

Contents lists available at ScienceDirect

Field Crops Research



journal homepage: www.elsevier.com/locate/fcr

Seasonal climate conditions impact the effectiveness of improving photosynthesis to increase soybean yield

Yufeng He^a, Megan L. Matthews^{a,b,*}

^a Carl R. Woese Institute for Genomic Biology, University of Illinois at Urbana-Champaign, IL 61801, USA
 ^b Department of Civil and Environmental Engineering, University of Illinois at Urbana-Champaign, IL 61801, USA

ARTICLETRIO	A B S T R A C T
Keywords: Photosynthetic assimilation Field climate conditions Soybean growth modeling Soybean yield estimation RuBP regeneration	<i>Context:</i> Photosynthetic stimulations have shown promising outcomes in improving crop photosynthesis, including soybean. However, it is still unclear to what extent these changes can impact photosynthetic assimilation and yield under long-term field climate conditions. <i>Objective:</i> In this paper, we present a systematic evaluation of the response of canopy photosynthesis and yield to two critical parameters in leaf photosynthesis: the maximum carboxylation rate of ribulose-1,5-bisphosphate carboxylase/oxygenase (V_{cmax}) and the maximum electron transport of the ribulose-1,5-bisphosphate regeneration rate (J_{max}). <i>Methods:</i> Using the field-scale crop model Soybean-BioCro and ten years of observed climate data in Urbana, Illinois, U.S., we conducted sensitivity experiments to estimate the changes in canopy photosynthesis, leaf area index, and biomass due to the changes in V_{cmax} and J_{max} . <i>Results:</i> The results show that 1) Both the canopy photosynthetic assimilation (A_n) and pod biomass yields were more sensitive to the changes in J_{max} , particularly at high atmospheric carbon-dioxide concentrations ([CO ₂]); 2]. Higher [CO ₂] undermined the effectiveness of increasing the two parameters to improve A_n and yield; 3) Under the same [CO ₂], canopy light interception and canopy respiration were key factors that undermined improvements in A_n and yield; 4) A canopy with smaller leaf area index tended to have a higher yield improvement, and 5) Increases in assimilations and yields were highly dependent on growing-season climatic conditions. The solar radiation, temperature, and relative humidity were the main climate drivers that impacted the yield improvement ment, and they had opposite correlations with improved yield during the vegetative phase compared to the reproductive phase. <i>Conclusions:</i> In a world with elevated [CO ₂], genetic engineering crop photosynthesis should focus more or improving J_{max} . Further, long-term climate conditions and yield at the field scale.

1. Introduction

Climate change and population growth are pressing problems and will continue to pose immediate risks to food security and increase demands on food production (Godfray & Garnett, 2014; Hasegawa et al., 2018). A central topic in plant science is to improve crop yields by increasing photosynthetic efficiency (Long et al., 2015; Ort et al., 2015; Raines, 2011; Simkin et al., 2019; Zhu et al., 2008). Transgenic

engineering of the photosynthetic pathway has been shown to increase photosynthesis rates in a wide variety of crop types, such as tobacco (López-Calcagno et al., 2020), soybean (Hay et al., 2017), wheat (Parry et al., 2011), maize (Salesse-Smith et al., 2018) and rice (Ku et al., 2000).

Increasing carbon-dioxide concentration ([CO₂]) in the atmosphere could lead to greater photosynthesis rates (Ainsworth & Rogers, 2007; Leakey et al., 2009b; Specht et al., 1999). In addition to [CO₂], these

* Corresponding author at: Department of Civil and Environmental Engineering, University of Illinois at Urbana-Champaign, IL 61801, USA. *E-mail address:* mlmatth2@illinois.edu (M.L. Matthews).

https://doi.org/10.1016/j.fcr.2023.108907

Received 21 October 2022; Received in revised form 31 January 2023; Accepted 22 March 2023 Available online 29 March 2023 0378-4290/© 2023 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/). rates are also highly dependent on other environmental factors, such as temperature and light. Model results also suggest alterations of the kinetic properties of Ribulose-1,5-bisphosphate carboxylase-oxygenase (Rubisco) and/or the regeneration of Ribulose-1,5-bisphosphate (RuBP) would increase photosynthetic assimilation rates under future climate conditions (Long et al., 2006). However, the extent to which these improvements would impact crop photosynthesis and yield under different field climate conditions is still unknown.

The correlation between assimilation and yield in annual crops is complex (Buttery et al., 1981; Evans & Fischer, 1999; Gifford & Evans, 1981; Specht et al., 1999). A recent study has shown that there is a large variability in the extent of these correlations with yield due to complex impacts across biological and temporal scales (Wu et al., 2023). For example, the effectiveness of photosynthetic enhancement could be undermined by increased photorespiration (Curtis et al., 1969) and nitrogen requirements (Sinclair & Wit, 1976; Yin et al., 2022). Further, it has been shown that a short-term, hourly or daily, gain in photosynthesis assimilation from increased [CO2] does not necessarily translate into a long-term yield gain (Sims et al., 1998). Particularly under constantly varying field conditions, the final yield is an integration of daily and hourly changes over the entire growing season. Therefore, determining the main contributor to photosynthesis at a seasonal scale is critical to evaluate the potential of photosynthetic improvements on yields. Using models can help us understand the overall sensitivities of assimilations and yields on changing climate conditions for a long climatically significant period, providing information for identifying new photosynthetic improvements.

In the steady-state biochemical model of photosynthesis, known as the Farguhar-von Caemmerer-Berry (FvCB) model (von Caemmerer, 2000), the net assimilation rate (A_n) is limited by three processes: 1) A_c , the rate of Rubisco carboxylation; 2) A_i , the rate of RuBP regeneration, as determined by the electron transport fluxes; and 3) A_p , the triose phosphate utilization (TPU) rate. The TPU rate has been found to rarely be the limiting factor at the current and the near-future atmospheric [CO2] levels due to its high internal [CO2] requirement (Kumarathunge et al., 2019; Sharkey, 2019). Most studies have focused on improving Ac and Ai to increase photosynthesis assimilation (López-Calcagno et al., 2020). Corresponding to A_c and A_j are two commonly used parameters: the maximum carboxylation rate of Rubisco (V_{cmax}) and the maximum electron transport and RuBP regeneration rate (J_{max}) . These two high-level metrics are often used as measures of the photosynthetic efficiency of transgenic plants (López-Calcagno et al., 2020; Suzuki et al., 2009; Wu et al., 2019) or plants grown under different environments (Bernacchi et al., 2005; Galmés et al., 2015).

