

Transient expression in *Nicotiana benthamiana* for rapid functional analysis of genes involved in non-photochemical quenching and carotenoid biosynthesis

Lauriebeth Leonelli¹, Erika Erickson^{1,2}, Dagmar Lyska^{1,2} and Krishna K. Niyogi^{1,2,*}

¹Howard Hughes Medical Institute, Department of Plant and Microbial Biology, University of California, Berkeley, CA 94720-3102, USA, and

²Molecular Biophysics and Integrated Bioimaging Division, Lawrence Berkeley National Laboratory, Berkeley, CA 94720, USA

Received 5 April 2016; revised 30 June 2016; accepted 4 July 2016; published online 15 September 2016.

*For correspondence (e-mail niyogi@berkeley.edu)

SUMMARY

Plants must switch rapidly between light harvesting and photoprotection in response to environmental fluctuations in light intensity. This switch can lead to losses in absorbed energy usage, as photoprotective energy dissipation mechanisms can take minutes to hours to fully relax. One possible way to improve photosynthesis is to engineer these energy dissipation mechanisms (measured as non-photochemical quenching of chlorophyll *a* fluorescence, NPQ) to induce and relax more quickly, resulting in smaller losses under dynamic light conditions. Previous studies aimed at understanding the enzymes involved in the regulation of NPQ have relied primarily on labor-intensive and time-consuming generation of stable transgenic lines and mutant populations – approaches limited to organisms amenable to genetic manipulation and mapping. To enable rapid functional testing of NPQ-related genes from diverse organisms, we performed *Agrobacterium tumefaciens*-mediated transient expression assays in *Nicotiana benthamiana* to test if NPQ kinetics could be modified in fully expanded leaves. By expressing *Arabidopsis thaliana* genes known to be involved in NPQ, we confirmed the viability of this method for studying dynamic photosynthetic processes. Subsequently, we used naturally occurring variation in photosystem II subunit S, a modulator of NPQ in plants, to explore how differences in amino acid sequence affect NPQ capacity and kinetics. Finally, we functionally characterized four predicted carotenoid biosynthesis genes from the marine algae *Nannochloropsis oceanica* and *Thalassiosira pseudonana* and examined the effect of their expression on NPQ in *N. benthamiana*. This method offers a powerful alternative to traditional gene characterization methods by providing a fast and easy platform for assessing gene function *in planta*.

Keywords: non-photochemical quenching, PSBS, carotenoid biosynthesis, xanthophyll cycle, transient assay, photosynthesis, *Nicotiana benthamiana*, *Nannochloropsis*, *Thalassiosira*, lutein epoxide.

INTRODUCTION

Conservative projections of population growth have indicated that by 2050 the world's population will exceed 9.2 billion (Raftery *et al.*, 2012). This population forecast portends an increase in crop demand for both food and industrial uses—a need which must be met by improving crop yields, as sources of arable land are limited (Wu *et al.*, 2014). This foreseeable challenge prompts the question: How can we engineer plants to be more productive?

With respect to theoretical maximum yields of sunlight to biomass conversion, plants use only one-third of the light energy available for photochemistry (Zhu *et al.*, 2010).

This inefficient energy usage is the combinatorial result of limited light penetrance into the canopy as well as photoprotective mechanisms that dissipate absorbed energy as heat during periods of light saturation (Melis, 2009; Endo *et al.*, 2014). The latter of these two factors is referred to as non-photochemical quenching of chlorophyll *a* fluorescence (NPQ). NPQ is vital for protecting plants during periods of high light stress, especially in environments with fluctuating light (Külheim *et al.*, 2002). However, sustained NPQ can result in wasteful dissipation of light energy in limiting light (Zhu *et al.*, 2004; Murchie and Niyogi, 2011),

and engineering of NPQ kinetics and capacity present attractive targets for improving photosynthetic yields (Zhu *et al.*, 2010; Long *et al.*, 2015).

Several different excitation quenching mechanisms contribute to what is collectively known as NPQ. These components include feedback de-excitation (qE), zeaxanthin-dependent quenching (qZ), photoinhibition (qI), and state transitions (qT), which are distinguished by the timescales on which they act (Müller *et al.*, 2001). The most rapidly responding quenching component, qE, is induced and relaxes over the course of seconds to minutes and requires zeaxanthin formation, a pH gradient and, in plants, the photosystem II subunit S (PSBS) protein (Demmig-Adams *et al.*, 1996; Niyogi *et al.*, 1998; Li *et al.*, 2000, 2002a). In *Arabidopsis*, zeaxanthin is a product of the xanthophyll cycle and its accumulation is regulated by the antagonistic activities of two enzymes, violaxanthin de-epoxidase (AtVDE) and zeaxanthin epoxidase (AtZEP) (Figure 2a). In excess light, a pH gradient builds up across the thylakoid membrane and the resulting low lumen pH activates AtVDE, which catalyzes the conversion of violaxanthin to zeaxanthin via the intermediate antheraxanthin (Yamamoto *et al.*, 2004). This acidification of the lumen also activates AtPSBS, which is thought to act as a pH sensor (via protonation of two conserved lumen-facing glutamate residues) that facilitates the induction of qE (Li *et al.*, 2002b, 2004). The exact role of AtPSBS remains unknown, but its presence is required for qE *in vivo*, and its importance is underscored by its existence in all photosynthetic land plants studied to date (Bonente *et al.*, 2008).

Mechanistic insights into NPQ have been obtained by studying mutants isolated from *Arabidopsis thaliana* and *Chlamydomonas reinhardtii*, however many other photosynthetic organisms independent of the green lineage perform NPQ (Goss and Lepetit, 2015). These organisms, like the diatom *Thalassiosira pseudonana* and the eustigmatophyte *Nannochloropsis oceanica*, contain a variety of different carotenoids and proteins involved in light harvesting and photoprotection (Brown, 1987; Coesel *et al.*, 2008; Vieler *et al.*, 2012). This rich biodiversity presents an opportunity to discover new mechanisms of NPQ, along with the genes involved. To this end, advancements in next-generation sequencing (NGS) have produced a burst of genome sequence information to aid with gene predictions. However, basic biological tools to study gene function in emerging model organisms are still in the developmental phase, stalling efforts to fully characterize these genes.

In an effort to circumvent this bottleneck and characterize potentially interesting genes involved in energy dissipation and carotenoid biosynthesis, we took advantage of a system that has been used extensively by the plant pathology community: transient *Agrobacterium tumefaciens*-mediated gene expression in *Nicotiana benthamiana*

(Rossi *et al.*, 1993; Kapila *et al.*, 1997; Van der Hoorn *et al.*, 2000; Armbruster *et al.*, 2016). This system exploits the natural ability of *A. tumefaciens* to genetically modify plant cells, enabling expression of candidate genes *in planta* in a matter of days. This system is particularly well suited for the study of algal genes involved in carotenoid biosynthesis and photosynthesis, as introduced genes can use endogenous pigments as substrates as well as existing photosynthetic machinery as a scaffold. In addition, the activity of genes expressed in the same leaf can be directly compared, because proteins accumulate in the same background.

In this study, we tested the viability of transient expression for modifying dynamic chloroplast processes by examining the effects of AtVDE, AtZEP, and AtPSBS overexpression on NPQ in fully expanded *N. benthamiana* leaves. We then used this method to directly compare the kinetics and quenching capacity resulting from expression of PSBS homologs from an alga, a bryophyte, and a vascular plant (*C. reinhardtii*, *Physcomitrella patens*, and *A. thaliana*, respectively) and showed that PSBS from *P. patens* confers the highest quenching capacity of those tested. Finally, we used the *N. benthamiana* platform to explore the function of four predicted carotenoid biosynthetic genes from emerging model organisms *T. pseudonana* and *N. oceanica*. Several of these proteins demonstrated predicted activity, whereas others, specifically the zeaxanthin epoxidase homolog from *N. oceanica* (NoZEP1), showed relaxed substrate specificity and formed lutein epoxide when expressed *in planta*.

