



## Review



**Cite this article:** Long SP. 2025 Needs and opportunities to future-proof crops and the use of crop systems to mitigate atmospheric change.

*Phil. Trans. R. Soc. B* **380**: 20240229.

<https://doi.org/10.1098/rstb.2024.0229>

Received: 12 January 2025

Accepted: 22 January 2025

One contribution of 21 to a theme issue ‘Crops under stress: can we mitigate the impacts of climate change on agriculture and launch the ‘Resilience Revolution’?’.

### Subject Areas:

plant science

### Keywords:

food security, rising CO<sub>2</sub>, drought, flooding, *Miscanthus*, rising temperature

### Author for correspondence:

Stephen P. Long

e-mail: [slong@illinois.edu](mailto:slong@illinois.edu)

# Needs and opportunities to future-proof crops and the use of crop systems to mitigate atmospheric change

Stephen P. Long

Institute for Genomic Biology and Departments of Plant Biology and of Crop Sciences, University of Illinois at Urbana-Champaign, Urbana, IL 61801, USA

SPL, 0000-0002-8501-7164

Predicted changes in atmospheric composition and climate affecting crop productivity are reviewed. These include changes in both average conditions and extreme events, with respect to temperature, drought, flooding and surface ozone, coupled with rising atmospheric [CO<sub>2</sub>]. Impacts on, and means to adapt, crops to these changes are reviewed and outlined. Particular emphasis is given to (i) the results from open air field manipulations of surface atmosphere, temperature and soil water to understand impacts and adaptation and (ii) demonstrated genetic manipulations of photosynthesis and water use that could support future food supply under current and future conditions. Finally, attention is given to means by which crop systems could serve as CO<sub>2</sub> collectors and carbon storage systems. Here, apparent opportunities are outlined for (i) manipulations of crops to enhance carbon storage and (ii) use of high-productivity sustainable perennial C<sub>4</sub> grasses coupled with carbon capture and storage.

This article is part of the theme issue ‘Crops under stress: can we mitigate the impacts of climate change on agriculture and launch the ‘Resilience Revolution’?’.

*U.S. Secretary of State Anthony Blinken at the Global Solutions for Food Security Event in New York in September 2023 ‘if we don’t get this right, I actually don’t think anything else really, really matters’ [1].*

## 1. Predicted future needs

Secretary Blinken’s remark truly underlines the gravity of our current situation and the seriousness of failing to address future food security. Recent and past history shows us that food shortages result in sharp price rises for any available food, leading to high levels of starvation, disruption of the social and political order of a country, and mass migrations [2–8]. By 2050–60 crops will experience a significantly different environment from today. Depending on location, a few changes will benefit crops, but the majority will lead to yield losses unless crops are future-proofed to anticipated change. Atmospheric [CO<sub>2</sub>] reached 427 p.p.m. in 2024 and is projected to be approximately 600 p.p.m. by 2050–60 [9,10]. This assumes the ‘representative concentration pathway’ (RCP) 8.5 of continued high emissions. With lower emissions, the concentration increase would of course be lower. However, actual increases have tracked RCP 8.5 for the past 20 years, with little sign of deviation [10]. This is underlined by the fact that the period from 2022 to 2024 saw the largest two-year jump in atmospheric [CO<sub>2</sub>] of any time since records began [9]. In

June 2023, the average global temperature relative to the pre-industrial level, exceeded 1.5°C, and it remains above 1.5°C [11,12]. Yet, 1.5°C was the limit of warming in the Paris Agreement, a legally binding international treaty on climate change. It was adopted in 2016 by 194 individual countries, plus the EU, following the UN Climate Change Conference (COP21) [13]. Its failure, within just seven years from signing, signals a clear lack of commitment to addressing the problem with any urgency. Given the apparent absence of sufficient political will, RCP 8.5 (SSSP5-8.5 in the IPCC AR6 report [10]) is used here to address what may well come to pass by mid-century. While we may hope for a better scenario, future-proofing of cropping systems should prepare for the worst, as essential insurance.

Under this scenario, the global average temperature will rise another 1.2°C by 2050–60 to 2.7°C above pre-industrial temperature. This will be accompanied by more extreme temperature events. For example, in eastern Europe, the number of days in which 35°C is exceeded is predicted to quadruple from around 10 days in 2023 to 40 days. In the Caribbean, the number of days in which the temperature exceeds 41°C could rise from around 50 to 200 [10]. Precipitation will increase as a global average by 6%, but this will vary greatly between regions. It will very likely increase at high latitudes and in monsoon climates and likely decrease over large parts of the subtropics [10]. However, increased temperature and surface to atmosphere water vapour pressure deficit (VPD) will increase evapotranspiration and lower water use efficiency (WUE) [14]. Therefore, despite more precipitation, annual mean total column soil moisture is predicted to decline in most of South America, the central plains of North America, southern Africa, the Mediterranean and much of China and Australia, while increasing in equatorial Africa, central Asia and most high latitude regions. Precipitation on the wettest day of the year and hence the likelihood of flooding and run-off are projected to increase in all locations by 10–40%, with the current one-in-ten-year heavy rainfall event becoming a one-in-three-year event and 30% more intense under the SSSP5-8.5 scenario [10].

Why is future-proofing crop production against these changes so important? In the absence of climate change, a meta-analysis of 57 detailed global food scenario quantitative projections showed that the world will need between 35 and 56% more food by 2050 [15]. This is driven by three global trends: increased food wastage as the proportion of the global population living in urban areas increases, increased consumption *per capita* of meat and dairy and a rising global population [16]. If steady increases in temperature and drought are added, the maximum projected demand rises up to 62% [15]. However, the increase in crop losses to extreme events (fires, heat waves, floods and extreme droughts) raises the production capacity needed to provide sufficient food and reserves by a further 15–20% [17]. These extreme events will cause regional food supply shocks increasingly as climate change progresses, requiring increased food reserves as a buffer against starvation in the affected areas [17,18]. In short, to provide sufficient food and reserves, it may be necessary to almost double the yield per unit of land in current use. If supply fails to meet demand, then expansion of the crop production footprint and more destruction of natural habitats—in particular tropical forests—is an inevitable consequence [19,20]. This would further exacerbate climate change and the current rapid rate of biodiversity loss [21]. The number going hungry rose from 541 million in 2017 to 731 million in 2023 [22], a trend that can only continue as growth in demand exceeds production. The following sections outline the impacts and potential mitigation of specific aspects of change in atmospheric composition and climate on cropping systems.

## 2. Rising atmospheric [CO<sub>2</sub>]

Open-air elevation of [CO<sub>2</sub>] using free-air concentration enrichment (FACE) technology has provided us with the most realistic data on the direct effects of future [CO<sub>2</sub>] levels on crops [23]. In the absence of other changes and stresses, yields of C<sub>3</sub> crops are substantially increased, with modern elite cultivars of rice and soybean showing yield increases of approximately 30% with elevation of [CO<sub>2</sub>] to anticipated 2050–60 levels. C<sub>4</sub> crops—maize and sorghum—do not show a yield increase, since they are already [CO<sub>2</sub>]-saturated at today's already elevated [CO<sub>2</sub>] levels [24]. However, under drought conditions yields can be increased, since stomata of both C<sub>3</sub> and C<sub>4</sub> show reduced aperture and conductance in elevated [CO<sub>2</sub>], resulting in improved conservation of soil water [24]. Elevated [CO<sub>2</sub>] does not always benefit crop production in a drought environment. In years when soil moisture is high during crop establishment, elevated [CO<sub>2</sub>] can lead to poorer root development, which then impacts the crop if soil moisture is depleted during the reproductive stage [25]. While yield may be increased, quality is generally depressed, particularly the protein content of non-legumes [26]. However, there is wide variation in protein and mineral content within the germplasm of the major grain crops, suggesting that breeding could quite easily address this quality loss [24].

Atmospheric [CO<sub>2</sub>] averaged 200 p.p.m. during the 2 Myr prior to the Industrial Revolution, the period during which our crops' ancestors evolved [27]. In November 2024, the level had reached 424 p.p.m., more than double the level of 2 Myr ago [9,27]. Half of this increase has occurred in just the past 50 years, a very short period for natural or breeder selection to afford adaptation to these elevated conditions [9]. Analysis of limitations to photosynthesis suggests that metabolic control has been altered by this elevation. In C<sub>3</sub> crops, control has shifted to become predominantly ribulose-1:5-bisphosphate (RuBP) regeneration-limited from RuBP carboxylase-oxygenase (Rubisco) limitation at lower [CO<sub>2</sub>] levels [28]. This indicated that upregulation of proteins limiting RuBP regeneration could further increase yields under elevated [CO<sub>2</sub>]. Plastid sedoheptulose-1:7-bisphosphatase (SbPase) is one enzyme that has been shown to limit RuBP regeneration [29]. Consistent with this theory, transgenic upregulation of SbPase in both tobacco and soybean resulted in significantly greater increases in photosynthesis and productivity in FACE than under current atmospheric conditions [30,31]. In contrast, the elevation of [CO<sub>2</sub>] that has already occurred has shifted the metabolic control of photosynthetic CO<sub>2</sub> assimilation in C<sub>4</sub> crops from phospho-enol pyruvate carboxylase (PEPC) to Rubisco [32]. This inferred that the elevation of Rubisco would increase photosynthesis and productivity in C<sub>4</sub> crops. Consistent with this theory, transgenic upregulation of Rubisco in maize, sorghum and sugarcane resulted in increased photosynthesis at current [CO<sub>2</sub>], but not at pre-industrial levels. This also resulted in increased productivity in greenhouse conditions and—for sorghum—in replicated four-row plot field trials [16,33,34]. While these modifications that have increased productivity of both

C<sub>3</sub> and C<sub>4</sub> crops under current and future conditions of elevated [CO<sub>2</sub>], they involved transgenesis. However, it is very likely that the same increases could be achieved by editing the promoter region of these genes, potentially avoiding or minimizing lengthy deregulation of such improved germplasm [16,34,35].