 V_{cmax} and J_{max} have been increased by 10–30% by increasing the content and/or activity of certain photosynthetic enzymes in the Calvin–Benson cycle, resulting in an increase of 10–20% in A_n and leaf biomasses in tobacco (Lefebvre et al., 2005; López-Calcagno et al., 2020; Rosenthal et al., 2011; Simkin et al., 2015). Similar improvements have also been accomplished in transgenic soybeans with 4–14% increases in A_n and 4–8% increases in V_{cmax} and J_{max} , which are expected to prevent a decrease in yield under the combined effects of future warming and elevated [CO₂] (Köhler et al., 2016).

Most studies have focused on the sensitivities of environmental conditions, such as changing [CO₂], temperature, and light (Cai et al., 2018), where V_{cmax} and J_{max} are treated as derived parameters from the A- C_i curve. However, this approach alone leaves a knowledge gap in systematically understanding the sensitivity of the two parameters under different environments. While V_{cmax} and J_{max} directly contribute to the magnitudes of A_c and A_j , respectively, changing these parameters, either individually or together, alters how often each rate is limiting assimilation during a long-term simulation. Therefore, a comprehensive sensitivity study on the two parameters can determine the respective contributions of Rubisco and RuBP regeneration under a wide range of climate conditions, which can be used to support effective lab testing. This information may further help identify strategies for improving

photosynthesis best suited for current and future climate conditions.

When designing strategies for increasing photosynthetic assimilation and yield it is also essential to consider canopy gradients and microclimates. Not only can climate alter crop development progress (He et al., 2020), but the impacts of the same climate condition during different developmental phases will have varying effects on crop yield (Wu et al., 2023). For example, an optimum light and temperature condition may differ in determining the final yield for the reproductive and vegetative phases. Many real-world conditions that can be simulated by advanced crop growth models are difficult, if not impossible, to replicate in the lab. These conditions may include light attenuation due to leaf canopy shading effects, constant radiation changes due to cloud cover, and extreme weather conditions in temperature and precipitation. Using real climate field data instead of controlled lab data, we can have a more realistic response of photosynthesis and yield and explore the variabilities in climate driving forcing and predicted quantities.

In this paper, we use a crop growth model, Soybean-BioCro (Matthews et al., 2022), to simulate soybean growth using observed climate data and quantify the impacts of changing V_{cmax} and J_{max} on A_n and yields. We further evaluate how climate conditions impact the effectiveness of increasing V_{cmax} and J_{max} at a seasonal scale. Our specific objectives are to 1) estimate the changes in A_n and yields with a range of changes in V_{cmax} and J_{max} under current and future [CO₂] conditions; 2) examine the causes of reduced returns in A_n and yields for a 20% increase in V_{cmax} and J_{max} ; and 3) quantify the contribution of each climate driver (including solar radiation, air temperature, precipitation, relative humidity and wind speed) to the yield gain for the vegetative and reproductive phases respectively.

2. Material and methods

2.1. Model description

All of the field-scale soybean simulations were implemented using Soybean-BioCro, a crop growth model that is part of the BioCro model framework (Lochocki et al., 2022; Matthews et al., 2022) . The model calibration and validation for Soybean-BioCro were conducted against four years (i.e., 2002, 2004, 2005, and 2006) of observed climate data, as well as soybean biomass measurements (Pioneer 93B15) collected at the SoyFACE facility (40.04°N, 88.23°W) at the University of Illinois at Urbana-Champaign (Matthews et al., 2022; Morgan et al., 2005). Soybean-BioCro incorporates the Farguhar-von Caemmerer-Berry (FvCB) model and Ball-Berry stomatal conductance model to simulate a 10-layer canopy photosynthesis that contains sunlit and shaded leaves (Fig. S1). It then calculates photothermal development rate and uses logistic functions to partition assimilated carbon into biomass. The growth and senescence of leaf, stem, grain, and root biomasses are estimated at an hourly time step and integrated throughout the growing season (Matthews et al., 2022).

2.2. Definition of main quantities

We define the following metric for representing the relative change of variables,

$$Relative \ change = \frac{V_{XJX} - CTL}{|CTL|} * 100\%$$
(1)

Where CTL is the model simulation with the default values of V_{cmax} and J_{max} . VxJx represents the simulation experiments with changed values in the two parameters, where *x* represents the amount of change in percentage.

The photosynthetic water use efficiency (WUE) at the canopy level (Medrano et al., 2015) is calculated as,

$$WUE = \frac{A_n}{E} (\mu mol \ mmol^{-1})$$
⁽²⁾

Where A_n is the canopy-level net assimilation rate, and *E* is the canopy-level evapotranspiration rate. The leaf level evapotranspiration is estimated by the Penman-Monteith evapotranspiration model, which is then integrated over the canopy layers to obtain *E* (Fig. S1).

2.3. Data and experiment design

Ten years of weather data from 2006 to 2015 in Bondville, Illinois, were used for the model simulations. The driving climate variables include air temperature, precipitation, solar radiation, relative humidity, and wind speed. All observed climate data were obtained from the Surface Radiation Budget Network (https://gml.noaa.gov/grad/surfrad /) except for the precipitation. The precipitation data were received from the Illinois Climate Network (https://www.isws.illinois.edu/dat/).

Two key parameters in the FvCB model, the maximum carboxylation rate of Rubisco (V_{cmax}) and the maximum electron transport and RuBP regeneration rate (J_{max}), were investigated in a series of model sensitivity experiments. For the control experiments (CTL), the default values of the two parameters are, V_{cmax} = 110 and J_{max} = 195 μ mol m⁻² s⁻¹ (Matthews et al., 2022). For the sensitivity experiments (VxJx), we scaled V_{cmax} and J_{max} by a range of values from – 50% to + 50% at a 5% step size. Four [CO₂] levels were simulated: 400, 600, 800, and 1000 ppm, where 400 ppm is approximately the current level of the atmospheric [CO₂]. The 600 and 1000 ppm values are in the range of the predicted [CO₂] in an extreme scenario for 2050 and 2100, respectively (Joos et al., 2001). The abbreviation V20J20 was used to represent the experiments with a 20% increase in both V_{cmax} and J_{max} .

We then estimated the absolute and relative changes (Eq. 1) in six selected variables of interest: the harvestable biomasses of pod and shoot (sum of pod, leaf, and stem), seasonal averages of daily maximum and mean of A_n , maximum leaf area index (LAI) during the growing season and average of daily mean WUE. The relative changes represent the effectiveness of changing V_{cmax} and J_{max} at corresponding climate and [CO₂] conditions. Unless otherwise specified, A_n refers to the canopy-level A_n .

2.4. Gradient descent

. .