RESULTS AND DISCUSSION

NPQ in *N. benthamiana* is altered by the transient expression of Arabidopsis genes

Because fully developed leaves contain mature chloroplasts and assembled photosystems, we tested whether the addition of Arabidopsis genes involved in photoprotection could alter photosynthesis in tobacco leaves at such a late stage of development. AtPSBS, AtVDE, and AtZEP are known to play vital roles in NPQ in Arabidopsis. We expressed these genes in fully expanded *N. benthamiana* leaves using *A. tumefaciens*-mediated delivery and observed effects on NPQ that were consistent with results observed in stable transgenics of Arabidopsis and other higher plants (Li *et al.*, 2002c; Chen and Gallie, 2012). NPQ capacity increased dramatically in leaf spots over-expressing AtPSBS (Figure 1a, b), similar to the effect of increased AtPSBS protein accumulation in Arabidopsis (Li *et al.*, 2002c). Leaf spots expressing AtVDE showed a more rapid NPQ induction than those expressing a control protein (GUS), whereas spots expressing AtZEP exhibited slower induction and never reached NPQ levels comparable to the GUS control (Figure 1a, b). AtZEP also appeared

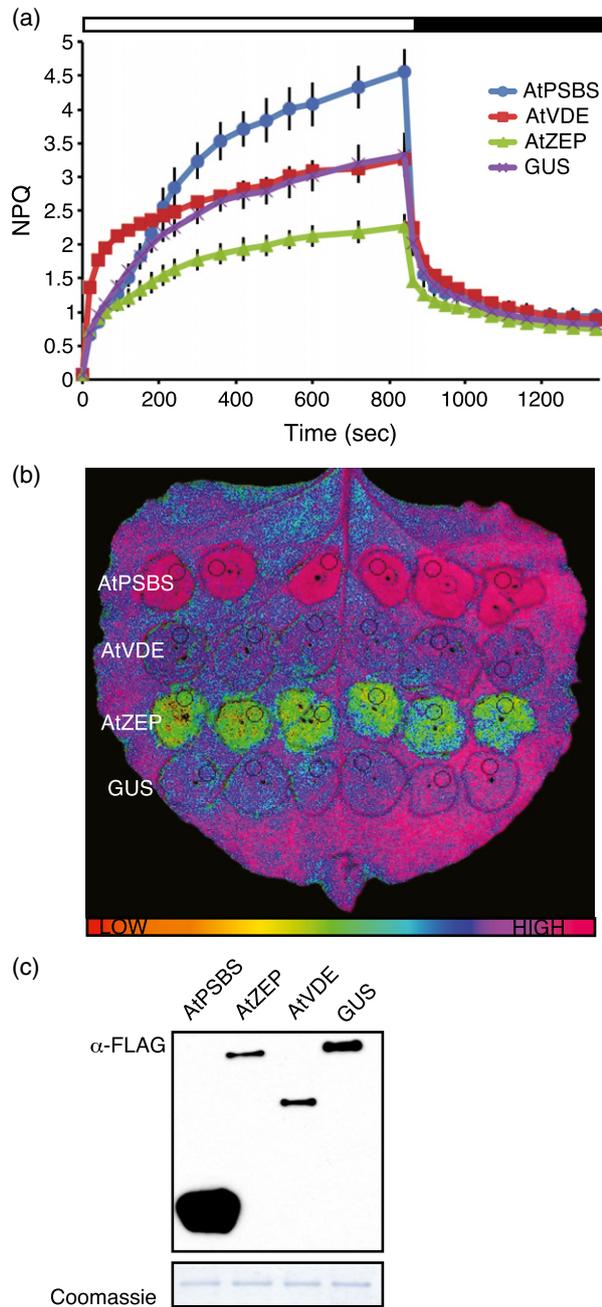


Figure 1. Over-expression of *Arabidopsis thaliana* NPQ-related genes in *Nicotiana benthamiana*.

(a) NPQ measurements corresponding to leaf spots over-expressing FLAG-tagged *AtPSBS*, *AtVDE*, or *AtZEP* with *GUS* as a negative control. White bar indicates $600 \mu\text{mol photons m}^{-2} \text{sec}^{-1}$ actinic light exposure, whereas black bar denotes the dark recovery period. Error bars represent standard deviation ($n = 6$).

(b) False-colored image of NPQ taken with IMAGING-PAM M-Series (Heinz Walz) of a leaf expressing *AtPSBS*, *AtVDE*, *AtZEP*, and *GUS* 10 min after high light exposure. Rainbow bar indicates relative amount of NPQ.

(c) Upper panel: Immunoblot analysis of tissue collected from panel (b) and probed with α -FLAG. Lower panel: Coomassie loading control.

to increase the speed of NPQ relaxation. All expressed proteins were tagged with a C-terminal FLAG tag (DYKDDDDK) and accumulated to detectable levels 48 h post infiltration (Figure 1c). These results suggest that by fine-tuning the expression of these proteins, NPQ kinetics can be engineered to better match the kinetics of light fluctuation.

To further investigate the NPQ phenotypes described above, we analyzed pigment composition in leaf spots before and after high light treatment. *AtVDE* and *AtZEP* act antagonistically, converting violaxanthin to zeaxanthin and back, respectively, through the intermediate antheraxanthin (Figure 2a). Zeaxanthin formation is involved in both the qE and qZ components of NPQ and is crucial for effective photoprotection (Niyogi *et al.*, 1998; Nilkens *et al.*, 2010). Compared to the *GUS* control, over-expression of *AtVDE* caused a marked decrease in violaxanthin accumulation and a concomitant increase in zeaxanthin even before high light treatment (Figure 2b, c). With zeaxanthin already present in these leaf spots, qE is poised for activation, explaining the rapid NPQ induction kinetics similar to that of *Arabidopsis npq2* plants that lack ZEP activity (Niyogi *et al.*, 1998). VDE activity depends on a conformational change that occurs when the pH of the thylakoid lumen decreases in saturating light conditions (Arnoux *et al.*, 2009). Thus, our finding that zeaxanthin accumulated under low light is surprising. Even after a 12-h dark acclimation period, zeaxanthin and antheraxanthin were still detectable in leaf spots over-expressing *AtVDE* (Figure S1). One possible explanation for VDE activity in the dark was recently suggested based on studies of *Ulva sp.* (Xie *et al.*, 2013). Using various electron transport chain (ETC.) and NPQ inhibitors, Xie *et al.* (2013) showed that VDE remained active in *Ulva sp.* in low light and implicated chlororespiration-mediated electron transport as a possible mechanism to generate sufficient lumenal acidification to activate VDE. Alternatively, an over-abundance of *AtVDE* protein crowded into the tight space of the thylakoid lumen may result in higher background activity due to an increased association of *AtVDE* with its membrane-bound substrate, facilitating zeaxanthin production even under low light conditions. In its inactive state, VDE is soluble in the thylakoid lumen, however, once activated it associates with the lipid MGDG and anchors to the membrane where it acts upon violaxanthin embedded in protein complexes to produce zeaxanthin (Hager and Holocher, 1994; Rockholm and Yamamoto, 1996).

Unlike VDE, ZEP is constitutively active. In leaf spots expressing *AtZEP*, zeaxanthin did not accumulate despite 15 min of high light treatment (Figure 2c). This absence of zeaxanthin explains the attenuated NPQ response to high light seen in Figure 1(a), similar to the phenotype of the *Arabidopsis npq1* mutant that lacks VDE activity and is