### 3. Tropospheric ozone

Tropospheric ozone (O<sub>3</sub>) is a secondary pollutant formed by the action of sunlight on volatile organic compounds and nitrogen oxides in polluted air masses. As these air masses drift into the countryside from the large urban areas in which they are formed, they continue to produce ozone in the presence of sunlight. These masses can drift across entire continents and even between continents. While reductions in fossil fuel use might lessen the problem, this will be likely counteracted by rising temperatures which enhance photochemical ozone formation. [O<sub>3</sub>] levels of ≥40 ppb can be damaging to crop productivity [36]. Pre-industrial levels are considered to have been about 5–15 ppb [37]. Today, levels of >100 ppb can be frequently found in rural areas of the US corn belt, with significantly higher levels in the major crop production areas of China and India [38,39]. A regression analysis of historical yield, climate and [O<sub>3</sub>] data for the USA was used to determine the losses resulting from [O<sub>3</sub>] for maize (*Zea mays*) and soybean (*Glycine max*) between 1980 and 2011. In rain-fed fields, this showed a loss owing to ozone of approximately 5% for soybean and approximately 10% for maize, costing some \$9 billion annually [40]. Globally, losses of the major food crops could be in the region of 10% and rising [39]. Overcoming this would provide a substantial piece of the increased food supply needed. A key question is how future conditions of higher temperatures and drought will interact with O<sub>3</sub> damage. While higher temperatures and drought generally coincide with higher surface [O<sub>3</sub>] levels, a 15 year open-air fumigation study showed that drought and elevated temperatures did not alter the vulnerability of soybean to a given level of ozone [41].

Ozone effects its damage on the plant by entering the leaf via the stomata, where it dissolves into the apoplastic fluid, producing an array of damaging active oxygen species [42]. An early downstream response is the loss of Rubisco [43]. Rising [CO<sub>2</sub>] means that leaves could assimilate the same amount of CO<sub>2</sub> with lower stomatal conductance. Engineering or selecting for lower stomatal conductance would also decrease O<sub>3</sub> uptake and damage, while upregulating Rubisco content could also lessen its impact. Antioxidant systems can remove the damaging oxidizing radicals produced from O<sub>3</sub> absorption in the leaf [43]. Upregulation of these would also lessen ozone damage. Studies of the impacts of O<sub>3</sub> on a range of maize, wheat and rice collections have revealed considerable variation in O<sub>3</sub> tolerance and enabled the identification of loci associated with ozone tolerance and susceptibility [44–46]. This suggests a means for marker-assisted introgression of alleles conferring O<sub>3</sub> tolerance to these major food crops.

### 4. Rising temperature

Rising temperatures will make conditions better or worse for crop production, depending on location. In the cool temperate zone, more frost-free days allow a longer growing season, facilitating the planting of longer season maturity types or allowing double cropping, such as winter cereal followed by soybean or a spring cereal within a single year. It will also allow the cultivation of crops at ever-higher latitudes, provided those locations have adequate soil quality for these crops. However, higher temperatures may also benefit weeds, pests and diseases. In addition, while growth and development may be accelerated, higher temperatures will reduce the length of the grain/seed-filling window, leading to lower yields. Crops that require insect pollination will suffer if rising temperatures cause a disparity between flowering and the pollinators' period of abundance. On balance, with a further 1.2°C temperature rise by mid-century, yields are projected to be reduced by 9% (wheat), 16% (rice) or 5% (soybean) for current cultivars. In the tropics and warm temperate zone, increased frequency of extreme heat will likely increase incidence of crop failures, particularly when combined with drought [17]. The frequency of years when temperatures exceed thresholds for damage during critical growth stages will increase in all regions.

Ambient temperatures have a profound effect on plant growth, influencing all stages of development from germination and shoot growth to flower, fruit and seed formation, affecting both crop quality and yield. Higher temperatures can impair crop yields by decreasing photosynthesis, increasing respiration, and affecting development, pollination, fertilization and fruit/seed development. Higher temperatures will also mean higher leaf to air VPDs. The water vapour saturation pressure of air rises exponentially with temperature. Very few studies of elevated temperature have controlled VPD and the drying stress that it drives. As such, it is impossible in most cases to separate whether damage is a direct effect of temperature or indirect due to increased VPD [47].

In the absence of other stresses, the optimum temperature for crop photosynthesis is typically about 25°C for C<sub>3</sub> crops and 35°C for C<sub>4</sub> crops. Increases in temperature beyond these optima result in losses in photosynthesis, with a steep decline when the optimum is exceeded by about 10°C. In maize, photosynthesis declines by 50–60% at 40°C [48]. Across crops, this has been associated with a sharp decline in the activity of Rubisco activase (Rca), which serves to remove inhibitors from Rubisco. Clearly, there are wild species that can tolerate considerably higher temperatures, such as desert plants, and they may prove a source of more thermotolerant forms or inform edits that could improve the thermostability of Rca. Even within the germplasm of individual crops, variation in Rca temperature tolerance has been identified, including single residue changes that would improve thermal stability [49–51]. Small heat-shock proteins (HSPs), which serve to protect photosynthetic proteins, are induced by high temperatures. Upregulation of these HSPs may increase thermal tolerance of photosynthesis and other physiological processes [52].

Crops are generally more sensitive to elevated temperature during reproductive growth. Temperatures in excess of 35–39°C during this phase of growth generally lead to large or complete crop losses. In maize at  $\geq 30^\circ\text{C}$ , endosperm cell and amyloplast division are slowed, affecting sink strength and kernel size [53]. Pollen viability decreases at  $\geq 35^\circ\text{C}$ . Duration of pollen viability after anthesis is strongly dependent on moisture content, which in turn is strongly affected by the large elevation of VPD at these high temperatures. The result is that pollen viability will be low or absent by the time silking occurs [48]. While vegetative growth in soybean has an optimum temperature of  $30^\circ\text{C}$ , the optimum temperature for reproductive growth and development post-anthesis is just  $23^\circ\text{C}$ . An increase in the mean temperature above  $23^\circ\text{C}$  causes declines in seed growth rate, seed size and partitioning of biomass to grain (harvest index), reaching zero at  $39^\circ\text{C}$ . The viability of soybean pollen begins to decline at  $30^\circ\text{C}$ , with complete failure at  $47^\circ\text{C}$  [48]. Significant variation in pollen viability and pollen tube development was found in a survey of 44 genotypes [54], suggesting that with a wider search of germplasm, including the wild ancestor *Glycine soja* whose distribution extends from Siberia to sub-tropical China [55], better tolerance could be found. Rice similarly shows an optimum for vegetative growth of  $33^\circ\text{C}$ , but  $25^\circ\text{C}$  for grain development. The yield declines by about 10% for each  $1^\circ\text{C}$  above  $25^\circ\text{C}$  until  $35^\circ\text{C}$ , where yield is zero. This is associated with reduced pollen viability and production at temperatures above  $25^\circ\text{C}$  [48]. Significant genotypic variations in heat tolerance for percent filled grains, pollen production, pollen shed and pollen viability were found respectively across 14, 3 and 4 rice genotypes/ecotypes for heat tolerance, with greater pollen heat tolerance found in upland genotypes [56–58]. This suggests that a much wider search coupled with genome-wide association analysis would likely uncover considerable variation and associated loci that may be used in increasing reproductive viability at elevated temperatures. Of these major food crops, wheat shows the lowest optimum temperature for grain development, at just  $15^\circ\text{C}$ . This is primarily owing to the shortening of the grain filling window as temperatures increase, even in the absence of any reduction in photosynthesis. The increase in mean temperature during grain filling by each degree Celsius above  $15^\circ\text{C}$  lowers wheat yield by 6% [59]. A survey of 304 diverse elite winter wheat showed significant variation in heat stress tolerance of grain filling, identifying lines of potential value for breeding future-proofed wheat [60].

## 5. Drought and water use efficiency

By 2050, global yield losses to drought in maize are projected to rise to 21.3% from a previous average of 12.0% for the period 1961–2006, and for wheat from 9.6% to 15.5%. Of the areas of current crop production, the proportion that are drought-affected will rise most in Africa and Oceania, from the present 22 and 15%, respectively, to 59 and 58% by the end of the century [61]. Considerable effort was expended in identifying potential drought tolerance genes through screening of *Arabidopsis* mutants and relatives, which greatly expanded our understanding of the molecular responses of this plant to drought [62–64]. However, these have not translated into increased drought tolerance in crops, to date. The introduction of the *Bacillus subtilis* cold shock protein B (cspB), which downregulates ethylene production in drought, into maize appears to have improved drought tolerance and yield [65]. Two years of trials in Nigeria showed a significant yield increase under moderate drought conditions [66]. RNAi suppression of farnesyltransferase increased the Absciscic acid (ABA) sensitivity of guard cells and in turn, the drought tolerance of canola (*Brassica napus*), improving yields by 26% in field trials [65,67]. Despite these advances and other promising biotechnological approaches [65], these innovations have not been widely deployed.