To find the steepest path on the heatmap of the relative change against V_{cmax} and J_{max} , we used a simple gradient descent algorithm as follows,

$$X_{n+1} = X_n - \alpha \nabla f(X_n) \tag{3}$$

Where X_n is a two-dimensional coordinate of V_{cmax} and J_{max} at the n_{th} step. α is the step size of path searching (α =0.001 was used in the simulations). The gradient of the function *f* can be calculated as,

$$\nabla f(X) = \begin{pmatrix} \frac{\partial f}{\partial x} \\ \frac{\partial f}{\partial y} \\ \frac{\partial f}{\partial y} \end{pmatrix}$$
(4)

Where x and y represent the V_{cmax} and J_{max} , respectively. To estimate the derivatives of discrete functions, we used the following central difference approximation method,

$$\frac{\partial f}{\partial x} = \lim_{h \to 0} \frac{f(x+h,y) - f(x-h,y)}{2h}$$
(5)

$$\frac{\partial f}{\partial y} = \lim_{h \to 0} \frac{f(x, y+h) - f(x, y-h)}{2h}$$
(6)

Where h= 0.01, and f(x,y) is estimated using a non-linear spline interpolation from the Akima package in R (Akima & Gebhardt, 2022).

2.5. Bootstrap of climate data and partial rank correlation

To better represent the day-to-day variability of the 10-year observed climate data, we used a bootstrap method to create 1000 scenarios of annual climate data based on a re-sampling process for all days of the year (DOY). For each bootstrapped climate scenario, all of the climate drivers were randomly sampled on a daily basis from one of the 10 years of weather data. For a given DOY, all 24 hours for all of the climate variables were sampled from the same year. This sampling strategy preserves the diurnal relationships and any relationships between the climate drivers (e.g., sunlight and temperature, precipitation and humidity). The bootstrapped climate scenarios were statistically generated to better represent the 10-year variability, and do not represent future climate conditions.

To evaluate the relationships between pod biomass gains and climate conditions at the vegetative and reproductive phases, we used partial rank correlation to represent each climate driver's contribution to the biomass gain. The partial rank correlation coefficients (PRCC) were calculated using the epiR package in R (Stevenson et al., 2022).

3. Results

3.1. Sensitivity of assimilation, LAI, and yield to the changes in V_{cmax} and J_{max}

Biomasses, LAI, and A_n were all impacted by the changes in V_{cmax} and J_{max} . Larger values were predicted for all variables as the atmospheric [CO₂] increased (Fig. 1). At 400 ppm, the pod and shoot biomasses ranged from 4.9–7.0 and 6.3–9.6 Mg/ha, respectively, compared with their controls of 6.6 and 9.1 Mg/ha. At 800 ppm, these ranges shifted to 6.1–8.0 and 8.1–11.2 Mg/ha with the controls of 7.7 and 10.8 Mg/ha. Similar trends were observed at 400 and 800 ppm for: LAI with ranges of 4.3–8.1 and 5.4–9.9 m²/m² and the respective controls of 7.3 and 9.3 m²/m², daily mean A_n with ranges of 3.6–5.5 and 4.7–6.4 μ mol m^{-2} s^{-1} and the controls of 5.2 and 6.2 μ mol $m^{-2} s^{-1}$ and the controls of 33.4 and 42.3 μ mol $m^{-2} s^{-1}$.

Although higher [CO₂] increased assimilation and yield in general, the increases were seen in both the sensitivity experiments (VxJx) and the corresponding controls (CTL). To better understand the effectiveness of changing V_{cmax} and J_{max} we quantified the relative difference between VxJx and CTL at all [CO₂] levels (Fig. 2). Varying V_{cmax} and J_{max} rates had a more significant impact on assimilation and yield under lower levels of atmospheric [CO₂] (Fig. 2). When compared with their controls, increasing both V_{cmax} and J_{max} at 400 ppm resulted in increases of up to 18% in the maximum A_n , 13% in the LAI and 8% in both the shoot and pod biomasses (Fig. 2). Decreasing V_{cmax} and J_{max} resulted in losses of up to 46% in the maximum A_n , 43% in the LAI, 33% in the shoot and 29% in the pod (Fig. 2). This trend was also observed at higher [CO₂], but with a decreasing range of impacts as [CO₂] increased, such as the pod biomass which was only increased by 4.2% at 800 ppm versus 8% at 400 ppm (Fig. 2).

In all simulated scenarios, the variables of interest were more sensitive to changes in J_{max} than V_{cmax} . Starting from the control experiments (Fig. 2, gray triangles), the path with the largest ascending gradient was mainly determined by increasing J_{max} , particularly at [CO₂] above 400 ppm, where increasing V_{cmax} had little to no impact as indicated by the gradient paths (Fig. 2, black dotted lines). The model was more sensitive to changes in J_{max} as the assimilation rate was almost entirely limited by A_j , the rate of RuBP regeneration (Fig. 3b). To achieve maximal gains at 400 ppm, however, increasing V_{cmax} was still necessary to maximize the yield and assimilation as increasing J_{max} increased the transition point of the intracellular carbon, C_i , where the limiting rate switched between A_c and A_j from ~280 ppm to ~410 ppm at high light (Fig. 3a). At a lower light condition of 800 $\mu mol m^{-2} s^{-1}$, the transition point of C_i was reduced to ~220 ppm and ~280 ppm



Fig. 1. Heatmaps of model estimations of the five variables at four $[CO_2]$ levels (400, 600, 800, and 1000 ppm) in responses to the changes in V_{cmax} and J_{max} . The variables include annual pod and shoot biomass, maximum LAI, daily mean, and maximum net assimilation rate (An). The control experiment (grey triangle) has the default values of V_{cmax} (=110) and J_{max} (=195). The values shown were calculated by the ten-year averages from 2006 to 2015.



Fig. 2. Heatmaps of the changes (Δ) in five variables at four [CO₂] levels in responses to the changes in *V_{cmax}* and *J_{max}*. The variables include annual pod and shoot biomass, maximum LAI, daily mean, and maximum net assimilation rate (An). The colors represent the level of changes (in %) of each sensitivity experiment regarding the control experiment (grey triangle). The values were calculated by the ten-year averages. The black dotted lines represent the fastest-changing paths with the largest gradients.



Fig. 3. (a) A-Ci curves of the A_c and A_j for the control case and the A_j with a 20% increase in J_{max} at a light intensity (Q) of 1500 µmol m⁻² s⁻¹. (b) Same as (a) but with a light intensity (Q) of 800 µmol m⁻² s⁻¹. (c) Frequency of the three limiting factors for the sunlit top layer in the Soybean-BioCro hourly simulations. (d) Frequency of the C_i levels for the sunlit top layer. This example used the data for the growing season of 2006 at 400 ppm [CO₂]. The A-Ci curves were obtained using the leaf-level Farquhar model with the same parameters as the full FvCB model in the Soybean-BioCro.



Fig. 4. Impacts of increasing V_{cmax} and J_{max} by 20% (V20J20) at four [CO₂] levels on the relative changes (%) of (a) Pod, (b) Shoot, (c) average of daily maximum A_n throughout the growing season, (d) average of daily mean A_n , (e) max LAI during the growing season and (f) average of WUE. $\Delta =$ V20J20 – CTL.