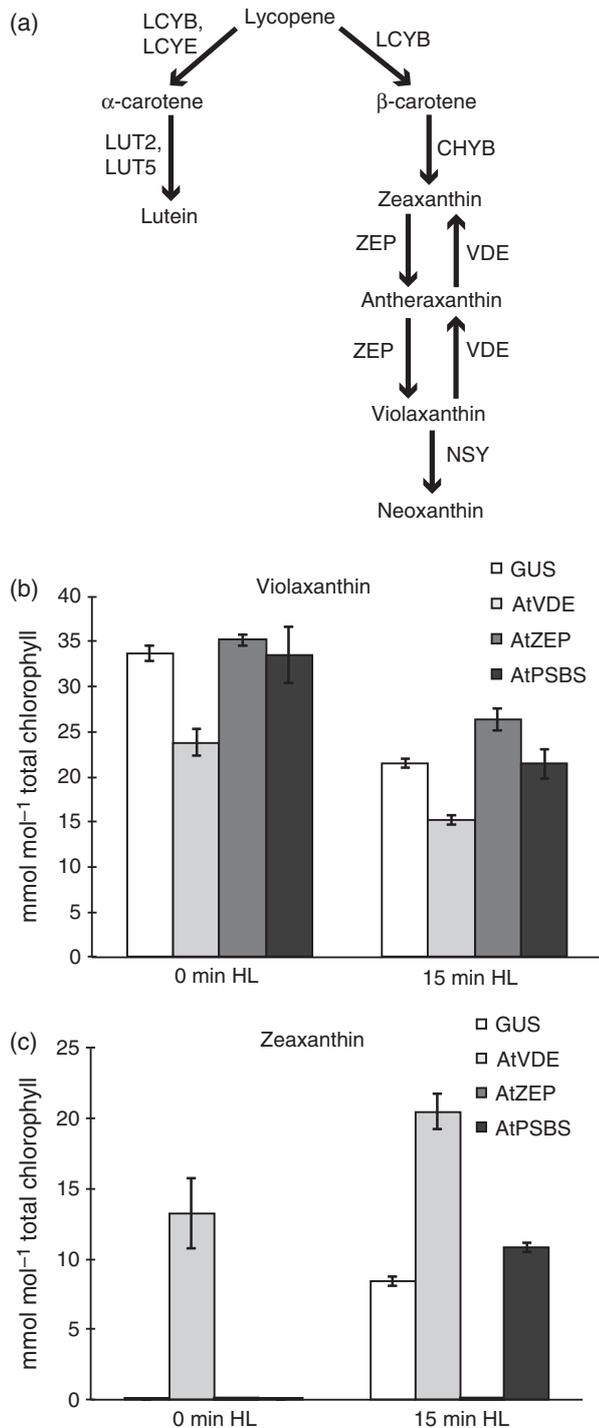


Figure 2. Xanthophyll accumulation in *N. benthamiana* leaf spots over-expressing *Arabidopsis thaliana* NPO-related genes. (a) Schematic diagram of a portion of the carotenoid biosynthetic pathway. Lycopene β -cyclase, LYCB; lycopene ϵ -cyclase, LYCE; β -carotene hydroxylase, CHYB; ϵ -ring hydroxylase, LUT2; violaxanthin de-epoxidase, VDE; zeaxanthin epoxidase, ZEP; neoxanthin synthase, NSY. (b, c) (b) Violaxanthin and (c) zeaxanthin accumulation in leaf spots over-expressing *AtPSBS*, *AtVDE*, or *AtZEP* with *GUS* as a negative control at time zero and after 15 min of high light treatment ($600 \mu\text{mol photons m}^{-2} \text{sec}^{-1}$). Error bars represent standard deviation ($n = 4$).

unable to accumulate zeaxanthin in high light (Niyogi *et al.*, 1998).

The de-epoxidation state (DES) of leaf spots expressing *AtVDE*, *AtZEP*, or *GUS* also reflected the trends observed in NPQ kinetics (Table 1). At time zero, leaf spots expressing *AtVDE* had a DES of 0.380, whereas the DES of *GUS* and *AtZEP* spots was 0. After 15 min of high light treatment, the DES of all spots increased, with *AtVDE*-expressing leaf spots still highest at 0.573, and the *GUS* control reaching a DES of only 0.319. Despite this difference in DES, both *AtVDE*- and *GUS*-expressing tissue achieved similar levels of NPQ, indicating that after reaching a certain threshold DES, the amplitude of NPQ is independent of DES.

These results demonstrate the plasticity of the photoprotective machinery in chloroplasts even at later stages of leaf development and indicate that this transient expression method can be used to study genes involved in photosynthesis and photoprotection. Moreover, this approach can, in principle, be used to examine gene function from any photosynthetic organism for which DNA sequence information is available, including species that are genetically intractable or otherwise difficult to work with, while also providing a consistent background for direct comparisons of gene function.

Distantly related *PSBS* genes show varying degrees of protonation-dependent quenching *in planta*

As demonstrated above, *PSBS* over-expression is an effective means of amplifying the qE component of energy dissipation. Although a lot of work has been done to characterize the role of this protein in NPQ (Brooks *et al.*, 2014), there is still much we can learn about *PSBS* by exploring natural variation between orthologs. *PSBS* genes were cloned from *Chlamydomonas reinhardtii* (*CrPSBS*), *Physcomitrella patens* (*PpPSBS*), and *Arabidopsis thaliana* (*AtPSBS*). All *PSBS* proteins contain two key lumen-exposed glutamate residues that become protonated in high light. These glutamates are crucial for *PSBS* activation, and when glutamine is substituted for each of these glutamates, the *PSBS* protein is rendered protonation-insensitive and does not activate when plants are exposed

Table 1 De-epoxidation states of leaf spots over-expressing various *Arabidopsis thaliana* genes involved in NPQ

	De-epoxidation state $(0.5A + Z)/(A + V + Z)$	
	Time 0	Time 15 min
GUS	0	0.319 ± 0.010
VDE	0.380 ± 0.049	0.573 ± 0.023
ZEP	0	0.113 ± 0.008
PSBS	0.013 ± 0.023	0.361 ± 0.013

to high light (Li *et al.*, 2002b). To compare the efficacy of PSBS-mediated NPQ amplification, PSBS genes from the species mentioned above were transiently expressed in *N. benthamiana*, along with protonation-insensitive mutant versions, and were scored for their ability to induce NPQ in response to high light. All three orthologs of PSBS enhanced qE (Figure 3a), including CrPSBS, which has not previously been shown to function *in vivo* (Bonente *et al.*, 2008). In each case, the enhanced qE was dependent on protonation of PSBS, as it required the conserved glutamate residues described in Li *et al.* (2004). PpPSBS conferred the highest levels of qE, which was somewhat surprising because *P. patens* uses both the algal-type LHCSR and the higher plant-type PSBS proteins to produce a robust high light response (Gerotto *et al.*, 2011). In fact, the majority of quenching in *P. patens* is attributed to LHCSR1 (Gerotto *et al.*, 2012), leading us to expect expression of PpPSBS alone in *N. benthamiana* to produce an intermediate level of quenching when compared to the higher plant version, AtPSBS. However, not only does PpPSBS confer higher qE than AtPSBS, it does so with a lower level of protein accumulation (Figure 3c).

Remarkably, CrPSBS also has the ability to confer quenching when expressed transiently in *N. benthamiana*, though to a lesser degree than either PpPSBS or AtPSBS. CrPSBS is one of the most strongly induced genes upon light exposure in *C. reinhardtii* (Duanmu *et al.*, 2013; Mettler *et al.*, 2014; Zones *et al.*, 2015), yet previous studies found no evidence that the protein accumulates or has any functionality in its native organism (Bonente *et al.*, 2008). Rather, *C. reinhardtii* relies on the algal-type LHCSR protein to facilitate qE (Peers *et al.*, 2009). Here we showed that when expressed in *N. benthamiana*, CrPSBS had protonation-dependent activity that enhanced the qE response relative to the inactive mutant forms of PSBS (Figure 3a). However, both active and protonation-insensitive forms of CrPSBS slightly reduced F_v/F_m values (Table 2) and showed a lag in Φ PSII recovery after high light treatment (Figure 3b). This reduction in PSII efficiency suggests a possible disruption to membrane organization when CrPSBS accumulates in higher plants regardless of qE activation. To ensure proper localization to the plant chloroplast, we replaced the first 44 amino acids of CrPSBS with the AtPSBS transit peptide (N-terminal 54 amino acid residues).

The three versions of PSBS presented in this study all demonstrated unique NPQ kinetics and capacities. These differences can be attributed to variations in amino acid sequence rather than protein levels, as PSBS accumulation did not necessarily correspond to NPQ capacity (e.g. despite greater amounts of CrPSBS, PpPSBS was better at facilitating quenching). Examining the differences between orthologs can provide insight into how PSBS can be engineered to activate more rapidly or generate a specific

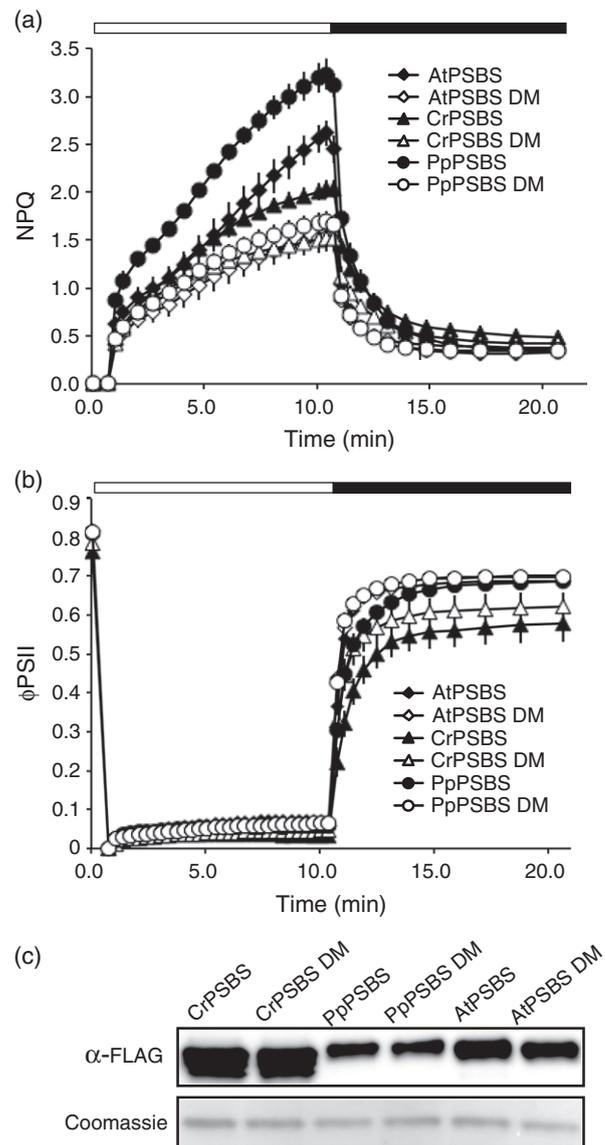


Figure 3. A comparison of PSBS activity from *A. thaliana*, *P. patens* and *C. reinhardtii*.

