

Available online at www.sciencedirect.com

SciVerse ScienceDirect



### **Elements of a dynamic systems model of canopy photosynthesis** Xin-Guang Zhu<sup>1,2</sup>, Qingfeng Song<sup>1,2</sup> and Donald R Ort<sup>3</sup>

Improving photosynthesis throughout the full canopy rather than photosynthesis of only the top leaves of the canopy is central to improving crop yields. Many canopy photosynthesis models have been developed from physiological and ecological perspectives, however most do not consider heterogeneities of microclimatic factors inside a canopy, canopy dynamics and associated energetics, or competition among different plants, and most models lack a direct linkage to molecular processes. Here we described the rationale, elements, and approaches necessary to build a dynamic systems model of canopy photosynthesis. A systems model should integrate metabolic processes including photosynthesis, respiration, nitrogen metabolism, resource remobilization and photosynthate partitioning with canopy level light, CO<sub>2</sub>, water vapor distributions and heat exchange processes. In so doing a systems-based canopy photosynthesis model will enable studies of molecular ecology and dramatically improve our insight into engineering crops for improved canopy photosynthetic CO<sub>2</sub> uptake, resource use efficiencies and yields.

#### Addresses

<sup>1</sup> State Key Laboratory of Hybrid Rice Research, CAS-MPG Partner Institute for Computational Biology, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Yueyang Road 320, Shanghai, 200031, China

<sup>2</sup> CAS Key Laboratory of Computational Biology, Chinese Academy of Sciences, CAS-MPG (Chinese Academy of Sciences-German Max Planck Society) Partner Institute for Computational Biology, Shanghai Institute of Biological Sciences, Chinese Academy of Sciences, Shanghai, 200031, China

<sup>3</sup> Global Change and Photosynthesis Research Unit, USDA ARS, Departments of Plant Biology and Crop Sciences, Institute of Genomic Biology, University of Illinois, Urbana, IL 61801, United States

Corresponding author: Zhu, Xin-Guang (zhuxinguang@picb.ac.cn)

#### Current Opinion in Plant Biology 2012, 15:237-244

This review comes from a themed issue on Physiology and metabolism Edited by Julian M Hibberd and Andreas PM Weber

Available online 9th February 2012

1369-5266/\$ - see front matter © 2012 Elsevier Ltd. All rights reserved.

DOI 10.1016/j.pbi.2012.01.010

### Introduction

On or near the 31st of October 2011, the global population reached 7 billion (http://www.worldometers.info/world-population/). Before plateauing in the middle of this century, the global population will have increased by another 2–4 billion humans [1], which along with improving

economic conditions in large parts of the developing world creates an unprecedented demand for food and energy [2]. An element of meeting the challenge of doubling the global food production will have to be increasing the efficiency of crop canopy photosynthesis [3<sup>•</sup>]. Crop yield is inherently related to the seasonal integral of net canopy photosynthesis. Canopy photosynthetic CO<sub>2</sub> uptake rate  $(A_c)$  is the integral of photosynthetic CO<sub>2</sub> uptake from both sunlit and shaded leaves. A<sub>c</sub> changes over the growing season as well as over the diurnal cycle. The difference in the light intensity dependence between canopy and leaf photosynthesis is shown in Figure 1. Photosynthesis of an individual leaf saturates at about 700  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> PPFD for a typical C3 and about 1000  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> PPFD for a C4 leaf. However, for a crop canopy containing several leaf layers, the A<sub>c</sub> for either C3 or C4 canopies shows little sign of saturation even at 2000  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> PPFD that is higher than full sunlight at most locations. Because of this fact, photosynthetic CO<sub>2</sub> uptake by shaded leaves can account for up to about 50% of  $A_{c}$  [4]. The historical misunderstanding of the relationship between photosynthesis and crop yields is largely owing to considering leaf instead of full canopy photosynthesis to derive the relationship [5,6].

Given that photosynthesis generates carbon and energy required for plant growth and development, photosynthesis is usually assumed to be optimized given that it has had >3.5 billion years of evolutionary fine tuning [7]. However, there is now a great deal of evidence to suggest that many photosynthetic components of crop plants, for example, Rubisco specificity factor [8], enzyme distributions [9], leaf chlorophyll concentration [10], leaf angle [11], and leaf nitrogen distribution [12<sup>•</sup>] are not optimized to achieve maximum  $A_{\rm c}$  (i.e. maximum net primary productivity). This lack of optimization to achieve maximum  $A_{\rm c}$  should not come as a surprise in that maximizing  $A_{\rm c}$  does not derive from selection for good competitors in natural ecosystems; maximizing  $A_c$  is a unique goal of agriculture. For example, an individual plant taller [13,14], greener [10], and having more horizontal leaves [15] than required for maximum  $A_c$  usually has a competitive advantage over its neighbors. Thus, plants adopt features beneficial for the individual at the cost of the community and therefore use resources, including light, inefficiently compared to strategies designed to maximize net primary productivity. Furthermore, selection in natural habitats favor individuals that invest resources to maintain sufficient defensive [16] or in preventative mechanisms to ensure their survival and competitive advantage when facing stress and other unpredictable environmental fluctuations [17,18<sup>•</sup>]. Additionally elements of global climate change, most notably increasing