(Fig. 3b). At 400 ppm of [CO₂], the hourly C_i showed few occurrences below 250 ppm and a small amount between 250 and 300 ppm (Fig. 3c). Consequently, more instances of A_c occurred as the limiting rate when the transition point of C_i increased (Fig. 3d). Thus, at current levels of atmospheric [CO₂], increasing both J_{max} and V_{cmax} were required to maximize assimilation, but at a higher [CO₂] the system became much less likely to be limited by A_c , therefore neglecting the contributions from increased V_{cmax} . In fact, at a higher [CO₂], V_{cmax} can even be decreased to 85% of its control without any negative impact on assimilation or biomass if J_{max} were unchanged or increased (Fig. 2).

The results from the sensitivity analysis (Figs. 1-2) represent each variable's interannual averages over the ten years. However, interannual variations were significant for all variables within those ten years. For example, increasing both V_{cmax} and J_{max} by 20% resulted in larger relative gains in some of the simulated years than in others (Fig. 4). At 400 ppm, the daily maximum A_n ranged from a 6.9%-10.6% increase (Fig. 4c), followed by the peak LAI with gains of 4.7%–7.9% (Fig. 4e), the daily mean An ranged from a 2.3%-5.5% increase (Fig. 4d), the shoot biomass ranged from a 1.6%-5.1% increase (Fig. 4b), and the pod biomass ranged from a -1.4%–5.3% change (Fig. 4a). In two of the ten years, the pod biomass was lower in the V20J20 scenario than the CTL. In 2007, there was a 0.5% loss and in 2014 there was a 1.4% loss in pod biomass. In the other eight years, the pod biomass increased by 3%-5.3% (Fig. 4a). Overall, the relative gains for each variable became consistently smaller as [CO₂] increased, but similar interannual patterns were predicted. The changes in WUE showed little similarity among the [CO₂] scenarios. Although there was a significant interannual variation over the simulated period, the 10-year average of the relative changes was minimal compared with the other variables (Fig. 4f).

3.2. Key factors that limit the effectiveness of increasing V_{cmax} and J_{max}

Although the daily maximum A_n increased more than the other variables, it was still limited to a 10% increase when V_{cmax} and J_{max} were both increased by 20% at 400 ppm (Fig. 4). The improvements in the daily maximum A_n were even smaller at higher [CO₂]. This reduced gain in assimilation and yield was largely determined by the amount of light received by the canopy. At a high light condition of 1500 μ mol $m^{-2} s^{-1}$, increasing V_{cmax} and J_{max} by 20% resulted in an increase in the leaf-level A_n by approximately 18% when C_i was between 400 and 600 ppm (Fig. 5a). While at a low light condition of 800 μ mol $m^{-2} s^{-1}$, the leaf-level A_n was only increased by 8% (Fig. 5b).

The leaf-level photorespiration was also greater with the increased V_{cmax} and J_{max} (Fig. S2). However, the increases in photorespiration

were rather small when compared with the increases in carboxylation under both high and low light conditions. At a C_i of 400 ppm, photorespiration was only increased by 10% of the increase seen in the carboxylation rate (Table S1). At a higher C_i of 600 ppm, the increase in photorespiration was only 7% of the increase in carboxylation (Table S1).

In the Soybean-BioCro simulations, the incoming light was mainly received by the sunlit part of the canopy. Throughout the growing season, the daily maximum solar radiation received by the sunlit canopy was about 700–800 μ mol $m^{-2} s^{-1}$ (Fig. 6a) due to the light interception calculated using an average leaf orientation of the canopy. Slightly lower sunlit radiation was estimated for the scenarios with increased V_{cmax} and J_{max} (V20J20) among all canopy layers, mostly distinctive at the bottom layers (Fig. 6b). This was due to a larger LAI simulated for the V20J20 scenarios, which caused more of the lower layers to be shaded. Consequently, there was a diminished effect on the daily maximum A_n at the bottom layers while there was a more prominent effect at the top layers (Fig. 6c). The combined effect from all layers determined the amount that A_n was increased.

The canopy dark respiration (R_d) was also increased with increased V_{cmax} and J_{max} , which led to further losses on the A_n . However, when compared with gross assimilation, R_d only contributed to a small percentage in the daily maximum A_n (Fig. S3a & c) when there was high light in a day. Therefore, the increases in the daily maximum A_n (\sim 8%) were close to those estimated at the leaf-level at 400 ppm (Fig. 5b). The contribution of R_d became particularly significant when considering the daily mean A_n (Fig. S3b & d), where changes in R_d can surpass changes in gross assimilation under low light and dark conditions. Therefore, the increases in A_n were further undermined (Fig. 4d) in its daily mean when compared with its daily maximum (Fig. 4 c).

3.3. Main climate drivers that determine the yield improvement

One thousand bootstrapped scenarios were generated from the tenyear observed climate data to better represent the climatic variability. Increases in pod biomass (Δ pod) varied from 0.4% to 5.7% among the bootstrapped samples when V_{cmax} and J_{max} were increased by 20% at the [CO₂] of 400 ppm (Fig. 7a). The lower quantile consisting of 250 samples (set-25) predicted 2.2% \pm 0.5% in Δ pod, and the upper quantile (set-75) predicted 4.4% \pm 0.4% in Δ pod. The Δ pod time series for the two sets started to diverge from the beginning of the predicted reproductive phase on about DOY 210 until the end of the growing season (Fig. 7b). A significant difference in the maximum LAI changes (Δ LAI) was found between the two sets (Fig. 7c), where set-25 predicted



Fig. 5. Relationships between net assimilation rate (A_n) and intercellular CO₂ concentration (C_i) at the leaf level under two light conditions, (a) $Q = 1500 \ \mu mol \ m^{-2} \ s^{-1}$ and (b) $Q = 800 \ \mu mol \ m^{-2} \ s^{-1}$. The sensitivity experiment (V20J20) has a 20% increase in both V_{cmax} and J_{max} . The A_n-C_i curves are obtained using the leaf-level Farquhar model with the same parameters as the full FvCB model used in this study.



Fig. 6. (a) Distribution of Qmax_sunlit for all layers and 1000 bootstrap samples for the CTL scenarios. (b) 10-layer profile of daily max radiation on the sunlit canopy (Qmax_sunlit) at [CO₂] of 400 ppm. (c) 10-layer profile of the maximum *An* of the sunlit canopy (An_sunlit). Both Qmax_sunlit and An_sunlit represent quantities per leaf area.