(a) NPQ of *N. benthamiana* leaf spots over-expressing FLAG-tagged PSBS from *A. thaliana* (AtPSBS), *P. patens* (PpPSBS) or *C. reinhardtii* (CrPSBS) when exposed to 10 min PAR 600 $\mu\text{mol photons m}^{-2} \text{sec}^{-1}$ (white bar) and 10 min dark recovery (black bar). DM denotes the protonation-insensitive double mutant form of each protein. Mutants are shown as open shapes.

(b) Φ PSII of leaf spots containing PSBS proteins. Measurements in (a) and (b) were obtained using the Dual-PAM-100 (Walz) and error bars in both (a) and (b) represent standard deviation ($n = 4$).

(c) Upper panel: Immunoblot probed with α -FLAG to determine PSBS protein accumulation. Lower panel: Coomassie loading control.

amount of quenching. Based on the sequence alignment of PSBS proteins from a broad range of species, many of the differences between alleles lie in or near the luminal loops that contain the glutamates believed to be protonated during activation (Figure S2). By exploring how changing these residues affects the pKa of activation and

Table 2 F_v/F_m values for leaf spots expressing various *PSBS* genes in *N. benthamiana*. *At*, *A. thaliana*; *Cr*, *C. reinhardtii*; *Pp*, *P. patens*. DM denotes a protonation-insensitive double mutant version of *PSBS*

	F_v/F_m
AtPSBS	0.81 ± 0.01
AtPSBS DM	0.81 ± 0.00
CrPSBS	0.76 ± 0.04
CrPSBS DM	0.78 ± 0.03
PpPSBS	0.81 ± 0.01
PpPSBS DM	0.81 ± 0.01

the intensity of quenching, we may learn more about *PSBS* function and ultimately discover how to tailor *PSBS* to produce an optimized NPQ response.

Chromalveolate carotenoid genes modify NPQ when expressed in planta

In addition to manipulating expression levels of known NPQ genes and comparing the efficiency of various alleles of *PSBS*, we used the *N. benthamiana*-*A. tumefaciens* system to investigate the function of putative carotenoid biosynthetic genes identified by homology searches from the marine algae *Nannochloropsis oceanica* and *Thalassiosira pseudonana*. Based on bioinformatic analysis, genes homologous to plant cytochrome P450-dependent β -carotene hydroxylase (*NoCYP97F5*) (Nelson, 2006), violaxanthin de-epoxidase (*NoVDE*, *TpDDE*), and zeaxanthin epoxidase (*NoZEP1*, *NoZEP2*) were cloned from *N. oceanica* and *T. pseudonana* and were expressed transiently in tobacco (Table S1). In certain cases, native transit peptides were replaced with that of AtPSBS (the first 56 amino acids) to ensure proper localization (Figure S3).

Functional annotations of genes from algae outside of the green lineage are often difficult to predict with confidence due to low sequence similarities. For instance, a BLAST search using the ZEP protein sequence from *Arabidopsis* identified two ZEP homologs from *N. oceanica*, *NoZEP1* and *NoZEP2*, both of which had less than 50% sequence identity to the original query (Table S1). Although both of these proteins accumulated when expressed in planta (Figure S4), only *NoZEP1* demonstrated epoxidase activity on available substrates (Figures 5e and 6c). While this result does not eliminate the possibility that *NoZEP2* has epoxidase activity, it does implicate *NoZEP1* as being the enzyme involved in the interconversion of violaxanthin, antheraxanthin, and zeaxanthin (the VAZ cycle) in *N. oceanica*. This case demonstrates not only the power of this method for validating gene function, but also provides us with an opportunity to explore how these genes from other organisms can affect NPQ in planta.

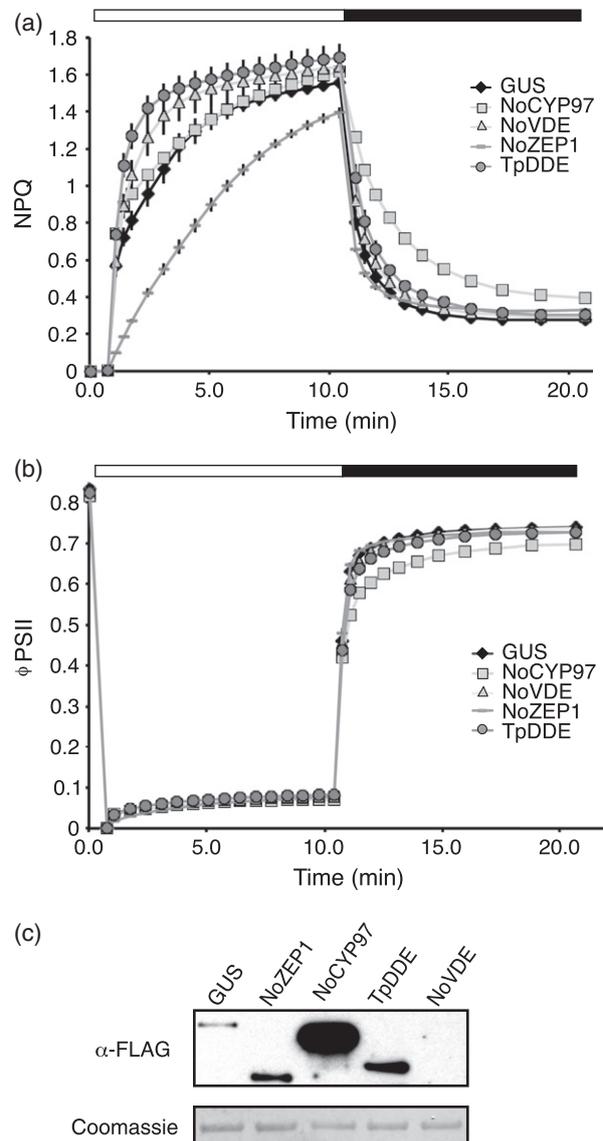


Figure 4. Transient expression of carotenoid biosynthetic genes from different algae in *N. benthamiana*.

(a) NPQ phenotypes associated with over-expression of genes predicted to be involved in carotenoid biosynthesis during 10 min high light exposure ($600 \mu\text{mol photons m}^{-2} \text{sec}^{-1}$, white bar) and 10 min dark recovery (black bar). *NoCYP97F5* is a cytochrome P450 from *N. oceanica* that shares sequence similarity with the cytochrome P450-type carotene hydroxylases from *A. thaliana*. *NoVDE* is the *N. oceanica* homolog of *AtVDE*, and *NoZEP1* is a homolog of *AtZEP*. *TpDDE* is an *AtVDE* homolog from *T. pseudonana*.

(b) Φ PSII of leaf spots expressing genes described above under aforementioned conditions. Measurements in (a) and (b) were obtained using the Dual-PAM-100 (Walz). Error bars in both (a) and (b) represent standard deviation ($n = 3$).

(c) Upper panel: Immunoblot probed with α -FLAG to determine algal protein accumulation. Lower panel: Coomassie loading control.

Remarkably, when expressed transiently in *N. benthamiana*, several of these distantly related algal genes altered the kinetics of plant-type NPQ (Figure 4a). Much like our finding that over-expression of *AtVDE* in *N. benthamiana*

caused faster induction of NPQ upon high light treatment, *NoVDE* and *TpDDE* expression showed similar trends relative to the *GUS* control, indicating an analogous function of these proteins. Similarly, over-expression of *NoZEP1* affected NPQ by dramatically slowing its induction kinetics, though unlike *AtZEP* over-expression, NPQ levels in leaf spots expressing *NoZEP1* eventually approached those of the *GUS* control after high light treatment. *NoCYP97F5* over-expression altered the overall shape of NPQ kinetics as well as severely impeding the speed of recovery in the dark. Additionally, leaf spots expressing *NoCYP97F5* appeared to suffer an accompanying reduction in Φ PSII recovery during the dark period compared to the control (Figure 4b). All proteins were C-terminally FLAG-tagged and, with the exception of *NoVDE*, accumulated to detectable levels after 48 h of expression (Figure 4c). Although *NoVDE* was not detected by immunoblotting, the NPQ induction phenotype leads us to believe that some protein must have accumulated, but was perhaps below detection limits or underwent some form of C-terminal processing.