The light response curves for both leaf (**a** and **b**) and canopy (**c** and **d**) photosynthesis in C3 (**a** and **c**) and C4 (**b** and **d**) plants. Simulations were conducted using WIMOVAC [44]. The canopy was assumed to have leaf area index of 4, random leaf distribution and inclination, solar zenith angle of  $30^{\circ}$  with 70% of incident light as direct light. Photosynthetic parameters for C3 plants were from [25] and for C4 plants were from [76]. PPFD: photosynthetic photon flux density.

atmospheric CO<sub>2</sub> concentration and temperature, are occurring at a pace that far exceeds the rate at which evolution of new adaptive features can occur. Indeed, the current Rubisco specificity factor in typical C3 plants is adapted to the CO<sub>2</sub> levels that existed for the ~20 M years that preceded the industrial revolution [8]. Finally, the boost of photosynthesis and biomass attendant to the insertion of a 'photorespiratory bypass' into plant chloroplasts using enzymes from *E. coli* [28] illustrates that there are more efficient alternatives for certain photosynthetic processes and enzymes that can be found in genetic resources to which plants do not have access without biotechnological intervention.

The deviation of photosynthesis from optimum and the difficulty of measuring canopy photosynthesis  $(A_c)$  in the field necessitate development of canopy photosynthesis models. Since the publication of the first mathematical model for estimating canopy photosynthesis [19], many models had been developed for agro-ecology or forest-ecology research for which there are several excellent reviews [20–22]. Our purpose here is to consider what is required to take the important next step of developing a full systems model of canopy photosynthesis. Such a model would enable studies of molecular ecology and provide insight to guide the engineering of crops for improved canopy photosynthetic  $CO_2$  uptake, crop yield and resource use efficiencies.

## Lessons learned from current canopy photosynthesis models

Among current canopy photosynthesis models the most well known are the: maximal productivity model, resource-use efficiency model, big-leaf model, sunlitshaded model, and multi-layer model. These models vary in the amount of physiological and environmental mechanism and detail that they incorporate [20]. Comparatively, the sunlit-shaded model is the best performer over a broad range of different time scales for which it has been validated, although other modeling approaches have specific advantages including simplicity of implementation and the ability to be combined with remote sensing data related to the interception of light by vegetation [20,23<sup>••</sup>,24<sup>•</sup>]. One major advantage of the sunlit-shaded model is its scalability and in particular its ability to directly incorporate leaf level physiological properties [23<sup>••</sup>]. Furthermore, important canopy architectural parameters, such as ratio of horizontal versus vertical projected leaf area ratios, leaf area index, and randomness of leaf angle distribution, are also represented in the model [24<sup>•</sup>], thereby providing it with the power to identify potential targets to engineer a higher  $A_c$ , for example, a decrease rather than increase in Rubisco specificity factor [8], decreasing leaf current chlorophyll concentration [10], and implementing a faster recovery from the photoprotective state to its full efficiency state [4]. Improving the light distribution inside a canopy through manipulating leaf angles is another approach to enhance  $A_c$  predicted by the sunlit-shaded model [11].

Current canopy photosynthesis models have also been used to study the interaction between canopy processes and environmental factors in both natural or managed ecosystems. For example, a sunlit-shaded canopy model has been used to predict that elevated  $CO_2$  can alter the magnitude or even the direction of responses of  $A_c$  to increase in temperature [25]. Changes in the fraction of diffuse light caused by clouds or other atmospheric particles were also forecast by this model to strongly influence  $A_{c}$  [26]. The influence of canopy height on  $A_{\rm c}$  can also be explored within current canopy models [27]. However, other important factors that impact canopy photosynthesis are not captured in current canopy photosynthesis models. For example, in cereal crops spike photosynthesis is known to play a significant role in determining crop yields [28], but this aspect has not been effectively represented in current canopy photosynthesis models for cereal crops.

## Elements of a dynamic systems model of canopy photosynthesis

The sunlit-shaded model, although clearly the most powerful of the existing canopy models, nevertheless has important constraints, which limit its accuracy in predicting canopy photosynthesis  $CO_2$  uptake rates  $(A_c)$  under numerous conditions of interest. Topping