Drought tolerance could also be improved by increasing crop WUE. Higher WUE results in a crop depleting soil water reserves more slowly, which can increase survival of transient droughts. Globally, 71.3% of freshwater withdrawals in 2020 were used in agriculture, primarily for irrigation. Despite declining freshwater availability, the proportion allocated to agriculture continues to rise. For example, India's freshwater use in agriculture doubled between 1975 and 2010 as its population and food demand rose. This trend has continued: 32% of agricultural land in India was irrigated in 2001, rising to 42% in 2020 [68]. In 2020, 21% of global cropland was irrigated and this land accounts for about 40% of total food production [68,69]. Even without any increase in future food demand, more currently rain-fed land would need to be brought into irrigation to maintain current yields. This is because rising VPD will lower WUE [14,47]. Given our dependence on irrigated land, it is hard to see how a 60%+ increase in food supply could be achieved within existing water resources. Solutions will be in more efficient use of irrigation water and bioengineering/breeding of more water use-efficient crops.

One opportunity results from rising  $[\text{CO}_2]$ : as  $[\text{CO}_2]$  rises, the same rates of assimilation can be achieved with lower stomatal conductance. Plants do respond to increased  $[\text{CO}_2]$  with partial stomatal closure, but this is insufficient to take full advantage of the elevated  $[\text{CO}_2]$  in the atmosphere. This is particularly the case for  $\text{C}_4$  crops, where photosynthesis is already  $[\text{CO}_2]$  saturated in the current atmosphere [16]. We therefore need to engineer or breed for decreased stomatal conductance to obtain the improved WUE that can be achieved under rising  $[\text{CO}_2]$ . Over-expression of Photosystem II Subunit S (PsbS) lowered the redox state of  $\text{Q}_\text{a}$ , the primary electron acceptor of Photosystem II, at all light levels, without affecting the  $\text{CO}_2$  assimilation rate but lowering stomatal conductance. The result was a 15% improvement in leaf-level WUE in field-grown tobacco and a 30% decrease in whole plant water use [70,71]. An alternative approach to affecting stomatal behaviour is to reduce stomatal density (SD). Elucidation of the genes controlling steps in stomatal development has allowed targeted transformation and edits to decrease SD [72]. Moderate reductions ( $<50\%$ ) in SD by transgenic expression of epidermal patterning factor (EPF) genes in both rice and wheat improved WUE by approximately 15–20% without affecting yield [73,74]. Similarly, moderate reductions in SD by constitutive expression of a synthetic EPF transgene in  $\text{C}_4$  sorghum lowered plant water use by approximately 15% without affecting  $\text{CO}_2$  uptake [75]. However, substantial variation in SD across the germplasm of different crops is likely. For example, SD varied 2.5-fold across 235 rice accessions [76]. By combining optical topometry and machine learning, a high-throughput phenotyping method has become available to rapidly screen large amounts of germplasm for variation in SD and other morphological properties of the stomatal apparatus. This will hugely shorten the time needed to screen large



collections of germplasm and would also aid phenotyping in the introgression of lower SD into elite cultivars [77]. Another factor affecting WUE is the speed of adjustment of stomatal conductance to fluctuations in light. Within crop canopies light is continually fluctuating; on transfer of leaves from sun to shade, CO<sub>2</sub> assimilation drops abruptly, but the stomata require many minutes to adjust to the greatly reduced flux. As a result, leaf WUE is low during this period of adjustment. Across the leaf canopy and the day, this amounts to a considerable lowering of crop WUE in the field environment [78–80]. There is again considerable variation in the speed of this adjustment within the germplasm of individual crops: for example, WUE during a light transient across 15 contrasting accessions of rice differed twofold [81].

## 6. Flooding

Ironically, just as the frequency, duration and intensity of drought events are predicted to increase, so is flooding. After droughts, flooding is the second largest cause of crop losses, accounting for 19% of global losses between 2008 and 2018 [82]. Flooding affects crop production by preventing planting or by causing plant mortality after planting by starving the roots of oxygen. Water itself is of course not toxic, but diffusion of gases—including oxygen—is some 10 000 times slower in water than in air, leading to rapid depletion of oxygen in flooded soils. Therefore, on flooding, soils rapidly become anaerobic, i.e. have insufficient oxygen for aerobic respiration. In the absence of oxygen as the terminal acceptor of mitochondrial electron transport, ATP supply is greatly diminished. In addition, soil bacteria seeking alternative electron acceptors will reduce many ions to forms that are toxic to the roots. For example, sulfates and sulfites are reduced to hydrogen sulfide, which is highly toxic to plants, although it is also a key signalling molecule for acclimation at low concentrations [83]. Wetland plants, including rice, avoid these problems by producing roots with aerenchyma, which are large air spaces within the root cortex that permit oxygen from the shoot to reach the roots. This allows them to function normally and produce an encasing oxidized soil layer that will re-oxidize toxic ions formed under the anaerobic conditions of the surrounding soil. Could this strategy be incorporated into the other major crops? A survey of the root anatomy of 256 different maize cultivars and relatives showed a tenfold variation in the proportion of root aerenchyma. Germplasm in which aerenchyma occupied more than one-quarter of the cross-sectional area, equivalent to that found in marsh plants, has been identified [84]. Defence against hypoxia on flooding of soils can also be induced. This shows considerable potential for breeding tolerance to flooding of soils. Hypoxia triggers ethylene biosynthesis, which acts as a signal to promote both antioxidant activity and aerenchyma formation [85]. However, in most current crop cultivars this is insufficient to avoid damage and loss. Understanding of the molecular basis of adaptive plant responses to soil hypoxia has been greatly advanced in recent years [86,87], opening an opportunity to upregulate these responses to protect roots during flooding. Root aerenchyma does not help if flooding is so severe that the shoot is also submerged and starved of oxygen. This can be a frequent occurrence for rice, where 35% of land planted to rice is prone to floods that can submerge the developing crop for days or weeks [88]. Here, however, remarkable progress has already been made. Surveying the ability of rice cultivars to survive up to two weeks of complete submergence, a tolerant cultivar was discovered and associated with an allele *Sub1A-1* at the *Submergence 1* locus. This gene is rapidly induced on submergence in contrast to other alleles, and allows the plant to survive. Subsequent introgression of *Sub1A-1* into flooding intolerant cultivars made them flooding tolerant [89]. This discovery then allowed marker-assisted introgression of *Sub1A-1* into a wide range of germplasm, particularly aiding poor farmers in the most flood-prone areas, who rapidly adopted the flood-tolerant lines [90].

Most potential solutions, as outlined above, have concerned single key variables of global change. Attention must also be given to atmospheric and climatic changes that include a combination of two or more stress factors, such as drought and heat. These could subject plants to complex conditions of stress combination that require additional molecular discovery and breeding efforts [24,25,41,91–93].

## 7. Modifying crops for carbon sequestration

The annual net flux of carbon dioxide into the terrestrial biosphere was estimated for the period 2010–19 to be 5.3 Gt (C) [10]. Arable crops occupy 1.7 billion acres worldwide. Today, for our major crops, about 50% of the above-ground biomass is harvested as grain or seed, and the remaining 50%—stem, leaves, chaff and other waste—can remain on the field and may be incorporated into the soil. A further significant fraction of biomass is root, which remains within the soil. Maize, rice, soy and wheat are the top four arable crops in terms of land area occupied: 721 Mha in 2021. Their aggregate average yield was 4.3 t ha<sup>-1</sup> in 2021 [94]. If we assume that the same amount of mass remained on this land area after harvest, plus another 50% as roots, then this would amount to 2.7 Gt (C) for just these four crops, assuming C is 40% of the dry biomass. Of course, this has little value as a carbon sink if this residue is decomposed within a few years, but if it could be kept in the soil, this would increase the net terrestrial sink by 50%. While deep tillage of agricultural soils created a past major source of CO<sub>2</sub> to the atmosphere, modern agronomic methods have begun to reverse losses of carbon from soils. No-till or minimal till soil cultivation have been facilitated by herbicide resistance traits and have resulted in some carbon accumulation [95,96]. How do we now retain substantially more carbon in the soil? Two opportunities in combination would allow this: modifying crop residues to be more recalcitrant to decomposition processes, so slowing return to the atmosphere, and larger and deeper root systems [97]. What is the opportunity and likelihood of this?

Increasing the amount of aerenchyma in roots, besides providing protection against flooding, also allows the development of more roots, including deeper roots, without requiring more investment in tissue mass. Typically, the water content of the soil increases with depth and the lower layers may be anaerobic. Aerenchyma will allow the root to function and gain water and

nutrients in these conditions [98], but on death, this root biomass will decompose far more slowly under anaerobic conditions than in the higher, more aerobic soil layers [99]. Deep root remains are also well below the depth of tillage [99]. Considerable genetic variation exists in the size, depth and architecture of root systems that could be exploited to increase both the efficiency of resource capture and soil carbon sequestration [84,100–102]. In parallel, factors controlling root architecture, rooting depth and root mass are being revealed at the molecular level [103–107].