To determine how each of the algal genes described above changes carotenoid levels, and in turn, to further understand how those changes underlie observed NPQ phenotypes, we examined pigment composition of leaf spots expressing these genes from both dark-acclimated and high-light-treated samples. In the case of leaf spots accumulating *NoVDE* or *TpDDE*, pigment profiles did not vary dramatically relative to the control in either dark-acclimated or high light treatments (Figure 5a–c). After pigment quantification, we found that *NoVDE* and *TpDDE* samples accumulated little to no zeaxanthin before high light treatment, but zeaxanthin levels were higher than that of the control after 10 min in high light (Figure 6a), as reflected by a higher DES (Table 3). These results suggest that both *NoVDE* and *TpDDE* are able to use violaxanthin as a substrate *in planta*, but they contrast with our findings that *AtVDE* over-expression resulted in appreciable amounts of zeaxanthin even before high light treatment, and nearly double the levels of the controls after treatment (Figure 2c). This disparity suggests that *NoVDE* and *TpDDE* might be regulated differently by lumen pH than *AtVDE*,

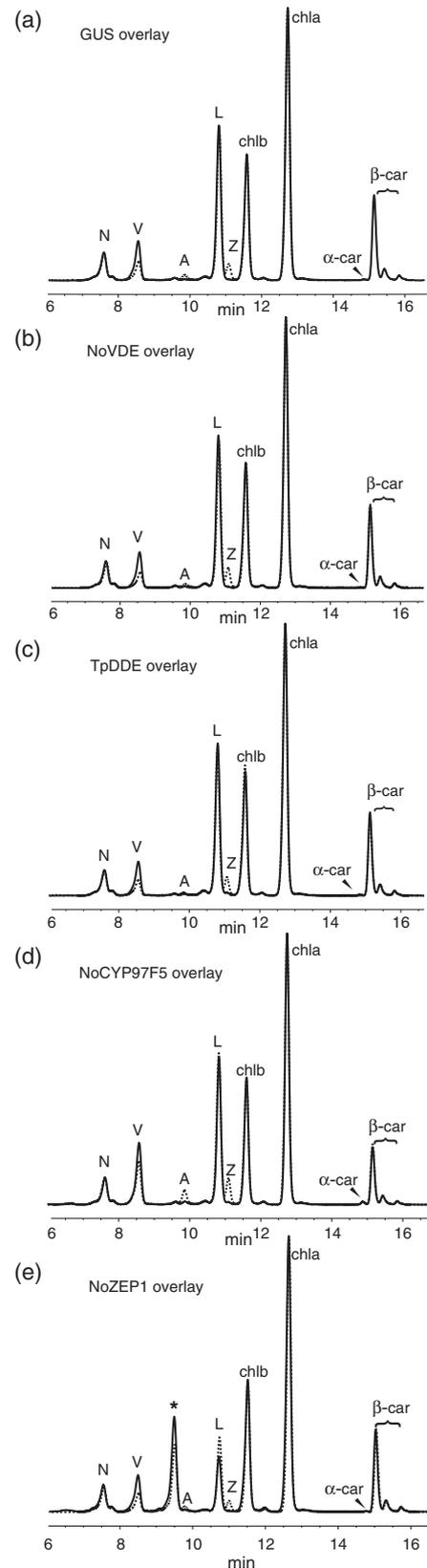


Figure 5. HPLC pigment profiles of leaf spots expressing predicted algal carotenoid biosynthesis genes in *N. benthamiana*. (a–e) (a) Pigments present in leaf spots expressing (a) *GUS*, (b) *NoVDE*, (c) *TpDDE*, (d) *NoCYP97F5*, and (e) *NoZEP1*. Solid black lines indicate pigment samples collected after a 20 min of dark acclimation, and dashed lines indicate pigment samples collected after 10 min high light exposure ($600 \mu\text{mol photons m}^{-2} \text{sec}^{-1}$). N, neoxanthin; V, violaxanthin; A, antheraxanthin; L, lutein; Z, zeaxanthin; chlb, chlorophyll *b*; chla, chlorophyll *a*; α -car, alpha-carotene; β -car, beta-carotene. Asterisk* in (e) denotes unknown peak later identified as lutein epoxide, Lx.

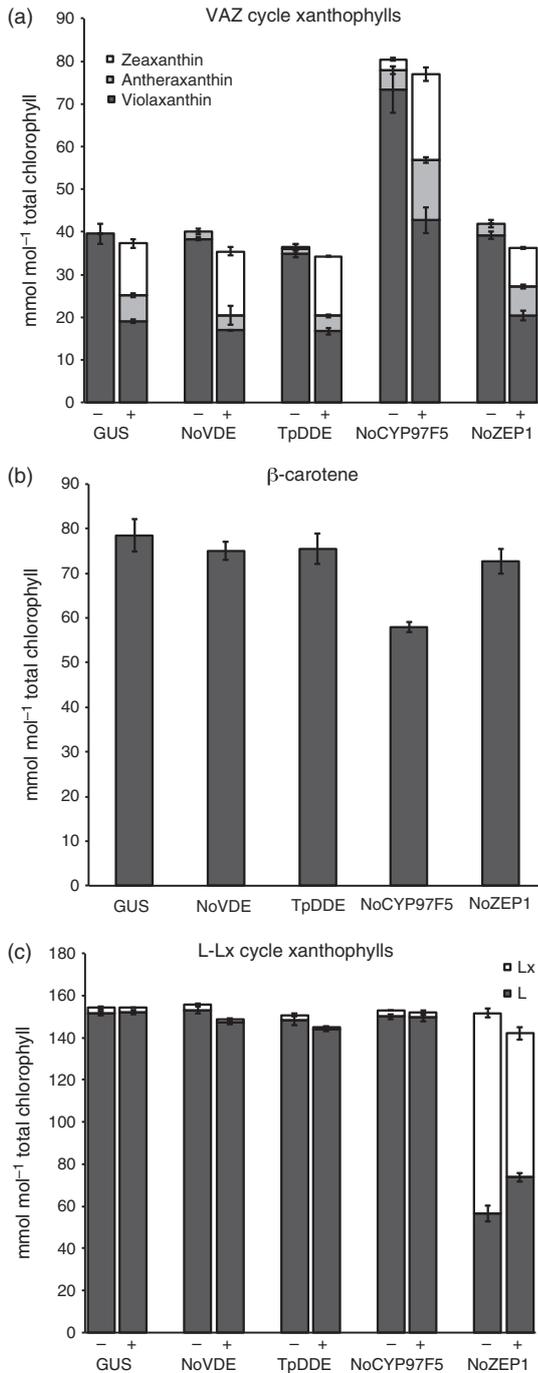


Figure 6. Pigment levels of *N. benthamiana* leaf tissue expressing various algal carotenoid biosynthesis genes.

(a) Levels of VAZ cycle xanthophylls in leaf discs expressing algal genes listed on the x-axis. '-' columns indicate samples collected before high light treatment, and '+' columns indicate samples collected after 10 min $600 \mu\text{mol photons m}^{-2} \text{sec}^{-1}$.

(b) β -Carotene levels in leaf discs expressing algal genes listed on the x-axis before high light treatment.

(c) Levels of L-Lx cycle xanthophylls in leaf discs expressing algal genes listed on the x-axis. '-' columns indicate samples collected before high light treatment, and '+' columns indicate samples collected after 10 min $600 \mu\text{mol photons m}^{-2} \text{sec}^{-1}$. L, lutein; Lx, lutein epoxide. All error bars represent standard deviation ($n = 4$).

Table 3 De-epoxidation states of leaf spots over-expressing various algal genes predicted to be involved in carotenoid biosynthesis before and after 10 min high light treatment. No, *N. oceanica*; Tp, *T. pseudonana*

	De-epoxidation state (0.5A + Z)/ (A + V + Z)	
	Time 0	Time 10 min
GUS	0	0.501 ± 0.017
NoVDE	0.009 ± 0.004	0.579 ± 0.015
TpDDE	0.024 ± 0.037	0.566 ± 0.013
NoZEP1	0.014 ± 0.004	0.420 ± 0.010
NoCYP97F5	0.059 ± 0.006	0.353 ± 0.019

although both can activate rapidly in light-saturating environments to speed the induction of NPQ (Figure 4a).