The influence of light and CO<sub>2</sub> heterogeneities on the estimates of total C3 canopy CO<sub>2</sub> uptake rate ( $A_c$ ). (a) Hypothetical PPFD at 10 different depths within a canopy of leaves; (b) the average PPFD of the 10 depths; (c) estimated  $A_c$  using actual PPFD at the different depths; (d) estimated  $A_c$  with the average PPFD.  $C_a$ : atmospheric CO<sub>2</sub> concentration. The leaf photosynthetic rates were calculated based on light response curve following [3]. The parameters used are:  $A_{sat}$  of 24.2 µmol m<sup>-2</sup> s<sup>-1</sup> at [CO<sub>2</sub>] concentrations of 380 ppm and 21.4 µmol m<sup>-2</sup> s<sup>-1</sup> at 340 ppm. The convexity factors are 0.71 with and 0.95 without photoinhibition. The initial slopes of CO<sub>2</sub> uptake versus light curves are 0.028 with and 0.055 without photoinhibition. A simplification used in this calculation is the assumption of the same extent of photoinhibition for leaves at all 10 depths in the canopy.

the list of its limitations is that the sunlit-shaded model predicts the 'average' light intensities for both sunlit and shaded leaves within a canopy [23<sup>••</sup>,24<sup>•</sup>], that is, the high level of spatial and temporal heterogeneity of light environments inside a canopy is not considered. The influence of this heterogeneity on the calculation of  $A_{c}$ is illustrated in Figure 2. Assuming no photoinhibition (i.e. no photoprotection),  $A_{\rm c}$  calculated with the average light level is 17% higher than that estimated with the actual light levels inside a canopy (Figure 2). With photoinhibition (i.e. photoprotective mechanisms engaged), the estimated  $A_c$  using the average light level is 7% higher than the estimate with actual light level (Figure 2). Here we used the simplification that the level of photoinhibition was same for all leaves regardless of depth within the canopy; had we included the gradient in photoinhibition that would actually exist it would have resulted in an even greater difference in  $A_c$  calculated using average versus actual light levels. Furthermore, as with light levels,  $CO_2$  levels within a canopy are also highly heterogeneous both temporary and spatially, for example, a 40 ppm gradual decrease of CO<sub>2</sub> concentration from the top to the center of a canopy height is common in deep maize or soybean canopies [29,30]. Given that mesophyll resistance can further decrease CO<sub>2</sub> concentration in chloroplast stroma [31] and the non-linear response of photosynthesis to CO<sub>2</sub> [32], ignoring the CO<sub>2</sub> gradient inside a canopy overestimated  $A_c$  by ~4% for a C3 crop (Figure 2). Another limitation of current canopy photosynthesis models, as also discussed in [33], is their lack of linkage between leaf physiological processes to molecular processes. As a result, such models cannot be used to identify molecular targets to engineer for higher  $A_c$ .

Figure 3a illustrates the key elements required for a mechanistic systems model of canopy photosynthesis to overcome the above-mentioned limitations. A brief summary of the differences between current canopy photosynthesis models and a fully mechanistic dynamic systems model of canopy photosynthesis is shown in Table 1. The basic components of the dynamic systems model of canopy photosynthesis include physical models of canopy microclimate, systems models of metabolic processes representing photosynthesis, respiration, nitrogen metabolism, resource re-mobilization and photosynthate partitioning in a plant, and algorithms for integrating these components to develop the complete model. In the past few decades, tremendous progress has been made in each of these different aspects. The most notable of the major advances are summarized here. First, models representing 3-D canopy architectures have been developed for different crops including maize [34–36], rice [37,38], and wheat [39]. Various forward and reverse light ray tracing algorithms combined with such 3D canopy architecture models have been developed to predict the light environment inside a canopy [27,40-42]. Compared to the large number of light distribution models, the CO<sub>2</sub> distribution inside a canopy has only been modeled in a limited number of crops, for example, in soybean [43]. The temperature and humidity profiles are usually predicted by canopy models based on the energy balance and stomatal conductance [43,44].

Major advances have also been made in developing mechanistic models of plant metabolic processes. So far, a number of kinetic models of photosynthesis and of plant primary metabolism have been developed [9,45,46]. To enable simulation of canopy photosynthesis during the growing season, various models of photosynthate partitioning were also developed, including descriptive allometry models, functional equilibrium models, and sink regulation models with different levels of mechanistic underpinnings [47,48°,49]. Recently, a mechanistic model of starch metabolism and sugar partitioning between shoot and root was developed, where measurable parameters, including maximal leaf growth rates, minimal levels of sugar in the leaves, and relationship between the minimal





The major elements of a systems model of canopy photosynthesis (a) and algorithms used to integrate metabolic model with canopy microclimate model (b). The major elements include physical models of canopy microclimate, systems models of metabolic processes representing photosynthesis, respiration, nitrogen metabolism, resource re-mobilization and photosynthate partitioning, and algorithms for integrating these components to develop the complete model (a). A hierarchical multi-scale approach is used to integrate these different models to develop the complete systems model of canopy photosynthesis (b).

sugar levels and maintenance respiration rates, were used as key parameters  $[50^{\circ}]$ . This model can predict the growth retardation of both starch-less and starch excess mutants compared to wildtype  $[50^{\circ}]$ , which represent a major advance toward building a mechanistic source-sink model.