A massive effort has been placed into understanding what makes plant cell walls difficult to deconstruct and how to bioengineer walls that can be more easily deconstructed for the efficient production of cellulosic biofuels [108–111]. This has also taught us how to make cell walls more recalcitrant so that crop residues will form a more effective soil carbon sink. From this prior work, it can be deduced that increased lignin content increases the recalcitrance of cell walls. Lignin is a random cross-linked polymer of three major monomers referred to as p-hydroxyphenyl (H), guaiacyl (G) and syringyl (S). Enrichment of the H content is shown to decrease polymerization and in turn recalcitrance in *Arabidopsis*. Conversely, lowering the H content should increase recalcitrance. Hydrophobic components of cell walls and surfaces are particularly recalcitrant. Plant cuticles consist primarily of cutin C32–36 polymers of C16 and C18 esterified and oxygenated fatty acids with small amounts of phenyl-propanoids [112]. They are by far the longest persisting components of plant remains in soils, typically being the only fossil remnants surviving thousands and millions of years [113]. Suberin is similarly hydrophobic, consisting of dimethyl esters of epoxyoctadecanedioic acids, with different levels of hydroxylation [114]. Decomposition of both cutin and suberin in soils is greatly slowed when complexed with lignin [115]. These findings suggest that to preserve a much higher proportion of root and crop residue in the soil would best be achieved by increasing the proportion of cutin and suberin, together with increased lignin of lower H content.

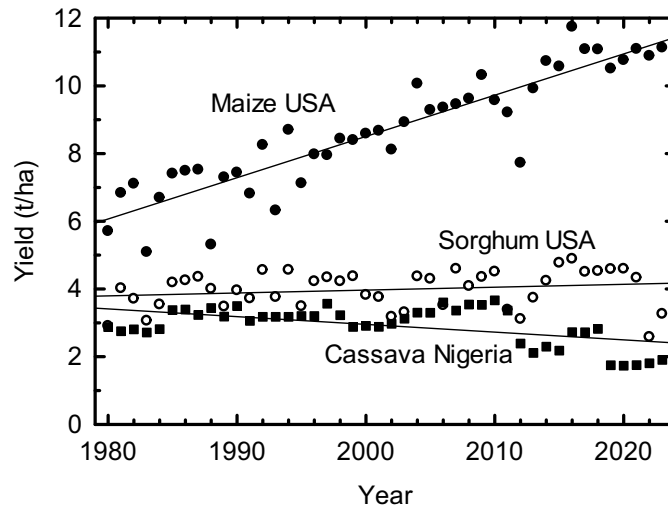
## 8. Perennial C<sub>4</sub> grasses for atmospheric CO<sub>2</sub> capture

Afforestation has been widely viewed as a means to offset carbon emissions. Forests, in their aggradation stage, may sequester about 3.2 t [CO<sub>2</sub>] per hectare into new biomass per year [116]. A mature crop of the perennial C<sub>4</sub> grass, *Miscanthus × giganteus* (*Miscanthus*), grown in Illinois, sequestered 130 t [CO<sub>2</sub>] per hectare in one year [117]. As productive as this may seem, plant breeders have identified substantially more productive *Miscanthus* genotypes [118]. *Miscanthus*, in common with other highly productive C<sub>4</sub> perennial grasses, such as switchgrass (*Panicum virgatum*) and prairie cordgrass (*Spartina pectinata*), produce an annual crop of shoots from perennating rhizomes [119,120]. In the autumn, the shoots senesce and translocate nutrients to the rhizome, which is then used by next year's crop of shoots the following spring [121]. The stems are harvested on completion of dry-down in the autumn. This makes these crops highly sustainable and able to produce biomass with very few or no inputs following establishment; further, they add large amounts of organic matter to the soil through the turnover of their large root and rhizome systems, which also serve to bind the soil [117,121]. The perenniality and sustainability of these crops make them particularly suitable to lands that are marginal for arable production because of their susceptibility to erosion or because soil quality is insufficient for the economic production of food crops. The performance of the three perennial grasses *Miscanthus*, switchgrass and energycane (*Saccharum officinarum* L. hybrids - fibre crops derived from sugarcane breeding) was assessed for marginal land capable of producing a crop under rain-fed conditions in the eastern USA. Yields were predicted to high spatial resolution based on gridded soil data and down-scaled current and future climate at each location. Selecting the most productive of these three crops for each location, it was estimated that these crops on this marginal land would remove 600 million tonnes of CO<sub>2</sub> from the atmosphere into harvestable shoot biomass per annum [118]. Much of the geology of the eastern USA is suitable for deep CO<sub>2</sub> storage. Using BECCS (bioenergy with carbon capture and storage), this biomass could be burnt to generate electricity and the resulting CO<sub>2</sub> captured and transferred to deep underground storage [118]. This CO<sub>2</sub> removal statistic does not take account of the additional benefits of displacement of fossil fuel use in electricity generation or the carbon that will accumulate in the soil under these crops. Such large-scale plantings would alter surface albedo and latent heat transfer to the atmosphere sufficient to cause a regional summer cooling of 1°C, which would significantly offset regional climate change-induced warming [122]. There are also key opportunities to similarly utilize somewhat less productive but drought tolerant desert perennials on degraded hot semi-arid lands [123–126].

## 9. The key role of plant breeding

The above sections have outlined many potential opportunities for adapting crops to future global change and even using crops to remove CO<sub>2</sub> from the atmosphere. Many of these opportunities require incorporating transgenes or introgression of advantageous alleles from crop relatives. While the acceptability of transgenic crops is slowly rising, particularly where it is most needed, the pathway to deregulation, where allowed, is still a long and expensive one [127,128]. Two developments, however, could greatly accelerate this process. In many cases, transgenic improvements involve upregulation by adding extra copies of a gene that the crop already has. This can now likely be achieved by editing the upstream non-coding region of the gene to, for example, remove repressor elements [35,129]. Such scarless editing is increasingly being accepted as non-transgenic [130]. Rapid progress in functional simulation of key proteins is allowing *in silico* testing of multiple amino acid residues to predict, for example, more efficient forms of key enzymes [131]. Again, these changes could be achieved by scarless editing, avoiding the need for transgenic upregulation.

Key to achieving any of these opportunities are the roles of plant breeders and seed systems. These changes can only be implemented by introgression of these into regionally adapted and accepted cultivars and identifying the genetic backgrounds



**Figure 1.** Average yields of maize and sorghum in the USA and cassava in Nigeria. Cassava is given as dry mass, assuming it has a moisture content of 70%. Data from [137].

in which the transgene, edit or allele from a crop relative is most effective [132]. Unfortunately, introducing just the opportunities outlined in the previous sections appears far beyond the current capacity for plant breeding. Just as plant molecular biological opportunities have mushroomed, the capacity of public domain plant breeding to implement these innovations has declined dramatically. The past four decades have seen a sharp decline in public plant breeding, offset to some extent by the ascendancy of a few multinational companies with vast plant breeding capability [133,134]. Unfortunately, most of this multinational effort must concentrate on short-term financial gain, which has been best served by a focus on North American hybrid maize improvement [135]. However, nearly 80% of maize produced in the USA is used for ethanol production or as animal feed [136], thereby contributing relatively little to global food security. Between 1980 and 2024, US maize yields doubled while sorghum improved by just 12% (figure 1). Maize improvement has largely resulted from massive investment by multinationals, while sorghum improvement has largely depended on depleted public domain efforts. Cassava, a major staple of much of Africa, showed no improvement in its average yield in Nigeria, the largest producer [137]. This is despite progress in breeding to counter increasing losses owing to pests, diseases and drought (figure 1) [138,139]. The example of maize in the USA shows what could be achieved with other crops in the public domain with similar investment. Although now aided with high-throughput phenotyping tools, marker-assisted and genomic breeding, it is hard to see how the opportunities for future-proofing our crops that are highlighted in this review and in accompanying articles can be implemented at the scale that is necessary. The discovery of the rice *Sub1A-1* allele, its introgression into a wide range of rice cultivars and subsequent availability through seed systems provide a shining example of what can be achieved in the public domain for some of the world's poorest farmers [89]. A second emerging example of such success through collaboration is the approval of cowpea carrying the *Bacillus thuringiensis* (Bt) *cry1Ab* gene in Nigeria and now Ghana. This was the first transgenic food crop approved for distribution to farmers in Africa, excepting South Africa's acceptance of bt maize. Its adoption by farmers prevents the devastating losses caused by the bean pod borer, *Maruca vitrata*, which can cost up to 90% of this key staple for the region [128,140]. These examples show the success that can be achieved when molecular biology, breeding and seed distribution systems work together for a common cause. But many more such examples will be needed if we are to avert famines on a scale not seen for decades. Achieving this requires the training of more plant breeders in both national and international programmes and access to relevant high-throughput phenotyping and genotyping facilities, especially in the countries that will continue to be most affected by food shortages. It also requires much swifter and more efficient deregulation of key biotechnologies. While there is still time, these needs deserve our most urgent attention.