Transient expression of *NoCYP97F5* and *NoZEP1* led to major differences in pigment profiles when compared to the *GUS* control. Similar to studies of β -carotene hydroxylase over-expression in *A. thaliana* (Davison *et al.*, 2002), the β -xanthophyll pool in leaf spots accumulating *NoCYP97F5* was two-fold higher than in the control (Figure 6a). Consistent with the findings of Davison *et al.* (2002), the increase in β -xanthophyll accumulation did not affect accumulation of α -branch xanthophylls (Figure 6c). β -Carotene levels, however, were 25% lower than in *GUS* controls (Figure 6b). This reduction of β -carotene and the concomitant increase in violaxanthin is explained by the additional β -carotene hydroxylase activity conferred by *NoCYP97F5* expression. It is notable that *NoCYP97F5* is the only predicted β -carotene hydroxylase in *N. oceanica*, which lacks a homolog of the di-iron type of β -carotene hydroxylase (Vieler *et al.*, 2012), yet it is capable of working alongside the native di-iron and CYP97C hydroxylases in *N. benthamiana*. Violaxanthin levels were twice as high in these samples, leading to twice as much zeaxanthin production upon high light treatment. However, the increase in zeaxanthin formation did not translate into enhanced NPQ (Figure 4a) as previously observed in stable *A. thaliana* lines expressing *AtCHYB* (Johnson *et al.*, 2008). In contrast to this earlier study that showed NPQ levels depend on the DES of the xanthophyll cycle pool, we found that DES was not an accurate predictor of NPQ in *N. benthamiana* leaf spots expressing *NoCYP97F5*. Despite achieving similar levels of NPQ after 10 min in high light, the DES of *NoCYP97F5* samples was 0.353, whereas *GUS* controls had a DES of 0.501 (Table 3). The NPQ relaxation was delayed in *NoCYP97F5* samples, as was the recovery of ΦPSII (Figure 4a, b). One possible explanation for the delay in NPQ relaxation is an increase in the contribution of qZ, a different zeaxanthin-dependent component of NPQ that relaxes on a longer timescale (Nilkens *et al.*, 2010) and would depend on endogenous NbZEP activity.

Most strikingly, pigment profiles of leaf spots expressing *NoZEP1* showed a marked decrease in lutein, accumulating less than half that of GUS controls (Figure 6c), and contained a major peak (denoted by an asterisk) which is absent in all other samples (Figure 5e). Because *NoZEP1* was predicted to have epoxidase activity and showed a severe reduction in lutein accumulation, we hypothesized that the new peak in these samples was likely lutein epoxide (Lx). To test this hypothesis, we analyzed the pigment composition of leaf tissue from *Cucumis sativus*, a plant known to contain Lx (Esteban *et al.*, 2009), and compared the retention time and absorbance spectrum of the known Lx reference against the unknown peak. Comparison of pigment profiles between *C. sativus* and *NoZEP1*-expressing leaf spots showed that the Lx peak from cucumber clearly overlapped with the unknown peak present in *NoZEP1* samples (Figure S5a). When absorbance spectra of these peaks were measured, the absorbance maxima were almost identical (Figure S5b, c), further supporting our hypothesis that the peak in question was Lx. Moreover, *NoZEP1* samples showed a 20% decrease in this pigment along with a simultaneous and stoichiometric increase in lutein after 10 min in high light (Figure 6c). This inter-conversion was likely due to the de-epoxidase activity of endogenous NbVDE, which also converted violaxanthin to zeaxanthin (Figure 6a). This pattern of pigment accumulation is consistent with the NPQ kinetics we observed. A rapid initial rise in NPQ has been shown to depend on the presence of lutein (Pogson *et al.*, 1998). Prior to high light exposure, *NoZEP1* samples had 60% of their lutein sequestered as lutein epoxide (Figure 6c), simulating lutein depletion and preventing a rapid induction of qE. During high light, Lx was converted to lutein and zeaxanthin accumulated, activating qE to allow leaf spots expressing *NoZEP1* to approach NPQ levels of controls (Figure 4a).

Nannochloropsis lacks α -carotenoids and consequently does not contain lutein or its derivatives (Brown, 1987; Vieler *et al.*, 2012). Nevertheless, the β -carotene hydroxylase activity of NoCYP97C has retained its substrate specificity as evidenced by the unchanged levels of lutein, derived from α -carotene (Figure 6c). In contrast, *NoZEP1* is apparently unable to distinguish between β rings of α - and β -carotenoids because lutein and zeaxanthin are both epoxidized when *NoZEP1* is expressed *in planta*. This example illustrates the two-fold benefit of screening algal carotenoid genes in this heterologous system: not only can gene function be assigned based on protein activity *in planta*, but the natural variation in substrate specificity of these enzymes can be exploited to create new pigment profiles. The resulting shifts in pigment composition can then be examined for their effect on photosynthesis and NPQ in an effort to discover new ways to optimize these pathways.

CONCLUSIONS

In this study, we show that transient expression in *N. benthamiana* is an effective tool for manipulating dynamic photosynthesis-related processes *in vivo*. Using this approach, we were able to manipulate NPQ kinetics in a predictable manner by over-expression of known genes involved in NPQ induction and relaxation. Additionally, we examined the efficacy of three distantly related PSBS proteins from an alga, a bryophyte, and an angiosperm, and found that all three proteins confer additional protonation-dependent NPQ when over-expressed in *N. benthamiana*. While PSBS from *P. patens* showed the fastest induction and greatest increase in quenching capacity, perhaps the most striking observation was the ability of CrPSBS to contribute to qE *in planta*, suggesting that it might also have a qE-related function in *C. reinhardtii*.

Beyond investigation of known contributors to NPQ, this method also allows for rapid identification of new components in carotenoid biosynthesis and energy dissipation from emerging model organisms. By expressing the algal genes described above, we were able to confirm predicted functions for NoVDE, TpDDE, NoCYP97F5, and *NoZEP1*. This study serves as proof of principle that photosynthetic genes from distantly related organisms can be assayed in this heterologous system. Furthermore, these new versions of carotenoid biosynthetic genes provide us with kinetically distinct alternatives to higher plant genes and present additional opportunities to manipulate NPQ in plants. Finally, expression of some of these algal genes in *N. benthamiana* can produce unexpected pigments that could influence photo-protection or light harvesting. For instance, *NoZEP1* over-expression produced lutein epoxide, a pigment that has been shown *in vitro* to increase energy transfer efficiency between chlorophyll *a* molecules in a reconstituted antenna protein (Matsubara *et al.*, 2007).

EXPERIMENTAL PROCEDURES

Gene sources

RNA from *A. thaliana* Columbia, *N. oceanica* CCMP1779, and *T. pseudonana* CCMP1335 was isolated using the TRIzol reagent (Life Technologies, Carlsbad, CA, USA) as per the manufacturer's instructions. cDNA was generated from these organisms with the Omniscript RT kit (Qiagen, Valencia, CA, USA), and genes of interest were amplified using the primers listed in Table S2.

Plant growth/material

N. benthamiana seeds were planted in Sunshine soil mix 4 and grown in 12 h day/night cycles in a Conviron growth chamber with light intensity ranging from 60 to 80 $\mu\text{mol photons m}^{-2} \text{sec}^{-1}$. Plants used in this study were between 4 and 6 weeks old.

Plasmid construction

AttB adapter sites were added to genes of interest by polymerase chain reaction (PCR) and recombined into gateway-compatible pDONR entry vectors using BP clonase II (Life Technologies). These pDONR vectors were used to recombine genes of interest into the pEARLEYGATE 100 destination vector using LR clonase II (Life Technologies).

Site-directed mutagenesis

Point mutations were introduced into PSBS using either the Quik-Change Lightning Multi site-directed mutagenesis kit (Agilent Technologies, Santa Clara, CA, USA) or the 3SM method (Follo and Isidoro, 2008). The PSBS transit peptide was added to algal genes using the splicing overlap extension PCR method (Warrens *et al.*, 1997). Primers used are listed in Table S2.

Agrobacterium tumefaciens transformation

A. tumefaciens strain GV3101 was transformed with 400 ng of each plasmid by snap freezing in liquid nitrogen for 2 min, followed by heat shock at 37°C for 5 min. Transformants were rescued in 200 µl LB with shaking at 28°C for 2–3 h and plated onto LB agar containing 30 µg/µl kanamycin and 50 µg/µl gentamycin. Single colonies appeared after 2 days of growth at 28°C.

Transient expression in *Nicotiana benthamiana*

A. tumefaciens strains were resuspended in 1 ml induction medium (10 mM MgCl₂, 10 mM MES pH5.6, 150 µM acetosyringone) and incubated for 2 h at 28°C. Cultures were then diluted to an A₆₀₀ of 0.5 and injected into *N. benthamiana* leaves using a blunt-end 1 ml syringe. Plants were placed under constant light (~70 µmol photons m⁻² sec⁻¹) for 48–60 h before light measurements, protein samples, and high pressure liquid chromatography (HPLC) samples were collected. In addition to technical replicates, at least three biological replicates were collected for light treatments and pigment samples.