In addition to the photosynthate partitioning, plants remobilize carbon and nitrogen from senescencing leaves at the lowest layers of a canopy for re-use to grow new leaves at the top of the canopy, which usually happens when the photosynthetic  $CO_2$  uptake of shaded leaves is less than its respiratory cost [12°,51]. This remobilization is correlated with leaf longevity, leaf mass per area and density [51,52], with leaf age potentially being used to program nitrogen allocation to maximize carbon gain [53]. Recently, these different aspects, that is, photosynthesis, respiration, cost of protein turnover and other maintenance processes, have been incorporated into dynamic canopy photosynthesis models [54] and used to explain the response of trees to elevated  $CO_2$  [55,56]. Also a new robust method to estimate the energetic cost for building plant primary metabolism studies has been developed [57<sup>•</sup>], which is an important step toward a complete mechanistic systems model.

### Table 1

Comparison of current canopy photosynthesis models and emergent dynamic systems model of canopy photosynthesis. In the section labeled as 'Availability of features in different models', the ' $\times$ ' represents 'not available' and ' $\sqrt$ ' represents 'available'. In the section labeled as 'Applications of the model', the ' $\times$ ' represents 'incapable to do' and ' $\sqrt$ ' represents 'capable to do'

		Radiation use efficiency model [77]	Big leaf model [78–80]	Sunlit-shaded model [23••,24•]	Structural Function model [22,27,60]	Dynamic systems model
Availability of features in different models	Canopy architecture	×	×	×	$\checkmark$	$\checkmark$
	Leaf photosynthetic properties at different layers of the canopy	×	×	$\checkmark$	$\checkmark$	$\checkmark$
	Interaction between photosynthesis, respiration and nitrogen metabolism	×	×	×	×	$\checkmark$
	Source sink interaction	×	×	×	$\checkmark$	$\checkmark$
Applications of the model	Predict canopy photosynthetic CO <sub>2</sub> uptake	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
	Identify genes to modify for increased canopy photosynthesis	×	×	×	×	$\checkmark$
	Study mechanisms underlying plant adaptation to different environments	×	×	$\checkmark$	$\checkmark$	$\checkmark$

How is it possible to integrate a canopy microclimate model with these different metabolic models to develop a complete dynamic systems model of canopy photosynthesis? This is computationally challenging since the involved processes in the models occur at drastically different time scales spanning from picoseconds  $(10^{-9} \text{ s})$ for the primary events in photosynthesis to metabolic processes at millisecond scale  $(10^{-3} \text{ s})$  up to processes occurring at minutes scale ( $\sim 10^2$  s), for example, changes in light environments inside the canopy. To overcome this difficulty, we are adopting a hierarchical multi-scale approach [58] (Figure 3). Briefly, first a 'divide-and-conquer' strategy is used to develop individual metabolic models and these models will then be combined to form an integrated plant primary metabolism model [3<sup>•</sup>]. Then, we will run the complete metabolic model for a variety of parameter sets to generate an ensemble of photosynthetic rates under diverse parameter sets. This ensemble of data will then be used as a lookup table when we run the complete model of canopy photosynthesis (Figure 3b). Finally, the photosynthate produced will be used within the model to grow new organs with full consideration of both the construction cost and also the allometrical constraints observed in nature [58,59]. Recently, a novel aspect oriented functional structural plant modeling approach has also been proposed to deal with the complex interactions among different processes in plant growth and development [60].

Parameterization and validation of such a complex multiscale model is equally challenging. Fortunately, a number of critical measurement techniques required to parameterize and validate such models have also recently emerged. For example, a versatile stereo imaging system has been developed to obtain 3D canopy architectural parameters [61]. A number of physiological parameters, for example, stomatal conductance, biomass, biomass composition, and so on, have been measured with high throughput spectroscopic methods [62-65]. At the leaf level, activities of multiple key enzymes involved in plant primary metabolism are being measured with robot-based platforms [66]. Development of advanced canopy photosynthesis chambers to measure total photosynthetic  $CO_2$ uptake for model validation is also on-going. Combined these provide the means to generate a complete and robust data set required to parameterize, test and validate systems models of canopy photosynthesis.

# What a highly mechanistic systems model of canopy photosynthesis can offer in the post-genomic era

A complete systems model of canopy photosynthesis linking processes from the molecular level through to the canopy level will provide a powerful new tool to both test and generate hypotheses regarding mechanisms underlying plant growth and survival strategies in natural environments. A global scale survey of leaf economic traits showed that relationships between leaf mass per area, longevity, leaf nitrogen content and photosynthetic parameters, were remarkably conserved [67]. Furthermore, when facing stress, plants usually respond by taking one of three well-conserved strategies, that is, competitive, stress tolerant or ruderal, each with different combinations of canopy architecture, leaf form, longevity, and leaf turnover rates [68]. One intriguing hypothesis underlying these conserved relationships is that plants can potentially utilize many different strategies to grow and survive, and thus the particular strategy chosen might be an optimal solution of the dynamic systems model constrained by the growth environment and the genetic background of the plant. Though ecological and physiological models coupled with optimization routines have been developed to study relationships between these features [12,51,69,70], these models offer little opportunity to explore the molecular mechanisms behind these relationships. By linking molecular process to leaf and canopy processes, a dynamic systems model of canopy photosynthesis can offer new avenues to study ecological and physiological processes or phenomena from a molecular perspective.