'Some see things as they are, and ask why. I dream of things that never were, and ask why not?' Robert F. Kennedy

**Ethics.** This work did not require ethical approval from a human subject or animal welfare committee.

**Data accessibility.** This article has no additional data.

**Declaration of AI use.** I have not used AI-assisted technologies in creating this article.

**Authors' contributions.** S.P.L.: conceptualization, writing—original draft, writing—review and editing.

**Conflict of interest declaration.** I declare I have no competing interests.

**Funding.** S.P.L. is supported by awards from Gates Agricultural Innovations grant investment ID 57248, the DOE Center for Advanced Bioenergy and Bioproducts Innovation (US Department of Energy, Office of Science, Biological and Environmental Research Program) under award number DE-SC0018420 and the Carbon Technology Research Foundation under award number 00850818. Any opinions, findings, and conclusions or recommendations expressed in this publication are those of the author and do not necessarily reflect the views of Gates Agricultural Innovations, the US Department of Energy or the Carbon Technology Research Foundation.

**Acknowledgements.** Ann Long is thanked for critical comments and advice on drafts.

## References

1. USDS. 2023 *Secretary Antony J. Blinken on the vision for adapted crops and soils: global solutions for food security event*. <https://2021-2025.state.gov/secretary-blinken-on-the-vision-for-adapted-crops-and-soils-at-the-united-nations-security-council>: U.S. Department of State.
2. d'Amour CB, Wenz L, Kalkuhl M, Christoph Steckel J, Creutzig F. 2016 Teleconnected food supply shocks. *Environ. Res. Lett.* **11**, 035007. (doi:10.1088/1748-9326/11/3/035007)
3. Schilling J, Hertig E, Trambly Y, Scheffran J. 2020 Climate change vulnerability, water resources and social implications in North Africa. *Reg. Environ. Chang* **20**, 1–12. (doi:10.1007/s10113-020-01597-7)
4. Soffiantini G. 2020 Food insecurity and political instability during the Arab Spring. *Glob. Food Secur.* **26**, 100400. (doi:10.1016/j.gfs.2020.100400)
5. Weinberg J, Bakker R. 2015 Let them eat cake: food prices, domestic policy and social unrest. *Confl. Manag. Peace Sci.* **32**, 309–326. (doi:10.1177/0738894214532411)
6. Bouton C. 2000 Les mouvements de subsistance et le problème de l'économie morale sous l'ancien régime et la Révolution Française. *Ann. Hist. Revolut. Fr.* **319**, 71–100. (doi:10.4000/ahrf.104)
7. Powderly WG. 2019 How infection shaped history: lessons from the Irish famine. *Trans. Am. Clin. Climatol. Assoc.* **130**, 127–135.
8. Taine HA. 2024 *The french revolution*. vol. 1. Whitefish, MT, USA: Kessinger Publishing.
9. NOAA. 2025 *During a year of extremes, carbon dioxide levels surge faster than ever*. Boulder, CO, USA: National Oceanic and Atmospheric Administration Global Monitoring Laboratory.
10. IPCC. 2021 *Climate change 2021: The Physical Science basis. Contribution of Working Group 1 to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge, UK: Cambridge University Press.
11. McCulloch MT, Winter A, Sherman CE, Trotter JA. 2024 300 years of sclerosponge thermometry shows global warming has exceeded 1.5 °C. *Nat. Clim. Chang.* **14**, 171–177. (doi:10.1038/s41558-023-01919-7)
12. Copernicus. 2024 *Copernicus: June 2024 marks 12th month of global temperature reaching 1.5°C above pre-industrial*. Reading, UK: European Centre for Medium-Range Weather Forecasts (ECMWF).
13. UNFCCC. 2016 *The Paris Agreement*. Paris, France: United Nations Framework Convention on Climate Change (UNFCCC).
14. Ort DR, Long SP. 2014 Limits on yields in the corn belt. *Science* **344**, 483–484. (doi:10.1126/science.1253884)
15. van Dijk M, Morley T, Rau ML, Saghai Y. 2021 A meta-analysis of projected global food demand and population at risk of hunger for the period 2010–2050. *Nat. Food* **2**, 494–501. (doi:10.1038/s43016-021-00322-9)
16. Salesse-Smith CE, Wang Y, Long SP. 2025 Increasing Rubisco as a simple means to enhance photosynthesis and productivity now without lowering nitrogen use efficiency. *New Phytol.* **245**, 951–965. (doi:10.1111/nph.20298)
17. Hasegawa T, Sakurai G, Fujimori S, Takahashi K, Hijioka Y, Masui T. 2021 Extreme climate events increase risk of global food insecurity and adaptation needs. *Nat. Food* **2**, 587–595. (doi:10.1038/s43016-021-00335-4)
18. Cottrell RS *et al.* 2019 Food production shocks across land and sea. *Nat. Sustain.* **2**, 130–137. (doi:10.1038/s41893-018-0210-1)
19. Fearnside PM, Figueiredo AMR, Bonjour SCM. 2013 Amazonian forest loss and the long reach of China's influence. *Environ. Dev. Sustain.* **15**, 325–338. (doi:10.1007/s10668-012-9412-2)
20. Lundberg C, Abman R. 2022 Maize price volatility and deforestation. *Am. J. Agric. Econ.* **104**, 693–716. (doi:10.1111/ajae.12246)
21. Meyfroidt P, Rudel TK, Lambin EF. 2010 Forest transitions, trade, and the global displacement of land use. *Proc. Natl Acad. Sci. USA* **107**, 20917–20922. (doi:10.1073/pnas.1014773107)
22. FAO. 2024 *Hunger and food insecurity*. Rome, Italy: Food and Agriculture Organization of the United Nations.
23. McLeod AR, Long SP. 1999 Free-air carbon dioxide enrichment (FACE) in global change research: a review. *Adv. Ecol. Res.* **28**, 1–56. (doi:10.1016/S0065-2504(08)60028-8)
24. Ainsworth EA, Long SP. 2020 30 years of free-air carbon dioxide enrichment (FACE): what have we learned about future crop productivity and its potential for adaptation? *Glob. Chang. Biol.* **26**, 27–49. (doi:10.1111/gcb.15375)
25. Gray SB *et al.* 2016 Intensifying drought eliminates the expected benefits of elevated carbon dioxide for soybean. *Nat. Plants* **2**, 16132. (doi:10.1038/nplants.2016.132)
26. Myers SS *et al.* 2014 Increasing CO<sub>2</sub> threatens human nutrition. *Nature* **510**, 139–142. (doi:10.1038/nature13179)
27. Clark PU, Shakun JD, Rosenthal Y, Köhler P, Bartlein PJ. 2024 Global and regional temperature change over the past 4.5 million years. *Science* **383**, 884–890. (doi:10.1126/science.ad1908)
28. Long SP, Ainsworth EA, Rogers A, Ort DR. 2004 Rising atmospheric carbon dioxide: plants FACE the future. *Annu. Rev. Plant Biol.* **55**, 591–628. (doi:10.1146/annurev.arplant.55.031903.141610)
29. Lefebvre S, Lawson T, Zakhleniuk OV, Lloyd JC, Raines CA, Fryer M. 2005 Increased sedoheptulose-1,7-bisphosphatase activity in transgenic tobacco plants stimulates photosynthesis and growth from an early stage in development. *Plant Physiol.* **138**, 451–460. (doi:10.1104/pp.104.055046)
30. Rosenthal DM, Locke AM, Khozaei M, Raines CA, Long SP, Ort DR. 2011 Over-expressing the C<sub>3</sub> photosynthesis cycle enzyme sedoheptulose-1-7 bisphosphatase improves photosynthetic carbon gain and yield under fully open air CO<sub>2</sub> fumigation (FACE). *BMC Plant Biol.* **11**, 123. (doi:10.1186/1471-2229-11-123)
31. Köhler IH, Ruiz-Vera UM, VanLoocke A, Thomey ML, Clemente T, Long SP, Ort DR, Bernacchi CJ. 2017 Expression of cyanobacterial FBP/SBPase in soybean prevents yield depression under future climate conditions. *J. Exp. Bot.* **68**, 715–726. (doi:10.1093/jxb/erw435)
32. Pignon CP, Long SP. 2020 Retrospective analysis of biochemical limitations to photosynthesis in 49 species: C<sub>4</sub> crops appear still adapted to pre-industrial atmospheric [CO<sub>2</sub>]. *Plant Cell Environ.* **43**, 2606–2622. (doi:10.1111/pce.13863)
33. Salesse-Smith CE, Sharwood RE, Busch FA, Kromdijk J, Bardal V, Stern DB. 2018 Overexpression of Rubisco subunits with RAF1 increases Rubisco content in maize. *Nat. Plants* **4**, 802–810. (doi:10.1038/s41477-018-0252-4)
34. Salesse-Smith CE *et al.* 2025 Adapting C<sub>4</sub> photosynthesis to atmospheric change and increasing productivity by elevating Rubisco content in sorghum and sugarcane. *Proc. Natl Acad. Sci. USA* **122**, e2419943122. (doi:10.1073/pnas.2419943122)
35. Patel-Tupper D, Kelikian A, Leipertz A, Maryn N, Tjahjadi M, Karavolias NG, Cho MJ, Niyogi KK. 2024 Multiplexed CRISPR-Cas9 mutagenesis of rice *PSBS1* noncoding sequences for transgene-free overexpression. *Sci. Adv.* **10**, eadm7452. (doi:10.1126/sciadv.adm7452)
36. Mills G, Buse A, Gimeno B, Bermejo V, Holland M, Emberson L, Pleijel H. 2007 A synthesis of AOT40-based response functions and critical levels of ozone for agricultural and horticultural crops. *Atmos. Environ.* **41**, 2630–2643. (doi:10.1016/j.atmosenv.2006.11.016)