Fig. 7. Changes in pod biomass and LAI due to an increase of V_{cmax} and J_{max} by 20% at [CO₂] of 400 ppm driven by bootstrap climate samples. (a) Frequency distribution of pod gains (Δ pod, %) for the 1000 bootstrap samples. (b) Time series of the absolute difference of pod biomass (Δ pod, Mg/ha), separated by lower (set-25; red line) and upper (set-75; black line) quantiles. Shaded bands represent one standard deviation for each set. (c) Same as (b), but for the difference of maximum LAI (Δ LAI). (d) The relationship between Δ LAI (%) and Δ pod (%) with a linear regression line (red).

consistently higher Δ LAI than set-75, which was inversely correlated with their Δ pod predictions. This was further confirmed by a significant negative correlation (r = -0.41, p < 0.001) between Δ LAI and Δ pod for all bootstrap samples (Fig. 7d).

The large variation in the predicted Δpod originated from different growing-season climate conditions that were generated using the boot-strap sampling of the 10-year period previously examined. Two growth phases of soybean are the vegetative and reproductive phases. Climate

conditions during each of these phases contributed to Δpod differently. Among the five climate variables, solar radiation (Q), air temperature (T), and relative humidity (RH) showed significant partial rank correlations with Δ pod at both growth phases (Fig. 8). During the vegetative phase, Q was negatively correlated with Δpod (PRCC=-0.37, p < 0.001), while T and RH were positively correlated with Δpod (PRCC=0.39 & 0.17, p < 0.001). In comparison, these correlations were reversed during the reproductive phase with similar correlation levels (Fig. 8). The opposing correlations between the two phases can be explained by examining the relationship between the climate drivers and $\Delta A_n / \Delta LAI$. In both phases, T was negatively correlated with ΔA_n and Q was positively correlated with ΔA_n (Fig. S4). Since a higher A_n during the vegetative phase led to a higher LAI and given that Δ LAI correlated negatively with Δpod (Fig. 7d), ΔA_n was negatively correlated with Δpod (Fig. S5a). A higher A_n during the reproductive phase, however, did not impact LAI and contributed directly to pod biomasses (Fig. S5b). Correlations between Δpod and the other two climate variables, precipitation and wind speed, were not statistically significant under the climate conditions at the study site (Fig. 8). The model simulations did not show significant water stress, so the results and their interpretations are limited to water non-limiting conditions.

4. Discussion

4.1. Increasing J_{max} contributes more to the improvements of assimilation and yield

At the current $[CO_2]$ of about 400 ppm, increasing the rate of Rubisco carboxylation has little impact, and thus there is a low potential for improving crop photosynthesis through increasing this rate in soybean (Ainsworth & Rogers, 2007; Leakey et al., 2009a; Wise et al., 2004). This conclusion is consistent with our model prediction that a more significant contribution was found in J_{max} than V_{cmax} (Fig. 2). At a high [CO₂] condition, assimilation is mainly limited by the capacity for RuBP regeneration (Long et al., 2004), same as our model representation of A_j being the main limiting factor under increased atmospheric [CO₂] (Fig. 3).

Previous modeling studies have demonstrated a possible nitrogen overinvestment in Rubisco in existing annual crops (Wu et al., 2019; Yin et al., 2022). At the higher [CO₂] level predicted for the future, there is an even smaller, and sometimes negligible, impact from increasing V_{cmax} , implying that Rubisco activity could be decreased without losing photosynthetic benefits from increased J_{max} (Fig. 2). Therefore, a potential pathway to further boost assimilation could focus on reallocating the resources like nitrogen from Rubisco to other parts of the plant.



Fig. 8. Partial rank correlation coefficients between Δ pod and climate variables are separated into the vegetative phase (indicated by 1) and reproductive phase (indicated by 2). The five climate variables are solar radiation (Q), air temperature (T), precipitation (P), relative humidity (RH), and wind speed (WS). The climate variables are the 1000 bootstrap samples from the 10-year observed data in Illinois. The three-asterisk symbols represent a significance level of the coefficients with p-values less than 0.001. The ns symbol stands for not significant.

4.2. A higher $[CO_2]$ lowers the effectiveness of increasing V_{cmax} and J_{max}

Higher atmospheric [CO₂] improved yields in both the CTL and VxJx simulations (Fig. 1), but the effectiveness of increasing V_{cmax} and J_{max} was reduced as [CO₂] increased (Figs. 2 & 4). A previous experimental study reported that transgenic soybeans showed larger V_{cmax} and J_{max} than the wildtypes under both ambient and elevated [CO₂] (Hay et al., 2017). The effectiveness of changing V_{cmax} and J_{max} can be viewed as the change in A_n per unit change of V_{cmax} or J_{max} . Following this definition, it is evident that increasing V_{cmax} or J_{max} was more effective in enhancing A_n at a lower [CO₂] (Hay et al., 2017). However, an increase in yield was still predicted under elevated [CO₂] with a higher V_{cmax} and J_{max} compared to the control (Fig. 1).

4.3. Why were increases in A_n and yield much lower than in V_{cmax} and J_{max} ?

Significant reductions in the returns in A_n and yield have been found in both C3 and C4 crops with boosted photosynthesis (Sinclair et al., 2004; Wu et al., 2018). Similarly, we estimated that a 20% increase in V_{cmax} and J_{max} led to less than a 10% gain in the maximum A_n and even lower gains in the mean A_n and pod biomasses (Fig. 4). The reduced gain in the maximum A_n was primarily due to the light interception at the canopy level. The percentage gain in A_n can approach 20% under high light, but is only around half of that under low light (Fig. 5). Canopy light use efficiency is a key factor that impacts photosynthesis and yield (Koester et al., 2014; Yin et al., 2022), therefore identifying optimal canopy structures could further improve the effectiveness of stimulating photosynthesis.

 R_d was another key factor that undermined the gain in A_n , particularly in the daily mean A_n due to diurnal variations of the increases in both gross assimilation and R_d (Fig. S3). It is important to consider both day and night hours to calculate the daily mean since the yield is continuously accumulated. This explains why increases in the daily mean A_n were much closer to and better correlated with gains in the yield than that in the daily maximum A_n (Fig. 4).

The increases in pod biomass were even less than that in the daily mean A_n due to additional losses from the development and carbon allocation processes, which can further undermine the effectiveness of increased V_{cmax} and J_{max} . The biomass partitioning varies significantly at different crop development stages (Matthews et al., 2022), causing a heterogeneous transfer of assimilated carbon into the final yields. Sensitivity analyses, as shown in this study, use coupled canopy photosynthesis and carbon allocation models and can reveal complex input-output information during the entire crop development period, which is helpful for decision-making toward an effective photosynthetic improvement to increase yield.

4.4. Canopy with smaller LAI can be more effective

Most of the light absorption occurs at the top of the canopy, with photosynthetic assimilation decreasing with depth (Amthor, 1994). Using a multiple-layer canopy model, we were able to capture an important feature that a soybean canopy with smaller LAI was generally more effective in gaining biomass for increased V_{cmax} and J_{max} (Fig. 7). This is mainly due to the changed light use among the vertical canopy profile, where a smaller canopy has decreased shading from top layers, increasing the light received by the lower leaves and thus producing a higher yield overall (Fig. 6). Since leaf growth mainly occurs during the vegetative phase, lower radiation and/or higher temperature during this period would lead to a smaller LAI and thus a higher yield (Fig. 8, S4 & S5). Previous experimental studies also found similar results, and an optimal LAI may exist for obtaining the highest efficiency in canopy photosynthesis, which does vary with crop types and environmental conditions (Srinivasan et al., 2017; Tagliapietra et al., 2018).