Chlorophyll fluorescence measurements

Video imaging was performed on detached *N. benthamiana* leaves with the IMAGING-PAM M-Series (Heinz Walz, Effeltrich, Germany) using 15 min of blue actinic light (600 µmol photons m⁻² sec⁻¹), followed by 10 min of dark recovery. Unless otherwise stated, samples were dark-acclimated for 20 min prior to measurements. Photosynthetic parameters were calculated as described (Brooks and Niyogi, 2011).

The Dual-PAM-100 (Heinz Walz) was used to measure chlorophyll *a* fluorescence of *N. benthamiana* leaf discs expressing various constructs involved in NPQ and carotenoid biosynthesis. Unless otherwise indicated, samples were dark-acclimated for 20 min prior to all treatments. Samples were treated with 10 min of red actinic light (660 µmol photons m⁻² sec⁻¹) followed by 10 min of dark recovery.

Immunoblot analysis

Leaf discs (0.5 cm²) expressing different constructs were collected and frozen in liquid nitrogen. Samples were powdered (6.5 m/sec for 60 sec) using the cryorotor of a Fastprep-24 (MP Bio) and resuspended in 25 µl of 3× SDS buffer. Samples were heated to 85°C for 3 min and then centrifuged at 14 000 rpm for 3 min to pellet cell debris. 1 µl of each sample was diluted into 6 µl of 1× SDS buffer

and loaded onto a pre-cast 10–20% Tris–glycine gel (Invitrogen) and run at 125 V for 2 h at 4°C. Proteins were transferred onto 0.45 µm PVDF (GE Healthcare, Pittsburg, PA, USA) using the Semi-Phor semi-dry transfer system (Hoefer Inc, Holliston, MA, USA), and membranes were blocked for 30 min at room temperature in 5% milk in TBST. Membranes were probed overnight at 4°C with rabbit anti-FLAG (1:5000) primary antibody (Sigma) in TBST. Secondary antibody was donkey anti-rabbit-HRP (Amersham/GE Healthcare, Pittsburg, PA, USA) used at 1:25 000.

HPLC analysis of pigments

Leaf discs (0.5 cm²) expressing different constructs were frozen in liquid nitrogen and powdered as described above; 100 µl acetone was added to each sample and gently vortexed for 15 sec. Cell debris was pelleted at 14 000 rpm for 3 min and the supernatant was collected. Samples were extracted a second time as described above, and supernatants were pooled to total 200 µl. Samples were analyzed with a Spherisorb 5 µm ODS1 column (Waters Corp, Milford, MA, USA) as previously described (Müller-Moulé *et al.*, 2002).

ACCESSION NUMBERS

AtPSBS: NP_175092; AtVDE: NP_172331; AtZEP: NP_851285; CrPSBS: XP_001689476; PpPSBS: XP_001778563; TpDDE: XP_002292080; NoZEP1: KU980906; NoZEP2: KU980907; NoVDE: KU980905; NoCYP97F5: KU980908.

ACKNOWLEDGEMENTS

We thank Tom Kleist for his contribution of *P. patens* cDNA, Thuy Truong for providing *C. reinhardtii* PSBS cDNA, David Nelson for assistance in cytochrome P450 nomenclature, Matthew Brooks for helpful discussions and technical support, Marilyn Kobayashi for general awesomeness, and other members of the Niyogi lab for their support. This work was supported by the Bill & Melinda Gates Foundation RIPE Project under a subaward from the University of Illinois. The work on expression of *A. thaliana* genes was funded by the Advanced Research Projects Agency-Energy (ARPA-E), U.S. Department of Energy, under Award Number DE-AR0000204. The work on expression of PSBS was supported by the U.S. Department of Energy, Office of Science, Basic Energy Sciences, Chemical Sciences, Geosciences, and Biosciences Division under field work proposal 449B. K.K.N. is an investigator of the Howard Hughes Medical Institute and the Gordon and Betty Moore Foundation (through Grant GBMF3070). The authors have no conflicts of interest to declare.

SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article.

Figure S1. Xanthophyll levels after 12 h of dark acclimation in leaf spots transiently expressing *AtVDE* or *GUS*.

Figure S2. A MUSCLE alignment of mature PSBS proteins from a broad range of species.

Figure S3. Protein sequences of algal genes isolated and used in this study.

Figure S4. Immunoblot showing protein accumulation of transiently expressed *NoZEP1* and *NoZEP2* in *N. benthamiana* leaf discs.

Figure S5. Identification of unknown peak present in *NoZEP1*-expressing *N. benthamiana* leaves.

Table S1. List of proteins and corresponding percent sequence similarity used to probe *N. oceanica* and *T. pseudonana* genomes for carotenoid biosynthetic genes.

Table S2. A list of primers used in this study.