37. Pavelin EG, Johnson CE, Rughooputh S, Touni R. 1999 Evaluation of pre-industrial surface ozone measurements made using Schönbein's method. *Atmos. Environ.* **33**, 919–929. (doi:10.1016/S1352-2310(98)00257-X)
38. Wang Y, Wild O, Ashworth K, Chen X, Wu Q, Qi Y, Wang Z. 2022 Reductions in crop yields across China from elevated ozone. *Environ. Pollut.* **292**, 118218. (doi:10.1016/j.envpol.2021.118218)
39. Van Dingenen R, Dentener FJ, Raes F, Krol MC, Emberson L, Cofala J. 2009 The global impact of ozone on agricultural crop yields under current and future air quality legislation. *Atmos. Environ.* **43**, 604–618. (doi:10.1016/j.atmosenv.2008.10.033)
40. McGrath JM, Betzelberger AM, Wang SW, Shook E, Zhu XG, Long SP, Ainsworth EA. 2015 An analysis of ozone damage to historical maize and soybean yields in the United States. *Proc. Natl Acad. Sci. USA* **112**, 14390–14395. (doi:10.1073/pnas.1509771112)
41. Li S, Montes CM, Aspray EK, Ainsworth EA. 2024 How do drought and heat affect the response of soybean seed yield to elevated O<sub>3</sub>? An analysis of 15 seasons of free-air O<sub>3</sub> concentration enrichment studies. *Glob. Chang. Biol.* **30**, e17500. (doi:10.1111/gcb.17500)
42. Long SP, Naidu SL. 2002 Effects of oxidants at the biochemical, cell and physiological levels, with particular reference to ozone. In *Air pollution and plant life* (eds JNB Bell, M Treshow), pp. 69–88, 2nd edn. West Sussex, UK: John Wiley & Sons, Ltd.
43. Ainsworth EA, Yendrek CR, Stith S, Collins WJ, Emberson LD. 2012 The effects of tropospheric ozone on net primary productivity and implications for climate change. *Annu. Rev. Plant Biol.* **63**, 637–661. (doi:10.1146/annurev-arplant-042110-103829)
44. Choquette NE *et al.* 2019 Uncovering hidden genetic variation in photosynthesis of field-grown maize under ozone pollution. *Glob. Chang. Biol.* **25**, 4327–4338. (doi:10.1111/gcb.14794)
45. Begum H *et al.* 2020 Genetic dissection of bread wheat diversity and identification of adaptive loci in response to elevated tropospheric ozone. *Plant Cell Environ.* **43**, 2650–2665. (doi:10.1111/pce.13864)
46. Ueda Y, Frimpong F, Qi Y, Matthus E, Wu L, Höller S, Kraska T, Frei M. 2015 Genetic dissection of ozone tolerance in rice (*Oryza sativa* L.) by a genome-wide association study. *J. Exp. Bot.* **66**, 293–306. (doi:10.1093/jxb/eru419)
47. Grossiord C, Buckley TN, Cernusak LA, Novick KA, Poulter B, Siegwolf RTW, Sperry JS, McDowell NG. 2020 Plant responses to rising vapor pressure deficit. *New Phytol.* **226**, 1550–1566. (doi:10.1111/nph.16485)
48. Hatfield JL, Boote KJ, Kimball BA, Ziska LH, Izaurralde RC, Ort D, Thomson AM, Wolfe D. 2011 Climate impacts on agriculture: implications for crop production. *Agron. J.* **103**, 351–370. (doi:10.2134/agronj2010.0303)
49. Scafaro AP, Gallé A, Van Rie J, Carmo-Silva E, Salvucci ME, Atwell BJ. 2016 Heat tolerance in a wild *Oryza* species is attributed to maintenance of Rubisco activation by a thermally stable Rubisco activase ortholog. *New Phytol.* **211**, 899–911. (doi:10.1111/nph.13963)
50. Degen GE, Worrall D, Carmo-Silva E. 2020 An isoleucine residue acts as a thermal and regulatory switch in wheat Rubisco activase. *Plant J.* **103**, 742–751. (doi:10.1111/tpj.14766)
51. Amaral J, Lobo AKM, Carmo-Silva E. 2024 Regulation of Rubisco activity in crops. *New Phytol.* **241**, 35–51. (doi:10.1111/nph.19369)
52. Hu SS, Ding YF, Zhu C. 2020 Sensitivity and responses of chloroplasts to heat stress in plants. *Front. Plant Sci.* **11**, 375. (doi:10.3389/fpls.2020.00375)
53. Commuri PD, Jones RJ. 2001 High temperatures during endosperm cell division in maize: a genotypic comparison under *in vitro* and field conditions. *Crop Sci.* **41**, 1122–1130. (doi:10.2135/cropsci2001.4141122x)
54. Salem MA, Kakani VG, Koti S, Reddy KR. 2007 Pollen-based screening of soybean genotypes for high temperatures. *Crop Sci.* **47**, 219–231. (doi:10.2135/cropsci2006.07.0443)
55. Li F, Sayama T, Yokota Y, Hiraga S, Hashiguchi M, Tanaka H, Akashi R, Ishimoto M. 2024 Assessing genetic diversity and geographical differentiation in a global collection of wild soybean (*Glycine soja* Sieb. et Zucc.) and assigning a mini-core collection. *DNA Res.* **31**, dsae009. (doi:10.1093/dnares/dsae009)
56. Prasad PVV, Boote KJ, Allen LH, Sheehy JE, Thomas JMG. 2006 Species, ecotype and cultivar differences in spikelet fertility and harvest index of rice in response to high temperature stress. *Field Crop. Res.* **95**, 398–411. (doi:10.1016/j.fcr.2005.04.008)
57. Coast O, Murdoch AJ, Ellis RH, Hay FR, Jagadish KSV. 2016 Resilience of rice (*Oryza* spp.) pollen germination and tube growth to temperature stress. *Plant Cell Environ.* **39**, 26–37. (doi:10.1111/pce.12475)
58. Das S, Krishnan P, Nayak M, Ramakrishnan B. 2014 High temperature stress effects on pollens of rice (*Oryza sativa* L.) genotypes. *Environ. Exp. Bot.* **101**, 36–46. (doi:10.1016/j.envexpbot.2014.01.004)
59. Bender J, Hertstein U, Black CR. 1999 Growth and yield responses of spring wheat to increasing carbon dioxide, ozone and physiological stresses: a statistical analysis of 'ESPACE-wheat' results. *Eur. J. Agron.* **10**, 185–195. (doi:10.1016/S1161-0301(99)00009-X)
60. Fu JM, Bowden RL, Jagadish SVK, Prasad PVV. 2023 Genetic variation for terminal heat stress tolerance in winter wheat. *Front. Plant Sci.* **14**, 1132108. (doi:10.3389/fpls.2023.1132108)
61. Li YP, Ye W, Wang M, Yan XD. 2009 Climate change and drought: a risk assessment of crop-yield impacts. *Clim. Res.* **39**, 31–46. (doi:10.3354/cr00797)
62. Seki M *et al.* 2002 Monitoring the expression profiles of 7000 *Arabidopsis* genes under drought, cold and high-salinity stresses using a full-length cDNA microarray. *Plant J.* **31**, 279–292. (doi:10.1046/j.1365-3113x.2002.01359.x)
63. Ingram J, Bartels D. 1996 The molecular basis of dehydration tolerance in plants. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **47**, 377–403. (doi:10.1146/annurev.arplant.47.1.377)
64. Zhu JK. 2002 Salt and drought stress signal transduction in plants. *Annu. Rev. Plant Biol.* **53**, 247–273. (doi:10.1146/annurev.arplant.53.091401.143329)
65. Waltz E. 2014 Beating the heat. *Nat. Biotechnol.* **32**, 610–613. (doi:10.1038/nbt.2948)
66. Oyekunle M, Adamu RS, Ndou E, Beyene Y, Abdulmalik MM, Oikeh SO. 2023 Efficacy of drought-tolerant and insect-protected transgenic TELA® maize traits in Nigeria. *Transgenic Res.* **32**, 169–178. (doi:10.1007/s11248-023-00345-x)
67. Wang Y *et al.* 2009 Shoot-specific down-regulation of protein farnesyltransferase ( $\alpha$ -subunit) for yield protection against drought in canola. *Mol. Plant* **2**, 191–200. (doi:10.1093/mp/ssn088)
68. Ritchie H, Roser M. 2024 How much water do we use? How did it change over time? In *Our World in Data* (ed. H Ritchie). <https://ourworldindata.org/water-use-stress>: Global Change Data Lab.
69. FAO. 2020 *Sustainable Food and Agriculture*. Rome, Italy: Food and Agricultural Organization of the United Nations.
70. Glowacka K *et al.* 2018 *Photosystem II Subunit S* overexpression increases the efficiency of water use in a field-grown crop. *Nat. Commun.* **9**, 868. (doi:10.1038/s41467-018-03231-x)
71. Turc B, Sahay S, Haupt J, de Oliveira Santos T, Bai G, Glowacka K. 2024 Up-regulation of non-photochemical quenching improves water use efficiency and reduces whole-plant water consumption under drought in *Nicotiana tabacum*. *J. Exp. Bot.* **75**, 3959–3972. (doi:10.1093/jxb/erae113)
72. Gray JE. 2007 Plant development: three steps for stomata. *Curr. Biol.* **17**, R213–R215. (doi:10.1016/j.cub.2007.01.032)
73. Caine RS *et al.* 2019 Rice with reduced stomatal density conserves water and has improved drought tolerance under future climate conditions. *New Phytol.* **221**, 371–384. (doi:10.1111/nph.15344)