4.5. On WUE

We found no significant changes in WUE for the simulated 10-year period when considering only the changes of V_{cmax} and J_{max} (Fig. 4f). This is because the canopy assimilation and transpiration can change simultaneously. Two studies on the photosynthetic stimulation of tobacco have demonstrated opposite results. (López-Calcagno et al., 2020) found increased WUE with increased V_{cmax} and J_{max} , while (Simkin et al., 2015) reported that these increases in A_n were accompanied by increased stomatal conductance and decreased WUE. Whether changing the two parameters would impact the WUE seems inconclusive. While yield is the primary focus for photosynthetic improvements, evaluating WUE is crucial to understanding regional water sustainability, breeding resilience under extreme climate conditions like drought, and designing optimal cropping systems with soybean (Baath et al., 2021).

5. Conclusions

Despite many efforts spent on improving plant photosynthetic assimilation and yield by increasing V_{cmax} and J_{max} , how effective such an approach is under varying environmental conditions remained uncertain. In this paper, we used a semi-mechanistic crop growth model, for the first time, to systematically evaluate the effectiveness of increasing V_{cmax} and J_{max} on soybean growth using real field climate data and its statistical synthetics. The detailed results showed how changes in A_n and biomasses respond to the changes in V_{cmax} and J_{max} under different [CO2] and field climatic conditions. This work provides a framework for evaluating the effectiveness of increasing or decreasing V_{cmax} and J_{max} on assimilation and yield. While it is crucial to explore the potential of boosting leaf photosynthesis at the plant level, to achieve better and potentially maximum effectiveness of these improvements, we should also consider designing better canopy structures and incorporate the impacts of seasonal climate variability in the region of interest.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data Availability

All codes and data used in this study are free to be downloaded at the GitHub repository (https://github.com/cropsinsilico/Soybean-Sensitivity).

Acknowledgments

This work was supported by the Realizing Increased Photosynthetic Efficiency (RIPE) project, which is funded by the Bill & Melinda Gates Foundation, Foundation for Food & Agriculture Research (FFAR), and the UK Foreign, Commonwealth and Development Office (FCDO) under grant number OPP1172157. The content of this publication is solely the responsibility of the authors and does not necessarily represent the official views of the funding agencies. We greatly thank the collective discussions and suggestions from members of the Long Lab on the manuscript. All computational simulations were done through the high-performance computation facility of the BioCluster at the Carl R. Woese Institute for Genomic Biology (IGB) at the University of Illinois at Urbana-Champaign (UIUC).

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.fcr.2023.108907.

References

- Ainsworth, E.A., Rogers, A., 2007. The response of photosynthesis and stomatal conductance to rising [CO₂]: mechanisms and environmental interactions. Plant Cell Environ. 30, 258–270. https://doi.org/10.1111/j.1365-3040.2007.01641.x.
- Akima, H., Gebhardt, A., 2022. akima: Interpolation of Irregularly and Regularly Spaced Data [WWW Document]. URL https://CRAN.R-project.org/package=akima.
- Amthor, J.S., 1994. Scaling CO₂-photosynthesis relationships from the leaf to the canopy. Photosynth Res. 39, 321–350. https://doi.org/10.1007/BF00014590.
- Baath, G.S., Northup, B.K., Rao, S.C., Kakani, V.G., 2021. Productivity and water use in intensified forage soybean-wheat cropping systems of the US southern Great Plains. Field Crops Res. 265, 108086 https://doi.org/10.1016/j.fcr.2021.108086.
- Bernacchi, C.J., Morgan, P.B., Ort, D.R., Long, S.P., 2005. The growth of soybean under free air [CO₂] enrichment (FACE) stimulates photosynthesis while decreasing in vivo Rubisco capacity. Planta 220, 434–446. https://doi.org/10.1007/s00425-004-1320-8
- Buttery, B.R., Buzzell, R.I., Findlay, W.I., 1981. Relationships among photosynthetic rate, bean yield and other characters in field-grown cultivars of soybean. Can. J. Plant Sci. 61, 190–197. https://doi.org/10.4141/cjps81-029.

von Caemmerer, S., 2000. Biochemical Models of Leaf Photosynthesis. Csiro publishing.