A systems model of canopy photosynthesis can also be a major new tool to guide crop engineering to improve  $A_c$ . Although considerable effort has gone into engineering higher rates and more efficient photosynthesis, the success to date has been modest [71–74]. A dynamic systems model of photosynthesis as envisioned here, which incorporates the interactions among canopy microclimate, primary metabolism, resource re-mobilization in senescencing leaves, and associated energetic costs for maintaining existing tissues and constructing new tissues [57<sup>•</sup>,75<sup>•</sup>], would provide a robustframework to systematically re-evaluate the potential of different canopy features, for example, leaf nitrogen content, leaf thickness, leaf composition, enzyme distributions, enzyme kinetics and so on to improve  $A_c$  from a whole systems' energy and resource use efficiency perspective.

A persistent dream of plant biotechnology has been the ability to accurately predict the consequences of modifying different molecular, cellular, leaf and canopy level properties on total canopy  $CO_2$  uptake rates, on canopy light, water and nitrogen use efficiencies and ultimately on the yield of crops before ever conducting a field experiment. While this dream may never be entirely fulfilled, with the rapid advances in plant science, model building algorithms and computation capacity, it is timely now to develop a heuristic integrative model that will take a large step toward that goal.

### Acknowledgements

Funding for authors' research is from National Science Foundation of China (Grant No. 30970213), Ministry of Science and Technology of China (Grant No. 2011DFA31070), the Bill & Melinda Gates Foundation (Grant No. OPP1014417), and the Young Talent Frontier Program of Shanghai Institutes for Biology Sciences/Chinese Academy of Sciences (Grant No. 09Y1C11501), and a CAS visiting professorship grant to DRO and the United States Department of Agriculture, Agricultural Research Service.

### **References and recommended reading**

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
  of outstanding interest
- 1. Cohen JE: Human population: the next half century. Science 2003, **302**:1172-1175.
- 2. Tilman D, Balzer C, Hill J, Befort BL: Global food demand and the sustainable intensification of agriculture. *Proc Nat Acad Sci USA* 2011, **108**:20260-20264.
- 3. Zhu X-G, Long SP, Ort DR: Improving photosynthetic efficiency

• for greater yield. Annu Rev Plant Biol 2010, 61:235-261. This paper explores and evaluates numerous options to engineer higher photosynthetic efficiency for greater crop yields.

- Zhu X-G, Ort DR, Whitmarsh J, Long SP: The slow reversibility of photosystem II thermal energy dissipation on transfer from high to low light may cause large losses in carbon gain by crop canopies. A theoretical analysis. J Exp Bot 2004, 55:1167-1175.
- 5. Zelitch I: The close relationship between net photosynthesis
- and crop yield. Bioscience 1982, 32:796-802.

The controversy of the relationship or lack of relationship between photosynthesis and crop yields was discussed.

- Lawlor DW: Photosynthesis, productivity and environment. J Exp Bot 1995, 46:1449-1461.
- 7. Blankenship RE: Origin and early evolution of photosynthesis. *Photosynth Res* 1992, **33**:91-111.
- Zhu X-G, Portis AR Jr, Long SP: Would transformation of C3 crop plants with foreign Rubisco increase productivity? A computational analysis extrapolating from kinetic properties to canopy photosynthesis. *Plant Cell Environ* 2004, 27:155-165.
- Zhu X-G, de Sturler E, Long SP: Optimizing the distribution of resources between enzymes of carbon metabolism can dramatically increase photosynthetic rate: a numerical simulation using an evolutionary algorithm. *Plant Physiol* 2007, 145:513-526.
- 10. Ort DR, Zhu X, Melis A: **Optimizing antenna size to maximize** photosynthetic efficiency. *Plant Physiol* 2011, **155**:79-85.
- Long SP, Zhu XG, Naidu SL, Ort DR: Can improvement in photosynthesis increase crop yields? *Plant Cell Environ* 2006, 29:315-330.
- 12. Anten NPR, During HJ: Is analysing the nitrogen use at the plant
   canopy level a matter of choosing the right optimization criterion? *Oecologia* 2011, 167:293-303.

This paper discussed the need for considering both the dynamics and also competition among different individuals in canopy photosynthesis models.