REFERENCES

- Armbruster, U., Leonelli, L., Correa Galvis, V., Strand, D., Quinn, E.H., Jonikas, M.C. and Niyogi, K.K. (2016) Regulation and levels of the thylakoid K⁺/H⁺ antiporter KEA3 shape the dynamic response of photosynthesis in fluctuating light. *Plant Cell Physiol.* **57**, 1557–1567.
- Arnoux, P., Morosinotto, T., Saga, G., Bassi, R. and Pignol, D. (2009) A structural basis for the pH-dependent xanthophyll cycle in Arabidopsis thaliana. *Plant Cell*, **21**, 2036–2044.
- Bonente, G., Passarini, F., Cazzaniga, S., Mancone, C., Buia, M.C., Tripodi, M., Bassi, R. and Caffarri, S. (2008) The occurrence of the *psbS* gene product in *Chlamydomonas reinhardtii* and in other photosynthetic organisms and its correlation with energy quenching. *Photochem. Photobiol.* **84**, 1359–1370.
- Brooks, M.D. and Niyogi, K.K. (2011) Use of a pulse-amplitude modulated chlorophyll fluorometer to study the efficiency of photosynthesis in Arabidopsis plants. *Methods Mol. Biol.* **775**, 299–310.
- Brooks, M., Jansson, S. and Niyogi, K. (2014) PsbS-dependent non-photochemical quenching. In *Non-Photochemical Quenching and Energy Dissipation in Plants, Algae and Cyanobacteria* (Demmig-Adams, B., Garab, G., Adams III, W. and Govindjee, eds). Netherlands: Springer, pp. 297–314.
- Brown, J.S. (1987) Functional organization of chlorophyll a and carotenoids in the alga, *Nannochloropsis salina*. *Plant Physiol.* **83**, 434–437.
- Chen, Z. and Gallie, D.R. (2012) Violaxanthin de-epoxidase is rate-limiting for non-photochemical quenching under subsaturating light or during chilling in Arabidopsis. *Plant Physiol. Biochem.* **58**, 66–82.
- Coesel, S., Obornik, M., Varela, J., Falcitore, A. and Bowler, C. (2008) Evolutionary origins and functions of the carotenoid biosynthetic pathway in marine diatoms. *PLoS ONE*, **3**, e2896.
- Davison, P.A., Hunter, C.N. and Horton, P. (2002) Overexpression of [beta]-carotene hydroxylase enhances stress tolerance in Arabidopsis. *Nature*, **418**, 203–206.
- Demmig-Adams, B., Gilmore, A.M. and Adams, W.W. (1996) Carotenoids 3: *in vivo* function of carotenoids in higher plants. *FASEB J.* **10**, 403–412.
- Duanmu, D., Casero, D., Dent, R.M. et al. (2013) Retrograde bilin signaling enables *Chlamydomonas* greening and phototrophic survival. *Proc. Natl Acad. Sci. USA*, **110**, 3621–3626.
- Endo, T., Uebayashi, N., Ishida, S., Ikeuchi, M. and Sato, F. (2014) Light energy allocation at PSII under field light conditions: how much energy is lost in NPQ-associated dissipation? *Plant Physiol. Biochem.* **81**, 115–120.
- Esteban, R., Olano, J.M., Castresana, J., Fernández-Marín, B., Hernández, A., Becerril, J.M. and García-Plazaola, J.I. (2009) Distribution and evolutionary trends of photoprotective isoprenoids (xanthophylls and tocopherols) within the plant kingdom. *Physiol. Plant.* **135**, 379–389.
- Follo, C. and Isidoro, C. (2008) A fast and simple method for simultaneous mixed site-specific mutagenesis of a wide coding sequence. *Biotechnol. Appl. Biochem.* **49**, 175–183.
- Gerotto, C., Alboresi, A., Giacometti, G.M., Bassi, R. and Morosinotto, T. (2011) Role of PSBS and LHCSR in *Physcomitrella patens* acclimation to high light and low temperature. *Plant, Cell Environ.* **34**, 922–932.
- Gerotto, C., Alboresi, A., Giacometti, G.M., Bassi, R. and Morosinotto, T. (2012) Coexistence of plant and algal energy dissipation mechanisms in the moss *Physcomitrella patens*. *New Phytol.* **196**, 763–773.
- Goss, R. and Lepetit, B. (2015) Biodiversity of NPQ. *J. Plant Physiol.* **172**, 13–32.
- Hager, A. and Holoche, K. (1994) Localization of the xanthophyll-cycle enzyme violaxanthin de-epoxidase within the thylakoid lumen and abolition of its mobility by a (light-dependent) pH decrease. *Planta*, **192**, 581–589.
- Johnson, M.P., Davison, P.A., Ruban, A.V. and Horton, P. (2008) The xanthophyll cycle pool size controls the kinetics of non-photochemical quenching in Arabidopsis thaliana. *FEBS Lett.* **582**, 262–266.
- Kapila, J., De Rycke, R., Van Montagu, M. and Angenon, G. (1997) An Agrobacterium-mediated transient gene expression system for intact leaves. *Plant Sci.* **122**, 101–108.
- Külheim, C., Agren, J. and Jansson, S. (2002) Rapid regulation of light harvesting and plant fitness in the field. *Science*, **297**, 91–93.
- Li, X.-P., Björkman, O., Shih, C., Grossman, A.R., Rosenquist, M., Jansson, S. and Niyogi, K.K. (2000) A pigment-binding protein essential for regulation of photosynthetic light harvesting. *Nature*, **403**, 391–395.
- Li, X.-P., Müller-Moulé, P., Gilmore, A.M. and Niyogi, K.K. (2002a) PsbS-dependent enhancement of feedback de-excitation protects photosystem II from photoinhibition. *Proc. Natl Acad. Sci. USA*, **99**, 15222–15227.
- Li, X.-P., Phippard, A., Pasari, J. and Niyogi, K.K. (2002b) Structure–function analysis of photosystem II subunit S (PsbS) *in vivo*. *Funct. Plant Biol.* **29**, 1131–1139.
- Li, X.-P., Gilmore, A.M. and Niyogi, K.K. (2002c) Molecular and global time-resolved analysis of a *psbS* gene dosage effect on pH- and xanthophyll cycle-dependent nonphotochemical quenching in photosystem II. *J. Biol. Chem.* **277**, 33590–33597.
- Li, X.-P., Gilmore, A.M., Caffarri, S., Bassi, R., Golan, T., Kramer, D. and Niyogi, K.K. (2004) Regulation of photosynthetic light harvesting involves intrathylakoid lumen pH sensing by the PsbS protein. *J. Biol. Chem.* **279**, 22866–22874.
- Long, S.P., Marshall-Colon, A. and Zhu, X.-G. (2015) Meeting the global food demand of the future by engineering crop photosynthesis and yield potential. *Cell*, **161**, 56–66.
- Matsubara, S., Morosinotto, T., Osmond, C.B. and Bassi, R. (2007) Short- and long-term operation of the lutein-epoxide cycle in light-harvesting antenna complexes. *Plant Physiol.* **144**, 926–941.
- Melis, A. (2009) Solar energy conversion efficiencies in photosynthesis: minimizing the chlorophyll antennae to maximize efficiency. *Plant Sci.* **177**, 272–280.
- Mettler, T., Mühlhaus, T., Hemme, D. et al. (2014) Systems analysis of the response of photosynthesis, metabolism, and growth to an increase in irradiance in the photosynthetic model organism *Chlamydomonas reinhardtii*. *Plant Cell*, **26**, 2310–2350.
- Müller, P., Li, X.-P. and Niyogi, K.K. (2001) Non-photochemical quenching. A response to excess light energy. *Plant Physiol.* **125**, 1558–1566.
- Müller-Moulé, P., Conklin, P.L. and Niyogi, K.K. (2002) Ascorbate deficiency can limit violaxanthin de-epoxidase activity *in vivo*. *Plant Physiol.* **128**, 970–977.
- Murchie, E.H. and Niyogi, K.K. (2011) Manipulation of photoprotection to improve plant photosynthesis. *Plant Physiol.* **155**, 86–92.
- Nelson, D.R. (2006) *Cytochrome P450 Nomenclature, 2004*. Totowa, NJ: Humana Press.
- Nilkens, M., Kress, E., Lambrev, P., Miloslavina, Y., Müller, M., Holzwarth, A.R. and Jahns, P. (2010) Identification of a slowly inducible zeaxanthin-dependent component of non-photochemical quenching of chlorophyll fluorescence generated under steady-state conditions in Arabidopsis. *Biochim. Biophys. Acta*, **1797**, 466–475.
- Niyogi, K.K., Grossman, A.R. and Björkman, O. (1998) Arabidopsis mutants define a central role for the xanthophyll cycle in the regulation of photosynthetic energy conversion. *Plant Cell*, **10**, 1121–1134.
- Peers, G., Truong, T.B., Ostendorf, E., Busch, A., Elrad, D., Grossman, A.R., Hippler, M. and Niyogi, K.K. (2009) An ancient light-harvesting protein is critical for the regulation of algal photosynthesis. *Nature*, **462**, 518–521.
- Pogson, B.J., Niyogi, K.K., Björkman, O. and DellaPenna, D. (1998) Altered xanthophyll compositions adversely affect chlorophyll accumulation and nonphotochemical quenching in Arabidopsis mutants. *Proc. Natl Acad. Sci. USA*, **95**, 13324–13329.
- Raftery, A.E., Li, N., Sevcikova, H., Gerland, P. and Heilig, G.K. (2012) Bayesian probabilistic population projections for all countries. *Proc. Natl Acad. Sci. USA*, **109**, 13915–13921.
- Rockholm, D.C. and Yamamoto, H.Y. (1996) Violaxanthin de-epoxidase. *Plant Physiol.* **110**, 697–703.
- Rossi, L., Escudero, J., Hohn, B. and Tinland, B. (1993) Efficient and sensitive assay for T-DNA-dependent transient gene expression. *Plant Mol. Biol. Rep.* **11**, 220–229.
- Van der Hoorn, R.A.L., Laurent, F., Roth, R. and De Wit, P.J.G.M. (2000) Agrobacterium infiltration is a versatile tool that facilitates comparative analyses of

- Avr9/Cf-9-induced and Avr4/Cf-4-induced necrosis. *Mol. Plant Microbe Interact.* **13**, 439–446.
- Vieler, A., Wu, G., Tsai, C.-H. et al.** (2012) Genome, functional gene annotation, and nuclear transformation of the heterokont oleaginous alga *Nannochloropsis oceanica* CCMP1779. *PLoS Genet.* **8**, e1003064.
- Warrens, A.N., Jones, M.D. and Lechler, R.I.** (1997) Splicing by overlap extension by PCR using asymmetric amplification: an improved technique for the generation of hybrid proteins of immunological interest. *Gene*, **186**, 29–35.
- Wu, G., Fanzo, J., Miller, D.D., Pingali, P., Post, M., Steiner, J.L. and Thaler-Mercer, A.E.** (2014) Production and supply of high-quality food protein for human consumption: sustainability, challenges, and innovations. *Ann. N. Y. Acad. Sci.* **1321**, 1–19.
- Xie, X., Gu, W., Gao, S., Lu, S., Li, J., Pan, G., Wang, G. and Shen, S.** (2013) Alternative electron transports participate in the maintenance of violaxanthin de-epoxidase activity of *Ulva* sp. under low irradiance. *PLoS ONE*, **8**, e78211.
- Yamamoto, H.Y., Bugos, R.C. and Hieber, A.D.** (2004) *Biochemistry and Molecular Biology of the Xanthophyll Cycle*. Netherlands: Springer.
- Zhu, X.G., Ort, D.R., Whitmarsh, J. and Long, S.P.** (2004) The slow reversibility of photosystem II thermal energy dissipation on transfer from high to low light may cause large losses in carbon gain by crop canopies: a theoretical analysis. *J. Exp. Bot.* **55**, 1167–1175.
- Zhu, X.G., Long, S.P. and Ort, D.R.** (2010) Improving photosynthetic efficiency for greater yield. *Annu. Rev. Plant Biol.* **61**, 235–261.
- Zones, J.M., Blaby, I.K., Merchant, S.S. and Umen, J.G.** (2015) High-resolution profiling of a synchronized diurnal transcriptome from *Chlamydomonas reinhardtii* reveals continuous cell and metabolic differentiation. *Plant Cell*, **27**, 2743–2769.