- Cai, C., Li, G., Yang, H., Yang, J., Liu, H., Struik, P.C., Luo, W., Yin, X., Di, L., Guo, X., Jiang, W., Si, C., Pan, G., Zhu, J., 2018. Do all leaf photosynthesis parameters of rice acclimate to elevated <scp>CO</scp> 2, elevated temperature, and their combination, in <scp>FACE</scp> environments? Glob. Chang Biol. 24, 1685–1707. https://doi.org/10.1111/gcb.13961.
- Curtis, P.E., Ogren, W.L., Hageman, R.H., 1969. Varietal effects in soybean photosynthesis and photorespiration 1. Crop Sci. 9, 323–328. https://doi.org/ 10.2135/cropsci1969.0011183×000900030021x.
- Evans, L.T., Fischer, R.A., 1999. Yield potential: its definition, measurement, and significance. Crop Sci. 39, 1544–1551. https://doi.org/10.2135/ cropsci1999.3961544x.
- Galmés, J., Kapralov, M. v, Copolovici, L.O., Hermida-Carrera, C., Niinemets, Ü., 2015. Temperature responses of the Rubisco maximum carboxylase activity across domains of life: phylogenetic signals, trade-offs, and importance for carbon gain. Photosynth Res 123, 183–201. https://doi.org/10.1007/s11120-014-0067-8.
- Gifford, R.M., Evans, L.T., 1981. Photosynthesis, carbon partitioning, and yield. Annu Rev. Plant Physiol. 32, 485–509. https://doi.org/10.1146/annurev. pp.32.060181.002413.
- Godfray, H.C.J., Garnett, T., 2014. Food security and sustainable intensification. Philos. Trans. R. Soc. B: Biol. Sci. 369, 20120273 https://doi.org/10.1098/rstb.2012.0273.
- Hasegawa, T., Fujimori, S., Havlík, P., Valin, H., Bodirsky, B.L., Doelman, J.C., Fellmann, T., Kyle, P., Koopman, J.F.L., Lotze-Campen, H., Mason-D'Croz, D., Ochi, Y., Pérez Domínguez, I., Stehfest, E., Sulser, T.B., Tabeau, A., Takahashi, K., Takakura, J., van Meijl, H., van Zeist, W.-J., Wiebe, K., Witzke, P., 2018. Risk of increased food insecurity under stringent global climate change mitigation policy. Nat. Clim. Chang 8, 699–703. https://doi.org/10.1038/s41558-018-0230-x.
- Hay, W.T., Bihmidine, S., Mutlu, N., Hoang, K., le, Awada, T., Weeks, D.P., Clemente, T. E., Long, S.P., 2017. Enhancing soybean photosynthetic CO₂ assimilation using a cyanobacterial membrane protein, ictB. J. Plant Physiol. 212, 58–68. https://doi. org/10.1016/j.jplph.2017.02.003.
- He, L., Jin, N., Yu, Q., 2020. Impacts of climate change and crop management practices on soybean phenology changes in China. Sci. Total Environ. 707, 135638 https:// doi.org/10.1016/j.scitotenv.2019.135638.
- Joos, F., Prentice, I.C., Sitch, S., Meyer, R., Hooss, G., Plattner, G.-K., Gerber, S., Hasselmann, K., 2001. Global warming feedbacks on terrestrial carbon uptake under the intergovernmental panel on climate change (IPCC) emission scenarios. Glob. Biogeochem. Cycles 15, 891–907. https://doi.org/10.1029/2000GB001375.
- Koester, R.P., Skoneczka, J.A., Cary, T.R., Diers, B.W., Ainsworth, E.A., 2014. Historical gains in soybean (Glycine max Merr.) seed yield are driven by linear increases in light interception, energy conversion, and partitioning efficiencies. J. Exp. Bot. 65, 3311–3321. https://doi.org/10.1093/jtb/eru187.
- Köhler, I.H., Ruiz-Vera, U.M., VanLoocke, A., Thomey, M.L., Clemente, T., Long, S.P., Ort, D.R., Bernacchi, C.J., 2016. Expression of cyanobacterial FBP/SBPase in soybean prevents yield depression under future climate conditions. erw435 J. Exp. Bot.. https://doi.org/10.1093/jtxb/erw435.
- Ku, M.S.B., Ranade, U., Hsu, T.-P., Cho, D., Li, X., Jiao, D.-M., Ehleringer, J., Miyao, M., Matsuoka, M., 2000. Photosynthetic performance of transgenic rice plants overexpressing maize C4 photosynthesis enzymes. In: Studies in Plant Science. Elsevier, pp. 193–204. https://doi.org/10.1016/S0928-3420(00)80015-4.
- Kumarathunge, D.P., Medlyn, B.E., Drake, J.E., Rogers, A., Tjoelker, M.G., 2019. No evidence for triose phosphate limitation of light-saturated leaf photosynthesis under current atmospheric CO 2 concentration. Plant Cell Environ. 42, 3241–3252. https://doi.org/10.1111/pce.13639.
- Leakey, A.D.B., Ainsworth, E.A., Bernacchi, C.J., Rogers, A., Long, S.P., Ort, D.R., 2009a. Elevated CO₂ effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. J. Exp. Bot. 60, 2859–2876. https://doi.org/10.1093/jxb/ erp096.
- Leakey, A.D.B., Xu, F., Gillespie, K.M., McGrath, J.M., Ainsworth, E.A., Ort, D.R., 2009b. Genomic basis for stimulated respiration by plants growing under elevated carbon dioxide. Proc. Natl. Acad. Sci. 106, 3597–3602. https://doi.org/10.1073/ pnas.0810955106.
- Lefebvre, S., Lawson, T., Fryer, M., Zakhleniuk, O. v, Lloyd, J.C., Raines, C.A., 2005. Increased Sedoheptulose-1,7-Bisphosphatase Activity in Transgenic Tobacco Plants Stimulates Photosynthesis and Growth from an Early Stage in Development. Plant Physiol. 138, 451–460. https://doi.org/10.1104/pp.104.055046.

- Lochocki, E.B., Rohde, S., Jaiswal, D., Matthews, M.L., Miguez, F., Long, S.P., McGrath, J. M., 2022. BioCro II: a software package for modular crop growth simulations. Silico Plants 4. https://doi.org/10.1093/insilicoplants/diac003.
- Long, S.P., Ainsworth, E.A., Rogers, A., Ort, D.R., 2004. Rising atmospheric carbon dioxide: plants face the future. Annu Rev. Plant Biol. 55, 591–628. https://doi.org/ 10.1146/annurev.arplant.55.031903.141610.
- Long, S.P., Zhu, X.-G., Naidu, S.L., Ort, D.R., 2006. Can improvement in photosynthesis increase crop yields? Plant Cell Environ. 29, 315–330. https://doi.org/10.1111/ j.1365-3040.2005.01493.x.
- Long, S.P., Marshall-Colon, A., Zhu, X.-G., 2015. Meeting the global food demand of the future by engineering crop photosynthesis and yield potential. Cell 161, 56–66. https://doi.org/10.1016/j.cell.2015.03.019.
- López-Calcagno, P.E., Brown, K.L., Simkin, A.J., Fisk, S.J., Vialet-Chabrand, S., Lawson, T., Raines, C.A., 2020. Stimulating photosynthetic processes increases productivity and water-use efficiency in the field. Nat. Plants 6, 1054–1063. https:// doi.org/10.1038/s41477-020-0740-1.
- Matthews, M.L., Marshall-Colón, A., McGrath, J.M., Lochocki, E.B., Long, S.P., 2022. Soybean-BioCro: a semi-mechanistic model of soybean growth. Silico Plants 4. https://doi.org/10.1093/insilicoplants/diab032.
- Medrano, H., Tomás, M., Martorell, S., Flexas, J., Hernández, E., Rosselló, J., Pou, A., Escalona, J.-M., Bota, J., 2015. From leaf to whole-plant water use efficiency (WUE) in complex canopies: limitations of leaf WUE as a selection target. Crop J. 3, 220–228. https://doi.org/10.1016/j.cj.2015.04.002.
- Morgan, P.B., Bollero, G.A., Nelson, R.L., Dohleman, F.G., Long, S.P., 2005. Smaller than predicted increase in aboveground net primary production and yield of field-grown soybean under fully open-air [CO₂] elevation. Glob. Chang Biol. 11, 1856–1865. https://doi.org/10.1111/j.1365-2486.2005.001017.x.
- Ort, D.R., Merchant, S.S., Alric, J., Barkan, A., Blankenship, R.E., Bock, R., Croce, R., Hanson, M.R., Hibberd, J.M., Long, S.P., Moore, T.A., Moroney, J., Niyogi, K.K., Parry, M.A.J., Peralta-Yahya, P.P., Prince, R.C., Redding, K.E., Spalding, M.H., van Wijk, K.J., Vermaas, W.F.J., von Caemmerer, S., Weber, A.P.M., Yeates, T.O., Yuan, J.S., Zhu, X.G., 2015. Redesigning photosynthesis to sustainably meet global food and bioenergy demand. Proc. Natl. Acad. Sci. 112, 8529–8536. https://doi.org/ 10.1073/pnas.1424031112.
- Parry, M.A.J., Reynolds, M., Salvucci, M.E., Raines, C., Andralojc, P.J., Zhu, X.-G., Price, G.D., Condon, A.G., Furbank, R.T., 2011. Raising yield potential of wheat. II. Increasing photosynthetic capacity and efficiency. J. Exp. Bot. 62, 453–467. https:// doi.org/10.1093/jxb/erq304.
- Raines, C.A., 2011. Increasing photosynthetic carbon assimilation in C3 plants to improve crop yield: current and future strategies. Plant Physiol. 155, 36–42. https:// doi.org/10.1104/pp.110.168559.
- Rosenthal, D.M., Locke, A.M., Khozaei, M., Raines, C.A., Long, S.P., Ort, D.R., 2011. Over-expressing the C3 photosynthesis cycle enzyme Sedoheptulose-1-7 Bisphosphatase improves photosynthetic carbon gain and yield under fully open air CO2funigation (FACE). BMC Plant Biol. 11, 123. https://doi.org/10.1186/1471-2229-11-123.
- Salesse-Smith, C.E., Sharwood, R.E., Busch, F.A., Kromdijk, J., Bardal, V., Stern, D.B., 2018. Overexpression of Rubisco subunits with RAF1 increases Rubisco content in maize. Nat. Plants 4, 802–810. https://doi.org/10.1038/s41477-018-0252-4.
- Sharkey, T.D., 2019. Is triose phosphate utilization important for understanding photosynthesis? J. Exp. Bot. 70, 5521–5525. https://doi.org/10.1093/jxb/erz393.
- Simkin, A.J., McAusland, L., Headland, L.R., Lawson, T., Raines, C.A., 2015. Multigene manipulation of photosynthetic carbon assimilation increases CO₂ fixation and biomass yield in tobacco. J. Exp. Bot. 66, 4075–4090. https://doi.org/10.1093/jxb/ erv204.