- 13. Givnish TJ: On the adaptive significance of leaf height in forest herbs. Am Nat 1982, 120:353-381.
- 14. Falster DS, Westoby M: Plant height and evolutionary games. *Trends Ecol Evol* 2003, **18**:337-343.
- Hikosaka K, Hirose T: Leaf angle as a strategy for light competition: optimal and evolutionarily stable lightextinction coefficient within a leaf canopy. *Ecoscience* 1997, 4:501-507.
- 16. Endara MJ, Coley PD: The resource availability hypothesis revisited: a meta-analysis. *Funct Ecol* 2011, **25**:389-398.
- Murchie EH, Pinto M, Horton P: Agriculture and the new challenges for photosynthesis research. New Phytol 2008, 181:532-552.
- 18. Smith AM, Stitt M: Coordination of carbon supply and plant

• growth. Plant Cell Environ 2007, **30**:1126-1149. The long-term and short-term strategies that plants use to optimize starch synthesis and degradation for growth are discussed. Starch metabolism is receiving increasing attention as a major regulator of plant growth and development.

- Monsi M, Saeki T: Uber den Lichtfaktor in den Pflanzengesellschaf- u ur die Stoffproduktion. Jpn J Bot 1953, 14:22-52.
- 20. Medlyn B, Barrett D, Landsberg J, Sands P, Clement R: Conversion of canopy intercepted radiation to photosynthate: review of modelling approaches for regional scales. *Funct Plant Biol* 2003, **30**:153-169.
- 21. Hirose T: Development of the Monsi-Saeki theory on canopy structure and function. *Ann Bot* 2005, **95**:483-494.
- DeJong TM, Da Silva D, Vos J, Escobar-Gutierrez AJ: Using functional-structural plant models to study, understand and integrate plant development and ecophysiology. *Ann Bot* 2011, 108:987-989.
- 23. dePury DGG, Farquhar GD: Simple scaling of photosynthesis
   from leaves to canopies without the errors of big-leaf models. Plant Cell Environ 1997, 20:537-557.

This represents a major advance in canopy photosynthesis modeling by considering the division of leaves into sunlit categories and shaded categories. This model is widely used in current physiological and ecological research.

- 24. Norman JM: Interfacing leaf and canopy light interception
- models. In Predicting Photosynthesis for Ecosystem Models, vol 2. Edited by Hesketh JD, Jones JW. Boca Raton, Florida: CRC Press; 1980:49-67.
- See comments for ref [23\*\*].
- 25. Long SP: Modification of the response of photosynthetic productivity to rising temperature by atmospheric CO<sub>2</sub> concentrations: has its importance been underestimated? *Plant Cell Environ* 1991, **14**:729-739.
- 26. Roderick ML, Farquhar GD, Berry SL, Nobel IR: **On the direct** effect of clouds and atmospheric particles on the productivity and structure of vegetation. *Oecologia* 2001, **129**:21-30.
- Sarlikioti V, de Visser PHB, Buck-Sorlin GH, Marcelis LFM: How plant architecture affects light absorption and photosynthesis in tomato: towards an ideotype for plant architecture using a functional-structural plant model. Ann Bot 2011, 108:1065-1073.
- Hohmann-Marriott MF, Blankenship RE: Evolution of photosynthesis. Ann Rev Plant Biol 2011, 62:515-548.
- Buchmann N, Ehleringer JR: CO<sub>2</sub> concentration profiles, and carbon and oxygen isotopes in C3 and C4 crop canopies. Agric Forest Meteorol 1998, 89:45-58.
- 30. Prueger JH, Hatfield JL, Parkin TB, Krustas WP, Kaspar TC: Carbon dioxide dynamics during a growing season in midwestern cropping systems. *Environ Manage* 2004, **33**:S330-S343.
- Niinemets U, Diaz-Espejo A, Flexas J, Galmes J, Warren CR: Role of mesophyll diffusion conductance in constraining potential photosynthetic productivity in the field. J Exp Bot 2009, 60:2249-2270.
- Farquhar GD, von Caemmerer S, Berry JA: A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. *Planta* 1980, 149:78-90.
- 33. Yin X, Struik PC: Modelling the crop: from system dynamics to systems biology. *J Exp Bot* 2010, **61**:2171-2183.
- Zhang Y, Lao CL, Li BG, Chen Y, Guo Y, Wang XP, Ma YT, Zhao M: Monte Carlo simulation of solar radiation in maize canopies and its visualisation. N Z J Agric Res 2007, 50:553-558.
- 35. Guo Y, Ma YT, Zhan ZG, Li BG, Dingkuhn M, Luquet D, De Reffye P: **Parameter optimization and field validation of the functional-structural model GREENLAB for maize**. *Ann Bot* 2006, **97**:217-230.
- Espana ML, Baret F, Aries F, Chelle M, Andrieu B, Prevot L: Modeling maize canopy 3D architecture – application to reflectance simulation. *Ecol Model* 1999, 122:25-43.
- Watanabe T, Hanan JS, Room PM, Hasegawa T, Nakagawa H, Takahashi W: Rice morphogenesis and plant architecture: measurement, specification and the reconstruction of structural development by 3D architectural modelling. Ann Bot 2005, 95:1131-1143.