74. Dunn J, Hunt L, Afsharinafar M, Meselmani MA, Mitchell A, Howells R, Wallington E, Fleming AJ, Gray JE. 2019 Reduced stomatal density in bread wheat leads to increased water-use efficiency. *J. Exp. Bot.* **70**, 4737–4748. (doi:10.1093/jxb/erz248)
75. Ferguson JN *et al.* 2024 Reducing stomatal density by expression of a synthetic epidermal patterning factor increases leaf intrinsic water use efficiency and reduces plant water use in a C<sub>4</sub> crop. *J. Exp. Bot.* **75**, 6823–6836. (doi:10.1093/jxb/erae289)
76. Phetluan W *et al.* 2023 Candidate genes affecting stomatal density in rice (*Oryza sativa* L.) identified by genome-wide association. *Plant Sci.* **330**, 111624. (doi:10.1016/j.plantsci.2023.111624)
77. Xie J, Fernandes SB, Mayfield-Jones D, Erice G, Choi M, E Lipka A, Leakey ADB. 2021 Optical topometry and machine learning to rapidly phenotype stomatal patterning traits for maize QTL mapping. *Plant Physiol.* **187**, 1462–1480. (doi:10.1093/plphys/kiab299)
78. Long SP, Taylor SH, Burgess SJ, Carmo-Silva E, Lawson T, De Souza AP, Leonelli L, Wang Y. 2022 Into the shadows and back into sunlight: photosynthesis in fluctuating light. *Annu. Rev. Plant Biol.* **73**, 617–648. (doi:10.1146/annurev-arplant-070221-024745)
79. Lawson T, Blatt MR. 2014 Stomatal size, speed, and responsiveness impact on photosynthesis and water use efficiency. *Plant Physiol.* **164**, 1556–1570. (doi:10.1104/pp.114.237107)
80. Eyland D, van Wesemael J, Lawson T, Carpentier S. 2021 The impact of slow stomatal kinetics on photosynthesis and water use efficiency under fluctuating light. *Plant Physiol.* **186**, 998–1012. (doi:10.1093/plphys/kiab114)
81. Acevedo-Siaca LG, Coe R, Wang Y, Kromdijk J, Quick WP, Long SP. 2020 Variation in photosynthetic induction between rice accessions and its potential for improving productivity. *New Phytol.* **227**, 1097–1108. (doi:10.1111/nph.16454)
82. FAO. 2021 *The impact of disasters and crises on agriculture and food security: 2021*. Rome, Italy: Food and Agriculture Organization of the United Nations.
83. Li Y, Sun D, Xu K, Jin L, Peng R. 2021 Hydrogen sulfide enhances plant tolerance to waterlogging stress. *Plants* **10**, 1928. (doi:10.3390/plants10091928)
84. Burton AL, Brown KM, Lynch JP. 2013 Phenotypic diversity of root anatomical and architectural traits in *Zea* species. *Crop Sci.* **53**, 1042–1055. (doi:10.2135/cropsci2012.07.0440)
85. Renziehausen T, Chaudhury R, Hartman S, Mustroph A, Schmidt-Schippers RR. 2024 A mechanistic integration of hypoxia signaling with energy, redox, and hormonal cues. *Plant Physiol.* **197**, kiae596. (doi:10.1093/plphys/kiae596)
86. Sasidharan R, Hartman S, Liu Z, Martopawiro S, Sajeev N, van Veen H, Yeung E, Voesenek L. 2017 Signal dynamics and interactions during flooding stress. *Plant Physiol.* **176**, 1106–1117. (doi:10.1104/pp.17.01232)
87. Reynoso MA *et al.* 2019 Evolutionary flexibility in flooding response circuitry in angiosperms. *Science* **365**, 1291–1295. (doi:10.1126/science.aax8862)
88. Bailey-Serres J, Lee SC, Brinton E. 2012 Waterproofing crops: effective flooding survival strategies. *Plant Physiol.* **160**, 1698–1709. (doi:10.1104/pp.112.208173)
89. Emerick K, Ronald PC. 2019 *Sub1* rice: Engineering rice for climate change. *Cold Spring Harb. Perspect. Biol.* **11**, a034637. (doi:10.1101/cshperspect.a034637)
90. Mackill DJ, Ismail AM, Singh US, Labios RV, Paris TR. 2012 Chapter six - Development and rapid adoption of submergence-tolerant (*Sub1*) rice varieties. In *Advances in agronomy* (ed. DL Sparks), pp. 299–352. San Diego, CA: Academic Press. (doi:10.1016/B978-0-12-394276-0.00006-8)
91. Peláez-Vico MA, Zandalinas SI, Deviredy AR, Sinha R, Mittler R. 2024 Systemic stomatal responses in plants: coordinating development, stress, and pathogen defense under a changing climate. *Plant Cell Environ.* **47**, 1171–1184. (doi:10.1111/pce.14797)
92. Zandalinas SI, Casal J, Rouached H, Mittler R. 2024 Stress combination: from genes to ecosystems. *Plant J.* **117**, 1639–1641. (doi:10.1111/tpj.16681)
93. Zandalinas SI, Peláez-Vico MÁ, Sinha R, Pascual LS, Mittler R. 2024 The impact of multifactorial stress combination on plants, crops, and ecosystems: how should we prepare for what comes next? *Plant J.* **117**, 1800–1814. (doi:10.1111/tpj.16557)
94. UN-FAO. 2023 *FAOSTAT*. Rome, Italy: Food and Agriculture Organization of the United Nations.
95. Wang H, Wang S, Yu Q, Zhang Y, Wang R, Li J, Wang X. 2020 No tillage increases soil organic carbon storage and decreases carbon dioxide emission in the crop residue-returned farming system. *J. Environ. Manag.* **261**, 110261. (doi:10.1016/j.jenvman.2020.110261)
96. Bernacchi CJ, Hollinger SE, Meyers T. 2005 The conversion of the corn/soybean ecosystem to no-till agriculture may result in a carbon sink. *Glob. Chang. Biol.* **11**, 1867–1872. (doi:10.1111/j.1365-2486.2005.01050.x)
97. Eckardt NA *et al.* 2023 Climate change challenges, plant science solutions. *Plant Cell* **35**, 24–66. (doi:10.1093/plcell/koac303)
98. Saengwilai P, Nord EA, Chimungu JG, Brown KM, Lynch JP. 2014 Root cortical aerenchyma enhances nitrogen acquisition from low-nitrogen soils in Maize. *Plant Physiol.* **166**, 726–735. (doi:10.1104/pp.114.241711)
99. Kell DB. 2011 Breeding crop plants with deep roots: their role in sustainable carbon, nutrient and water sequestration. *Ann. Bot.* **108**, 407–418. (doi:10.1093/aob/mcr175)
100. Fonta JE, Vejchasarn P, Henry A, Lynch JP, Brown KM. 2022 Many paths to one goal: identifying integrated rice root phenotypes for diverse drought environments. *Front. Plant Sci.* **13**, 959629. (doi:10.3389/fpls.2022.959629)
101. Jochua CN, Strock CF, Lynch JP. 2020 Root phenotypic diversity in common bean reveals contrasting strategies for soil resource acquisition among gene pools and races. *Crop Sci.* **60**, 3261–3277. (doi:10.1002/csc2.20312)
102. Lynch JP, Strock CF, Schneider HM, Sidhu JS, Ajmera I, Galindo-Castañeda T, Klein SP, Hanlon MT. 2021 Root anatomy and soil resource capture. *Plant Soil* **466**, 21–63. (doi:10.1007/s11104-021-05010-y)
103. Uga Y *et al.* 2013 Control of root system architecture by *DEEPER ROOTING 1* increases rice yield under drought conditions. *Nat. Genet.* **45**, 1097–1102. (doi:10.1038/ng.2725)
104. Li YX *et al.* 2024 The OsEIL1–OsWOX11 transcription factor module controls rice crown root development in response to soil compaction. *Plant Cell* **36**, 2393–2409. (doi:10.1093/plcell/koae083)
105. Makhoul M *et al.* 2024 Novel *PHOTOPERIOD-1* gene variants associate with yield-related and root-angle traits in European bread wheat. *Theor. Appl. Genet.* **137**, 125. (doi:10.1007/s00122-024-04634-9)
106. Nirmalaruban R *et al.* 2024 Root traits: a key for breeding climate-smart wheat (*Triticum aestivum*). *Plant Breed.* (doi:10.1111/pbr.13248)
107. Zhou SY, Zi XJ, Rao DY, Liu K, Yang L, Shen P, Wu B, Zhou F. 2024 Jasmonic acid mediates maize (*Zea mays* L.) roots response to soil nitrogen heterogeneity. *J. Plant Biol.* **67**, 241–255. (doi:10.1007/s12374-024-09428-6)
108. Loqué D, Scheller HV, Pauly M. 2015 Engineering of plant cell walls for enhanced biofuel production. *Curr. Opin. Plant Biol.* **25**, 151–161. (doi:10.1016/j.pbi.2015.05.018)
109. Matthews ML, Wang JP, Sederoff R, Chiang VL, Williams CM. 2020 Modeling cross-regulatory influences on monolignol transcripts and proteins under single and combinatorial gene knockdowns in *Populus trichocarpa*. *PLOS Comput. Biol.* **16**, e1007197. (doi:10.1371/journal.pcbi.1007197)
110. Matthews ML, Wang JP, Sederoff R, Chiang VL, Williams CM. 2021 A multiscale model of lignin biosynthesis for predicting bioenergy traits in *Populus trichocarpa*. *Comput. Struct. Biotechnol. J.* **19**, 168–182. (doi:10.1016/j.csbj.2020.11.046)
111. Wang JP, Matthews ML, Naik PP, Williams CM, Ducoste JJ, Sederoff RR, Chiang VL. 2019 Flux modeling for monolignol biosynthesis. *Curr. Opin. Biotechnol.* **56**, 187–192. (doi:10.1016/j.copbio.2018.12.003)