- Simkin, A.J., López-Calcagno, P.E., Raines, C.A., 2019. Feeding the world: improving photosynthetic efficiency for sustainable crop production. J. Exp. Bot. 70, 1119–1140. https://doi.org/10.1093/jxb/ery445.
- Sims, D.A., Luo, Y., Seemann, J.R., 1998. Comparison of photosynthetic acclimation to elevated CO₂ and limited nitrogen supply in soybean. Plant Cell Environ. 21, 945–952. https://doi.org/10.1046/j.1365-3040.1998.00334.x.
- Sinclair, T.R., Wit, C.T., 1976. Analysis of the carbon and nitrogen limitations to soybean yield. Agron. J. 68, 319–324. https://doi.org/10.2134/ agroni1976.00021962006800020021x.
- Sinclair, T.R., Purcell, L.C., Sneller, C.H., 2004. Crop transformation and the challenge to increase yield potential. Trends Plant Sci. 9, 70–75. https://doi.org/10.1016/j. tplants.2003.12.008.
- Specht, J.E., Hume, D.J., Kumudini, S., 1999. Soybean yield potential-a genetic and physiological perspective. Crop Sci. v (39), 1560–1570. https://doi.org/10.2135/ cropsci1999.3961560x.
- Srinivasan, V., Kumar, P., Long, S.P., 2017. Decreasing, not increasing, leaf area will raise crop yields under global atmospheric change. Glob. Chang Biol. 23, 1626–1635. https://doi.org/10.1111/gcb.13526.
- Stevenson, M., Sergeant, E., Nunes, T., Heuer, C., Marshall, J., Sanchez, J., Thornton, R., Reiczigel, J., Robison-Cox, J., Sebastiani, P., Solymos, P., Yoshida, K., Jones, G., Pirikahu, S., Firestone, S., Kyle, R., Popp, J., Jay, M., Reynard, C., Cheung, A., Singanallur, N., Szabo, A., Rabiee., A., 2022. epiR: Tools for the Analysis of Epidemiological Data.
- Suzuki, Y., Miyamoto, T., Yoshizawa, R., Mae, T., Makino, A., 2009. Rubisco content and photosynthesis of leaves at different positions in transgenic rice with an overexpression of RBCS. Plant Cell Environ. 32, 417–427. https://doi.org/10.1111/ j.1365-3040.2009.01937.x.
- Tagliapietra, E.L., Streck, N.A., Rocha, T.S.M., Richter, G.L., Silva, M.R., Cera, J.C., Guedes, J.V.C., Zanon, A.J., 2018. Optimum leaf area index to reach soybean yield potential in subtropical environment. Agron. J. 110, 932–938. https://doi.org/ 10.2134/agronj2017.09.0523.
- Wise, R.R., Olson, A.J., Schrader, S.M., Sharkey, T.D., 2004. Electron transport is the functional limitation of photosynthesis in field-grown Pima cotton plants at high temperature. Plant Cell Environ. 27, 717–724. https://doi.org/10.1111/j.1365-3040.2004.01171.x.
- Wu, A., Doherty, A., Farquhar, G.D., Hammer, G.L., 2018. Simulating daily field crop canopy photosynthesis: an integrated software package. Funct. Plant Biol. 45, 362. https://doi.org/10.1071/FP17225.
- Wu, A., Hammer, G.L., Doherty, A., von Caemmerer, S., Farquhar, G.D., 2019. Quantifying impacts of enhancing photosynthesis on crop yield. Nat. Plants 5, 380–388. https://doi.org/10.1038/s41477-019-0398-8.
- Wu, A., Brider, J., Busch, F.A., Chen, M., Chenu, K., Clarke, V.C., Collins, B., Ermakova, M., Evans, J.R., Farquhar, G.D., Forster, B., Furbank, R.T., Groszmann, M., Hernandez-Prieto, M.A., Long, B.M., Mclean, G., Potgieter, A., Price, G.D., Sharwood, R.E., Stower, M., van Oosterom, E., von Caemmerer, S., Whitney, S.M., Hammer, G.L., 2023. A cross-scale analysis to understand and quantify the effects of photosynthetic enhancement on crop growth and yield across environments. Plant Cell Environ. 46, 23–44. https://doi.org/10.1111/pce.14453.
- Yin, X., Gu, J., Dingkuhn, M., Struik, P.C., 2022. A model-guided holistic review of exploiting natural variation of photosynthesis traits in crop improvement. J. Exp. Bot. 73, 3173–3188. https://doi.org/10.1093/jxb/erac109.
- Zhu, X.-G., Long, S.P., Ort, D.R., 2008. What is the maximum efficiency with which photosynthesis can convert solar energy into biomass? Curr. Opin. Biotechnol. 19, 153–159. https://doi.org/10.1016/j.copbio.2008.02.004.