- Zheng BY, Shi LJ, Ma YT, Deng QY, Li BG, Guo Y: Comparison of architecture among different cultivars of hybrid rice using a spatial light model based on 3-D digitising. *Funct Plant Biol* 2008, 35:900-910.
- Evers JB, Vos J, Fournier C, Andrieu B, Chelle M, Struik PC: An architectural model of spring wheat: evaluation of the effects of population density and shading on model parameterization and performance. *Ecol Model* 2007, 200:308-320.
- Chelle M, Andrieu B: The nested radiosity model for the distribution of light within plant canopies. *Ecol Model* 1998, 111:75-91.
- 41. Pearcy RW: **Y** Plant, a three dimensional crown architecture model linking leaf physiology to crown performance. In 4th International Workshop on Fucntional Structural Plant Models. Edited by Godin C et al.: 7–11 June 2004, Montpellier, France: 2004:415.
- Pearcy RW, Yang WM: A three-dimensional crown architecture model for assessment of light capture and carbon gain by understory plants. *Oecologia* 1996, 108:1-12.
- Baldocchi D: A lagrangian random-walk model for simulating water vapor, CO<sub>2</sub> and sensible heat-flux densities and scalar profiles over and within a soybean canopy. *Boundary Layer Meteorology* 1992, 61:113-144.
- 44. Humphries SW, Long SP: WIMOVAC: a software package for modelling the dynamics of plant leaf and canopy photosynthesis. *Comput Appl Biosci* 1995, **11**:361-371.
- Laisk A, Eichelmann H, Oja V: C3 photosynthesis in silico. Photosynth Res 2006, 90:45-66.
- 46. Morgan JA, Rhodes D: Mathematical modeling of plant metabolic pathways. *Metab Eng* 2002, **4**:80-89.
- Pollock C, Farrar J, Tomos D, Gallagher J, Lu C, Koroleva O: Balancing supply and demand: the spatial regulation of carbon metabolism in grass and cereal leaves. *J Exp Bot* 2003, 54:489-494.
- 48. Minchin PEH, Thorpe MR, Farrar JF: A simple mechanistic model
  of phloem transport which explains sink priority. J Exp Bot 1993, 44:947-955.

This is a pioneering model providing a mechanistic explanation of sink capacity. The model is based on the hypothesis of bulk flow driven by an osmotically generated pressure gradient and unloading kinetics.

- Marcelis LFM, Heuvelink E, Goudriaan J: Modelling biomass production and yield of horticultural crops: a review. Sci Hortic 1998, 74:83-111.
- 50. Rasse DP, Tocquin P: Leaf carbohydrate controls over
   Arabidopsis growth and response to elevated CO<sub>2</sub>: an experimentally based model. New Phytol 2006, 172:500-513.

This work used measurable parameters and built a mechanistic model for carbon allocation, which is used to predict the differential growth patterns of mutants with alternations in starch metabolism. This model represents a significant step towards developing a mechanistic model of source sink interactions.

- 51. Hikosaka K: Leaf canopy as a dynamic system: ecophysiology and optimality in leaf turnover. Ann Bot 2005, 95:521-533.
- Poorter H, Niinemets Ü, Poorter L, Wright IJ, Villar R: Causes and consequences of variation in leaf mass per area (LMA): a meta-analysis. New Phytol 2009, 182:565-588.
- Field C: Allocating leaf nitrogen for the maximization of carbon gain: leaf age as a control on the allocation program. *Oecologia* 1983, 56:341-347.
- Franklin O, Ågren GI: Leaf senescence and resorption as mechanisms of maximizing photosynthetic production during canopy development at N limitation. *Funct Ecol* 2002, 16:727-733.
- 55. Franklin O: **Optimal nitrogen allocation controls tree responses** to elevated CO<sub>2</sub>. New Phytol 2007, **174**:811-822.
- 56. Franklin O, McMurtrie RE, Iversen CM, Crous KY, Finzi AC, Tissue DT, Ellsworth DS, Oren R, Norby R: Forest fine-root production and nitrogen use under elevated CO<sub>2</sub>: contrasting responses in evergreen and deciduous trees explained by a common principle. *Global Change Biol* 2009, **15**:132-144.

- 57. Piques M, Schulze WX, Hohne M, Usadel B, Gibon Y, Rohwer J,
- Stitt M: Ribosome and transcript copy numbers, polysome occupancy and enzyme dynamics in Arabidopsis. Mol Syst Biol 2009, 5:.

This work developed an approach to quantify the energetics associated with building proteins used in primary metabolism. Estimates of the energy needed for the protein synthesis required to establish metabolism is needed in order to calculate the energy associated with resource remobilization and new leaf formation.