112. Heredia A. 2003 Biophysical and biochemical characteristics of cutin, a plant barrier biopolymer. *Biochim Biophys Acta* **1620**, 1–7. (doi:10.1016/s0304-4165(02)00510-x)
113. Jones MK, Briggs DEG, Briggs DEG, Eglington G, Hagelberg E. 1999 Molecular taphonomy of animal and plant cuticles: selective preservation and diagenesis. *Phil. Trans. R. Soc. Lond. B* **354**, 7–17. (doi:10.1098/rstb.1999.0356)
114. Santos S, Cabral V, Graça J. 2013 Cork suberin molecular structure: stereochemistry of the C<sub>18</sub> epoxy and vic-diol ω-hydroxyacids and α,ω-diacids analyzed by NMR. *J. Agric. Food Chem.* **61**, 7038–7047. (doi:10.1021/jf400577k)
115. Angst G, Heinrich L, Kögel-Knabner I, Mueller CW. 2016 The fate of cutin and suberin of decaying leaves, needles and roots – inferences from the initial decomposition of bound fatty acids. *Org. Geochem.* **95**, 81–92. (doi:10.1016/j.orggeochem.2016.02.006)
116. Beadle CL, Long SP. 1985 Photosynthesis — is it limiting to biomass production? *Biomass* **8**, 119–168. (doi:10.1016/0144-4565(85)90022-8)
117. Dohleman FG, Heaton EA, Arundale RA, Long SP. 2012 Seasonal dynamics of above- and below-ground biomass and nitrogen partitioning in *Miscanthus x giganteus* and *Panicum virgatum* across three growing seasons. *Glob. Chang. Biol. Bioenergy* **4**, 534–544. (doi:10.1111/j.1757-1707.2011.01153.x)
118. Njuguna JN *et al.* 2023 Biomass yield in a genetically diverse *Miscanthus sacchariflorus* germplasm panel phenotyped at five locations in Asia, North America, and Europe. *Glob. Chang. Biol. Bioenergy* **15**, 642–662. (doi:10.1111/gcbb.13043)
119. Heaton EA, Dohleman FG, Long SP. 2008 Meeting US biofuel goals with less land: the potential of *Miscanthus*. *Glob. Chang. Biol.* **14**, 2000–2014. (doi:10.1111/j.1365-2486.2008.01662.x)
120. Boe A, Owens V, Gonzalez-Hernandez J, Stein J, Lee DK, Koo BC. 2009 Morphology and biomass production of prairie cordgrass on marginal lands. *Glob. Chang. Biol. Bioenergy* **1**, 240–250. (doi:10.1111/j.1757-1707.2009.01018.x)
121. Heaton EA, Dohleman FG, Long SP. 2009 Seasonal nitrogen dynamics of *Miscanthus x giganteus* and *Panicum virgatum*. *Glob. Chang. Biol. Bioenergy* **1**, 297–307. (doi:10.1111/j.1757-1707.2009.01022.x)
122. He YF, Jaiswal D, Liang XZ, Sun C, Long SP. 2022 Perennial biomass crops on marginal land improve both regional climate and agricultural productivity. *Glob. Chang. Biol. Bioenergy* **14**, 558–571. (doi:10.1111/gcbb.12937)
123. Davis SC, Ortiz-Cano HG. 2023 Lessons from the history of *Agave*: ecological and cultural context for valuation of CAM. *Ann. Bot.* **132**, 819–833. (doi:10.1093/aob/mcad072)
124. Wang Y, Smith JAC, Zhu XG, Long SP. 2023 Rethinking the potential productivity of crassulacean acid metabolism by integrating metabolic dynamics with shoot architecture, using the example of *Agave tequilana*. *New Phytol.* **239**, 2180–2196. (doi:10.1111/nph.19128)
125. Buckland CE, Thomas DSG, Jägermeyr J, Müller C, Smith JAC. 2023 Drought-tolerant succulent plants as an alternative crop under future global warming scenarios in sub-Saharan Africa. *Glob. Chang. Biol. Bioenergy* **15**, 1287–1308. (doi:10.1111/gcbb.13095)
126. Neupane D, Mayer JA, Niechayev NA, Bishop CD, Cushman JC. 2021 Five-year field trial of the biomass productivity and water input response of cactus pear (*Opuntia* spp.) as a bioenergy feedstock for arid lands. *Glob. Chang. Biol. Bioenergy* **13**, 719–741. (doi:10.1111/gcbb.12805)
127. Ongu I, Olaiye P, Alexandersson E, Mugwanya Zawedde B, Eriksson D. 2023 Biosafety regulatory frameworks in Kenya, Nigeria, Uganda and Sweden and their potential impact on international R&D collaborations. *GM Crop. Food* **14**, 1–17. (doi:10.1080/21645698.2023.2194221)
128. USDA-FAS. 2024 *Biotechnology and other new production technologies*. United States Department of Agriculture - Foreign Agricultural Service.
129. Karavolias NG *et al.* 2023 Paralog editing tunes rice stomatal density to maintain photosynthesis and improve drought tolerance. *Plant Physiol.* **192**, 1168–1182. (doi:10.1093/plphys/kiad183)
130. Strobbe S, Wesana J, Van Der Straeten D, De Steur H. 2023 Public acceptance and stakeholder views of gene edited foods: a global overview. *Trends Biotechnol.* **41**, 736–740. (doi:10.1016/j.tibtech.2022.12.011)
131. Shukla D, Chen JM. 2022 Integration of machine learning with computational structural biology of plants. *Biochem. J.* **479**, 921–928. (doi:10.1042/bcj20200942)
132. Kantar MB *et al.* 2019 The many-faced Janus of plant breeding. *Plants People Planet* **1**, 306–309. (doi:10.1002/ppp3.30)
133. Brush SB. 2009 The demise of public plant breeding. *BioScience* **59**, 441–443. (doi:10.1525/bio.2009.59.5.13)
134. Coe MT, Evans KM, Gasic K, Main D. 2020 Plant breeding capacity in U.S. public institutions. *Crop Sci.* **60**, 2373–2385. (doi:10.1002/csc2.20227)
135. USDA-ERS. 2025 *Farm sector income & finances - farm sector income forecast*. Washington, DC: United State Department of Agriculture - Economic Research Service.
136. Saavoss M, Capehart T, McBride W, Effland A. 2021 *Trends in production practices and costs of the US corn sector*, p. 35. Washington, DC, USA: United States Department of Agriculture- Economic Research Service.
137. FAOstat. 2025 *FAOstat*. Rome, Italy: Food and Agricultural Organization of the United Nations.
138. De Souza AP, Massenburg LN, Jaiswal D, Cheng S, Shekar R, Long SP. 2017 Rooting for cassava: insights into photosynthesis and associated physiology as a route to improve yield potential. *New Phytol.* **213**, 50–65. (doi:10.1111/nph.14250)
139. Ntui VO, Tripathi JN, Kariuki SM, Tripathi L. 2024 Cassava molecular genetics and genomics for enhanced resistance to diseases and pests. *Mol. Plant Pathol.* **25**, e13402. (doi:10.1111/mpp.13402)
140. Nboyine JA *et al.* 2024 Field performance of a genetically modified cowpea (*Vigna unguiculata*) expressing the Cry1Ab insecticidal protein against the legume pod borer *Maruca vitrata*. *Agronomy* **14**, 3055. (doi:10.3390/agronomy14123055)