- 58. Mäkelä A: Process-based modelling of tree and stand growth: towards a hierarchical treatment of multiscale processes. *Can J For Res* 2003, **33**:398-409.
- Niklas KJ: Plant allometry: is there a grand unifying theory? Biol Rev 2004, 79:871-889.
- Cieslak M, Seleznyova AN, Prusinkiewicz P, Hanan J: Towards aspect-oriented functional structural plant modelling. Ann Bot 2011, 108:1025-1041.
- Biskup B, Scharr H, Schurr U, Rascher UWE: A stereo imaging system for measuring structural parameters of plant canopies. *Plant Cell Environ* 2007, **30**:1299-1308.
- Munns R, James RA, Sirault XRR, Furbank RT, Jones HG: New phenotyping methods for screening wheat and barley for beneficial responses to water deficit. *J Exp Bot* 2010, 61:3499-3507.
- Metzner R, Thorpe MR, Breuer UWE, BlÜMler P, Schurr U, Schneider HU, Schroeder WH: Contrasting dynamics of water and mineral nutrients in stems shown by stable isotope tracers and cryo-SIMS. *Plant Cell Environ* 2010, 33:1393-1407.
- 64. Menzel MI, Tittmann S, BÜHler J, Preis S, Wolters N, Jahnke S, Walter A, Chlubek A, Leon A, Hermes N *et al.*: **Non-invasive determination of plant biomass with microwave resonators**. *Plant Cell Environ* 2009, **32**:368-379.
- Furbank RT, von Caemmerer S, Sheehy J, Edwards GE: C4 rice: a challenge for plant phenomics. Funct Plant Biol 2009, 36:845-856.
- 66. Gibon Y, Blaesing OE, Hannemann J, Carillo P, Hohne M, Hendriks JHM, Palacios N, Cross J, Selbig J, Stitt M: A robotbased platform to measure multiple enzyme activities in arabidopsis using a set of cycling assays: comparison of changes of enzyme activities and transcript levels during diurnal cycles and in prolonged darkness. *Plant Cell* 2004, 16:3304-3325.
- Wright IJ, Reich PB, Westoby M, Ackerly DD, Baruch Z, Bongers F, Cavender-Bares J, Chapin T, Cornelissen JHC, Diemer M *et al.*: The worldwide leaf economics spectrum. *Nature* 2004, 428:821-827.
- 68. Grime JP: Evidence for the existence of three primary strategies in plants and its relevance to ecological and evolutionary theory. *Am Nat* 1977, **111**:1169-1194.

- Hikosaka K: A model of dynamics of leaves and nitrogen in a plant canopy: an integration of canopy photosynthesis, leaf life span, and nitrogen use efficiency. *Am Nat* 2003, 162:149-164.
- McMurtrie RE, Norby RJ, Medlyn BE, Dewar RC, Pepper DA, Reich PB, Barton CVM: Why is plant-growth response to elevated CO<sub>2</sub> amplified when water is limiting, but reduced when nitrogen is limiting? A growth-optimisation hypothesis. *Funct Plant Biol* 2008, 35:521-534.
- Raines CA: Transgenic approaches to manipulate the environmental responses of the C<sub>3</sub> carbon fixation cycle. *Plant Cell Environ* 2006, 29:331-339.
- Feng L, Wang K, Li Y, Tan Y, Kong J, Li H, Li Y, Zhu Y: Overexpression of SBPase enhances photosynthesis against high temperature stress in transgenic rice plants. *Plant Cell Rep* 2007, 26:1635-1646.
- Lefebvre S, Lawson T, Zakhleniuk OV, Lloyd JC, Raines CA: Increased sedoheptulose-1,7-bisphosphatase activity in transgenic tobacco plants stimulates photosynthesis and growth from an early stage in development. *Plant Physiol* 2005, 138:451-460.
- Rosenthal D, Locke A, Khozaei M, Raines C, Long SP, Ort DR: Over-expressing the C3 photosynthesis cycle enzyme Sedoheptulose-1-7 Bisphosphatase improves photosynthetic carbon gain and yield under fully open air CO<sub>2</sub> fumigation (FACE). *BMC Plant Biol* 2011, 11:123.
- 75. Amthor JS: From sunlight to phytomass: on the potential
   efficiency of converting solar radiation to phyto-energy. New Phytol 2010, 188:939-959.

This work presents a nice summary of the energy required to build different components of a leaf.

- Collatz GJ, Ribas-Carbo M, Berry JA: Coupled photosynthesisstomatal conductance model for leaves of C4 plants. *Aust J Plant Physiol* 1992, 19:519-538.
- 77. Monteith JL: Climate and the efficiency of crop production in Britain. *Philos Trans R Soc London* 1977, **281**:277-294.
- Sellers PJ, Berry JA, Collatz GJ, Field CB, Hall FG: Canopy reflectance, photosynthesis, and transpiration. III. A reanalysis using improved leaf models and a new canopy integration scheme. *Remote Sens Environ* 1992, 42:187-216.
- 79. Amthor JS: Scaling CO<sub>2</sub> photosynthesis relationships from the leaf to the canopy. *Photosyn Res* 1994, **39**:321-350.
- Lloyd J, Farquhar GD: The CO<sub>2</sub> dependence of photosynthesis, plant growth responses to elevated atmospheric CO<sub>2</sub> concentrations and their interaction with soil nutrient status.
   1. General principles and forest ecosystems. *Funct Ecol* 1996, 10:4-32.